ARTICLE IN PRESS – J. Appl. Cryst.



JOURNAL OF APPLIED CRYSTALLOGRAPHY Tomographic X-ray scattering based on invariant reconstruction – analysis of the 3D nanostructure of bovine bone

ISSN 1600-5767

Proof instructions

Proof corrections should be returned by **23 February 2021**. After this period, the Editors reserve the right to publish your article with only the Managing Editor's corrections.

Please

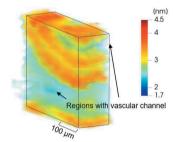
(1) Read these proofs and assess whether any corrections are necessary.

(2) Check that any technical editing queries highlighted in **bold underlined** text have been answered.

(3) Send corrections by e-mail to **ls@iucr.org**. Please describe corrections using plain text, where possible, giving the line numbers indicated in the proof. Please do not make corrections to the pdf file electronically and please do not return the pdf file. If no corrections are required please let us know.

Your article appears to qualify for open-access publication at no charge under a transformative deal with your institution. To purchase printed offprints, please complete the attached order form and return it by e-mail.

Please check the following details for your article



Thumbnail image for contents page

Synopsis: <u>Please provide a synopsis (of not more than two sentences) to appear in the Contents listing of the journal.</u>

Abbreviated author list: De Falco, P.; Weinkamer, R.; Wagermaier, W.; Li, C.; Snow, T. (1000000001-7146-6885); Terrill, N.J. (100000-0002-8783-1282); Gupta, H.S.; Goyal, P.; Stoll, M.; Benner, P.; Fratzl, P.

Keywords: SAXS; tomography; bovine bone; fibrolamellar unit; T parameter; small-angle X-ray scattering; scattering tomography; fibrolamellar bone

Open-access: Your article appears to qualify for open-access publication at no charge under a transformative deal with your institution.

How to cite your article in press Your article has not yet been assigned page numbers, but may be cited using the doi:

De Falco, P., Weinkamer, R., Wagermaier, W., Li, C., Snow, T., Terrill, N.J., Gupta, H.S., Goyal, P., Stoll, M., Benner, P. et al. (2021). J. Appl. Cryst. 54, https://doi.org/10.1107/S1600576721000881.

You will be sent the full citation when your article is published and also given instructions on how to download an electronic reprint of your article.

J. Appl. Cryst. (2021). 54

https://doi.org/10.1107/S1600576721000881

Files: j/vg5128/vg5128.3d j/vg5128/vg5128.sgml VG5128 FA IU-2112/57(16)2 2112/56(16)2 ()

Tomographic X-ray scattering based on invariant reconstruction - analysis of the 3D nanostructure

Paolino De Falco,^a Richard Weinkamer,^a* Wolfgang Wagermaier,^a Chenghao Li,^a Tim Snow,^b Nicholas J. Terrill,^b Himadri S. Gupta,^c Pawan Goyal,^d Martin Stoll,^{d,e} Peter Benner^d and Peter Fratzl^a*

^aDepartment of Biomaterials, Max Planck Institute of Colloids and Interfaces, Potsdam 14476, Germany, ^bDiamond Light Source Ltd. Diamond House Harwell Science and Innovation Campus. Didcot. Oxfordshire OX11.0DF. United Kingdom, ^cSchool of Engineering and Materials Science, Queen Mary University of London, London E1 4NS, United Kingdom, ^dMax Planck Institute for Dynamics of Complex Technical Systems, Sandtorstrasse 1, Magdeburg 39106, Germany, and ^eDepartment of Mathematics, TU Che nitz, Reichenhainer Strasse 41, Chemnitz 09126, Germany. *Correspondence e-mail: richard.weinkamer@mpikg.mpg.de, fratzl@mpikg.mpg.de

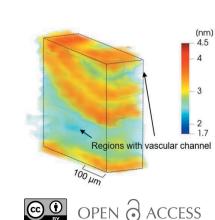
Small-angle X-ray scattering (SAXS) is an effective characterization technique for multi-phase nanocomposites. The structural complexity and heterogeneity of biological materials require the development of new techniques for the 3D characterization of their hierarchical structures. Emerging SAXS tomographic methods allow reconstruction of the 3D scattering pattern in each voxel but are costly in terms of synchrotron measurement time and computer time. To address this problem, an approach has been developed based on the reconstruction of SAXS invariants to allow for fast 3D characterization of nanostructured inhomogeneous materials. SAXS invariants are scalars replacing the 3D scattering patterns in each voxel, thus simplifying the 6D reconstruction problem to several 3D ones. Standard procedures for tomographic reconstruction can be directly adapted for this problem. The procedure is demonstrated by determining the distribution of the nanometric bone mineral particle thickness (T parameter) throughout a macroscopic 3D volume of bovine cortical bone. The T parameter maps display spatial patterns of particle thickness in fibrolamellar bone units. Spatial correlation between the mineral nanostructure and microscopic features reveals that the mineral particles are particularly thin in the vicinity of vascular channels.

1. Introduction

of bovine bone

Many biological materials incorporate nanoscopic mineral particles into an organic matrix, thereby reconciling conflicting material properties like strength and toughness (Weinkamer & Fratzl, 2016; Ritchie, 2011; Meyers et al., 2008). From both a biomedical (Xi et al., 2018; Milovanovic et al., 2015) and bio-inspired materials (Bouville et al., 2014; Studart, 2013) perspective, an important example is bone, which is a nanocomposite of stiff inorganic apatite particles embedded in a softer collagenous matrix (Weiner & Wagner, 1998; Fratzl & Weinkamer, 2007). The mineral particles in bone have roughly the shape of thin and elongated platelets with a thickness of 2-5 nm.

The characteristics of the mineral particles not only influence the mechanical performance but also provide hints about changes in bone physiology (Pathi et al., 2011). Previous work showed that the aspect ratio and staggered arrangement of the mineral particles affect the mechanical properties of bone (Jäger & Fratzl, 2000; Xi et al., 2018; Bar-On & Wagner, 2013). In general, the thickness of the mineral particles can be viewed



J. Appl. Cryst. (2021). 54

CRYSTALLOGRAPHY

JOURNAL OF

Received 8 September 2020 Accepted 25 January 2021

Edited by D. J. Svergun, European Molecular Biology Laboratory, Hamburg, Germany

as an indication of tissue age and normally correlates with the
degree of mineralization (Zizak *et al.*, 2003; Roschger *et al.*,
2001; Fratzl *et al.*, 1991), except in the case of *osteogenesis imperfecta*, the brittle bone disease (Fratzl-Zelman *et al.*,
2014). He *et al.* (2017) found that regions affected by cancer
metastasis in mouse models contain thinner and less oriented
mineral particles compared with healthy bone.

The high contrast in electron density between the inorganic 122 and organic components in bone makes scattering techniques 123 an attractive approach to characterize the mineral particles 124 (Rinnerthaler et al., 1999; Pabisch et al., 2013). A particularly 125 powerful approach is 2D scanning small-angle X-ray scat-126 tering (SAXS), where an X-ray beam is used to scan the 127 sample and provides maps of the local mineral nanostructure 128 with a spatial resolution of several micrometres or even less (Pabisch et al., 2013; Paris et al., 2000). The data obtained in 130 this way are 4D, with two real-space dimensions corresponding 131 132 to the mapping by scanning of the X-ray beam, and another two from the 2D SAXS patterns that correspond to planar 133 sections through reciprocal space. 134

For a number of research questions a higher-dimensional 135 mapping of mineral characteristics would be desirable. A way 136 of increasing the dimensionality of the information is to collect 3D scattering patterns using a thin sample, but measuring the 138 scattering signal under different angles by rotating the sample. 139 Here the data are quasi-2D in real space but with three 140 dimensions in reciprocal space. Combined SAXS and wide-141 angle X-ray scattering (WAXS) were used to investigate the 142 crystalline and morphological texture of mineral particles in 143 human vertebrae, showing a close relationship between the 144 c-axis orientation and the orientation distribution of the 145 mineral platelets, the plate normal being perpendicular to the 146 c axis (Jaschouz et al., 2003). In that study, it was shown that 147 148 the mineral platelets are aligned with the collagen fibers along the trabecula axis. In synchrotron scanning SAXS/WAXS with 149 a beam size of 1 µm, it was demonstrated that mineral plate-150 lets in human osteonal bone change their orientation over a 151 length scale of approximately the thickness of a lamella of \sim 5– 152 10 µm (Seidel et al., 2012; Wagermaier et al., 2006), in agree-153 ment with the previously proposed rotated twisted plywood 154 structure (Weiner et al., 1999). This technique can be 155 combined with serial sectioning and scanning the slices under 156 various angles. The result of such an experiment was a full 6D 157 data set with a 3D map of 3D SAXS patterns for a human 158 trabecula (Georgiadis et al., 2016), where results about ultra-159 structural 3D orientation were confirmed using polarized light 160 microscopy (Georgiadis et al., 2015). 161

An alternative approach to serial sectioning is SAXS 162 163 tomography. Instead of reconstructing the attenuation coefficients as in standard microcomputed tomography, the aim here is to measure a bulk sample under different directions of 165 the beam and to reconstruct 3D SAXS patterns in a 3D 166 volume. Under the assumption of structural isotropy, recon-167 structions of the 1D SAXS pattern were performed for 169 samples like injected polymers (Schroer et al., 2006), nanoporous glass (Feldkamp et al., 2009) and rat brains (Jensen et 170 al., 2011). Only recently was the full reconstruction of the 171

spatially heterogeneous anisotropic ultrastructure in bone (Liebi *et al.*, 2015) and tooth (Schaff *et al.*, 2015) achieved.

In SAXS measurements the sample is rotated around many different rotational axes. The reconstruction of the obtained 6D data set is computationally intensive and becomes tractable by assuming certain symmetries in the data (tensor tomography) and by fitting the 3D pattern by spherical harmonics (Liebi *et al.*, 2015). Another approach also preserves oriented scattering information by the introduction of virtual scattering axes (Schaff *et al.*, 2015). The application of regularization strategies during reconstruction can save experimental data acquisition time (Liebi *et al.*, 2018) and stronger assumptions on the symmetry of the SAXS pattern can substantially reduce the time needed for reconstruction (Gao *et al.*, 2019).

However, in many instances the full oriented 3D SAXS pattern is not required in every voxel of the reconstructed volume. Instead, information derived from the SAXS patterns such as a Porod-like or Guinier-like analysis (Jensen *et al.*, 2011) is sufficient. Here we develop an approach to reconstruct directly the spatial variation of the particle thickness parameter instead of the spatial variation of full 3D-SAXS patterns. While the original data to be reconstructed consist of a 3D reciprocal-space picture in each real-space voxel, using invariants of the small-angle scattering replaces the 3D-SAXS data by scalars. This effectively reduces the problem to the reconstruction of a 3D matrix of scalars as in conventional X-ray absorption tomography and allows the use of efficient algorithms that have previously been developed for microcomputed tomography.

To test our approach, we chose bovine fibrolamellar bone as our model system, mainly for structural reasons: (i) the cortex

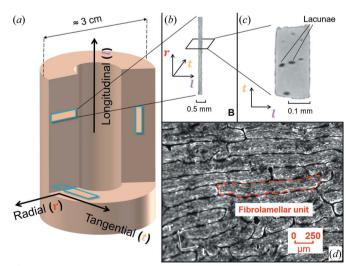


Figure 1

Macro- and microstructure of femoral bovine bone. (a) A representation of femoral bovine bone at the macroscale, showing the directions of the long axes (radial, tangential and longitudinal) of the measured samples. (b) A μ CT reconstruction of a sample with the long axis aligned to the radial direction of the femur, with an enlargement of a section shown in (c). (d) A light microscopy image showing the arrangement of fibrolamellar units at the microscale. The red dashed line outlines one such unit.

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

Files: j/vg5128/vg5128.3d j/vg5128/vg5128.sgml VG5128 FA IU-2112/57(16)2 2112/56(16)2 ()

Detecto

229 of a long bone is structurally anisotropic with a preferred direction along the long axis of the femur, (ii) the bone 230 231 consists of microscopic fibrolamellar units and these units are arranged in a regular brick-like fashion, with the shortest dimension oriented towards the bone center with a thickness 233 of approximately 200 µm (Fig. 1), and (iii) within the fibrolamellar unit structural features (different layers like the parallel-fibered layer and hypercalcified layer, or vascular 236 channels) are present and large enough to be resolved with 237 our beam size of 10 µm. The structural anisotropy of the 238 fibrolamellar bone is reflected in extremely anisotropic 239 mechanical properties, with the elastic modulus being 20 times 240 241 higher along the fiber direction than perpendicular to it (Seto et al., 2008). In the transition zone to the neighboring fibro-242 lamellar unit, lamellar bone is found, which again has a 243 preferred fiber orientation along the bone long axis and 244 contains blood vessels. Within both fibrolamellar and lamellar 245 246 bone a porous network is located. The lacunae of this lacunocanalicular network accommodate the cell bodies of 247 osteocytes, while their cell processes run in canaliculi. In 248 fibrolamellar bone of minipigs, the general orientation of the 249 canaliculi was found to be radial with tortuous and twisted pathways (Magal et al., 2014).

With the simplified reconstruction approach described in this paper, we were able to reconstruct spatial distributions of mineral particle characteristics in bovine fibrolamellar bone consisting of woven bone layers augmented by lamellar layers. A spatial correlation between the mineral nanostructure and microscopic features like vascular channels demonstrated that mineral particles are particularly thin in their vicinity.

2. Materials and methods

2.1. Samples

254

261

262

263

264

265

266

267 Four matchstick-like samples of bovine bone were used for both microcomputed tomography (μ CT) and synchrotron 268 scattering measurements (Fig. 1). All the samples were 269 obtained from the femur of a 23 month old cow, obtained from 270 a slaughterhouse. The mid-part of the diaphysis was cut into 271 approximately 2 cm thick pieces, the endosteal cancellous 272 273 bone was removed from the slice and the samples were stored at 251 K. Using a low-speed saw (Buehler Isomet, Düsseldorf, 274 Germany) under water cooling, plate-like samples were cut 275 under three different orientations, with the normal to the plate 277 pointing to the radial, longitudinal and tangential directions, respectively. Each plate was then polished to roughly 150 µm thickness. The plates were cut again to obtain stick-like 279 samples of approximately 4 mm in length [Fig. 1(b)] with a rectangular section of about $250 \times 150 \,\mu\text{m}$ [Fig. 1(c)]. The 281 long axes of the stick-like samples were aligned with one of the main directions of the femoral bovine bone [Fig. 1(a)] and are, therefore, referred to as radial (two samples), longitudinal 284 (one sample) and tangential (one sample). 285

Figure 2

(a)

The experimental setup adopted for the synchrotron measurements. (a) A schematic diagram showing SAXS signals acquired at different rotational angles φ . (b) A μ CT reconstruction of the sample with the rotational axis aligned to the radial direction shown in Fig. 1. In this sample, the normal to the main surface of the mineral particles is mainly parallel to the rotational axis of the sample. This allows particles to scatter the signal mainly along the vertical direction of the detector that we analyze by integrating the signal within an angular sector of $\Delta \chi = 6^{\circ}$.

(b)

Rotational axis (R II r

2.2. Micro-computed tomography (μ CT)

Rotational axis (R)

Grid

 μ CT measurements of all four samples were performed with the EasyTom 160 (RX solutions, Chavanod, France). In each measurement, the applied tube voltage was 60 kV and the integration times (duration of each tomographic projection) 11.0 s, resulting in a voxel size of 1.39 μ m³.

2.3. Synchrotron measurements

Two different synchrotron SAXS experiments were performed, on the μ Spot beamline at BESSY II (Germany) (Paris *et al.*, 2007) and on the I22 beamline at Diamond Light Source (DLS, UK) (Smith *et al.*, 2019). Throughout this paper, reported values of the experimental settings are separated by a forward slash (/), where the first value refers to BESSY II and the second to DLS. The monochromatic X-ray beam had an energy of 18 keV/14 keV and a beam size of 20 µm/10 µm, defined by a pinhole/secondary source slits. The sample-todetector distance was about 300 mm/5495 mm. Scattered signals were acquired by an EIGER X 9M/Pilatus P3-2M detector with an exposure time of 5 s/0.5 s.

In a scanning SAXS experiment the whole width of the sample was covered by the measurement grid, with the grid step defined by the step size between measurements. In both horizontal and vertical directions, the size of the grid step was equal to the beam size. The maximum number of horizontal scanned lines in the grid was 3/25. The same SAXS scan was repeated after rotating the sample along its long axis [Fig. 2(a)]. The measured set of angles θ ranged between 0° and 180° with an angular step of 3°/4°. Therefore, the number of measured SAXS patterns for each sample of bovine bone was 2745/45 000.

In addition to the scattering data, X-ray attenuation data were acquired using a diode with an exposure time of 0.3 s/ 0.5 s. The total time for collecting a data set at BESSY was about 45 h, while for the presented data set measured at DLS the total time amounted to around 8 h.

In the scattering experiments the rotational axis \mathcal{R} of the sample coincides with the long axis of our stick-like sample.

336

337

340

341

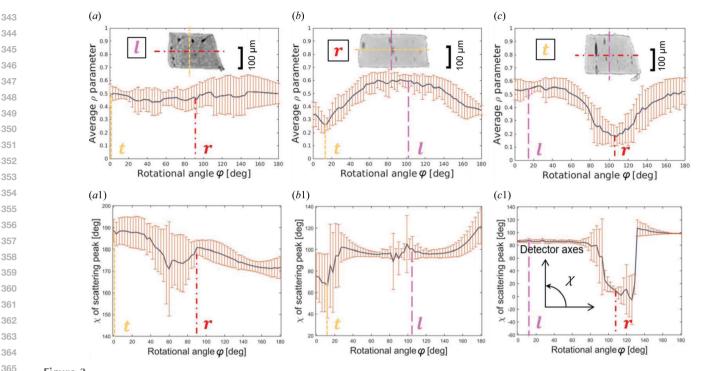


Figure 3

Macro- to nanostructure correlation. The plots show averaged values of the ρ parameter for the samples with the main axis aligned to the (a) longitudinal, (b) radial and (c) tangential directions of the samples. The values of the acquired ρ parameter at each rotational angle were averaged and the error bars indicate standard deviations. Dashed lines in each plot indicate the other two directions. (a1), (b1) and (c1) Plots of the position of the maximum of the scattering intensity (χ), where χ denotes the azimuthal angle, as a function of the rotational angle φ . [Labels are rather fuzzy and/or small - is a revised figure available?]

Samples were prepared in such a way that the longest extension is aligned with one of the main directions of the femoral bovine bone, *i.e.* the longitudinal, radial or tangential direction [Fig. 1(a)]. To link the rotational angle θ of the measurement with the position of the sample in the Cartesian coordinate system defined by the longitudinal, radial and tangential directions, sinograms of the attenuation coefficients were analyzed based on the known shape of sample cross sections. As an example, for the radial sample of Fig. 3(b) the tangential direction was determined to correspond to $\sim 12^{\circ}$.

3. SAXS-invariant tomography

3.1. SAXS invariants and platelet thickness

The SAXS intensity in 3D reciprocal space can be written as $I(q, \chi, \varphi)$, where q is the length of the scattering vector [q = $(4\pi/\lambda)\sin\theta$, where θ is half the scattering angle and λ is the wavelength of the incident radiation], χ the azimuthal angle measured with respect to an axis \mathcal{R} and φ the rotation angle around this axis \mathcal{R} . When the specimen is rotated around the axis \mathcal{R} defined by $\chi = -\pi/2$ or $\pi/2$, the intensity in this direction stays unchanged since the rotation axis remains fixed during the rotation. In the case of bone where there is a strong electron density contrast between the mineral and organic phases, a two-phase model can be used to describe the smallangle scattering and $I(q, \chi, \varphi)$ is taken proportional to the function $S(\mathbf{q})$ defined below. The proportionality constant

between the two functions will depend on instrumental parameters, as well as on the squared electron density difference between the organic and mineral phases. The vector \mathbf{q} is defined by its length q and the two angles χ and φ .

We consider a two-phase model with a function $\eta(\mathbf{r})$ that is equal to 1 if there is a particle at position r and is zero otherwise. Then the SAXS intensity will be proportional to

$$S(\mathbf{q}) = \frac{1}{V} \left| \int d\mathbf{r} \, \exp(-i\mathbf{q} \cdot \mathbf{r}) \, \eta(\mathbf{r}) \right|^2, \tag{1a}$$

$$= \frac{1}{V} \left| \int d\mathbf{r} \, \exp\left(-i\mathbf{q} \cdot \mathbf{r}\right) \left[\eta(\mathbf{r}) - \phi\right] \right|^2 \quad \left(\text{for } |\mathbf{q}| > 0 \right)$$
(1*b*)

$$= \frac{1}{V} \int d\mathbf{u} \, \exp\left(-i\mathbf{q} \cdot \mathbf{u}\right) \int d\mathbf{r} \left[\eta(\mathbf{r} + \mathbf{u}) - \phi\right] \left[\eta(\mathbf{r}) - \phi\right],\tag{1c}$$

where V is the probed volume and ϕ denotes the spatial average of η that just equals the volume fraction of particles in the volume V. In equation (1b), note that the subtraction of ϕ has no effect on $S(\mathbf{q})$ outside the origin of reciprocal space, q = 0. However, the average of $\eta(\mathbf{r})$ will formally generate a Dirac δ function at the origin that is practically invisible in SAXS. Therefore, the subtraction of the constant term ϕ ensures that the average of $[\eta(\mathbf{r}) - \phi]$ over the whole volume is zero and that there is no contribution of the Dirac δ function at q = 0 (which would otherwise contribute to the analytically

4 of 12 Paolino De Falco et al. • Tomographic X-ray scattering from bone

calculated integral intensity). This is the usual procedure in
the treatment of SAXS signals from two-phase systems, for
example in solution scattering, and the spherical average of
this expression is (Guinier & Fournet, 1955)

$$\overline{S}(q) = \langle S(\mathbf{q}) \rangle$$

$$= \int d\mathbf{u} \langle \exp(-i\mathbf{q} \cdot \mathbf{u}) \rangle \gamma(\mathbf{u})$$

$$= \int d\mathbf{u} \frac{\sin qu}{qu} \gamma(\mathbf{u})$$

$$= \int_{0}^{\infty} 4\pi u^{2} du \frac{\sin qu}{qu} \langle \gamma(\mathbf{u}) \rangle,$$

(2)

where the angle brackets denote the spherical average with respect to \mathbf{q} on the first two lines of the expression and with respect to \mathbf{u} on the last line. The first expression comes because only the exponential term depends on \mathbf{q} , and the second expression results from the averaging of the exponential term. In the last step, we rewrite the integration in spherical coordinates for the vector \mathbf{u} , and – since $(\sin qu)/qu$ does not depend on angle – we remain with a single integral with respect to u, provided we replace γ by its spherical average with γ denoting the correlation function,

$$\gamma(\mathbf{u}) = \frac{1}{V} \int d\mathbf{r} \left[\eta(\mathbf{r} + \mathbf{u}) - \phi \right] \left[\eta(\mathbf{r}) - \phi \right].$$
(3)

An inverse Fourier transform yields the expression for the spherically averaged correlation function (Guinier & Fournet, 1955):

$$\overline{\gamma}(u) = \gamma(\mathbf{u}) = \frac{1}{2\pi^2} \int_{0}^{\infty} q^2 \, \mathrm{d}q \, \frac{\sin qu}{qu} \, \overline{S}(q). \tag{4}$$

The consequence is the first SAXS invariant, the integral intensity

$$\int_{0}^{\infty} q^{2} \,\overline{S}(q) \,\mathrm{d}q = 2\pi^{2} \,\overline{\gamma}(0) = 2\pi^{2} \left(\phi - \phi^{2}\right) = 2\pi^{2} \,\phi \left(1 - \phi\right).$$
(5)

A Taylor expansion of the correlation function to the first order in u gives the second invariant, Porod's law (Guinier & Fournet, 1955), whereby S is the total amount of particle interface in the volume V:

$$\overline{\gamma}(u) = \phi(1-\phi) - \frac{S}{4V}u + \dots$$
(6)

The Fourier transform then gives the limit of the function $\overline{S}(q)$ for large q as

$$\overline{S}(q) \simeq \frac{S}{V} \frac{2\pi}{q^4} = \frac{P}{q^4}.$$
(7)

This has been used extensively [for a recent review, see Pabisch *et al.* (2013)] to determine an average thickness of particles through the parameter T defined as

J. Appl. Cryst. (2021). 54

$$T = \frac{4}{\pi P} \int_{0}^{0} q^2 \,\overline{S}(q) \,\mathrm{d}q = \frac{4\phi(1-\phi)}{S/V}.$$
 (8)

Here, ϕ and S/V are the volume fraction and the surface per unit volume of the particles, respectively. For thin particles with thickness *W*, this is well known to correspond to (Pabisch *et al.*, 2013)

 ∞

$$T \simeq 2(1 - \phi) W, \tag{9}$$

so that T roughly represents the mean particle thickness for a material with a particle volume fraction close to 50% (as in bone, for example).

The goal is now to generalize these expressions for integration along a rotation axis defined by the vector $\mathbf{q}_{\mathcal{R}} = (0, 0, q_{\mathcal{R}})$. We suppose that the specimen contains thin plates only. We first consider the contribution to the scattering by a single platelet oriented perpendicular to $\mathbf{q}_{\mathcal{R}}$ with a thickness W, and with breadth and length of B and L, respectively. Using equation (1a), the contribution of this particle to the total SAXS intensity can be written in Cartesian coordinates whereby z is along \mathcal{R} , yielding a well known result (Guinier & Fournet, 1955):

$$S_{1}(\mathbf{q}) = \frac{1}{V} \left| \int_{-L/2}^{L/2} \exp(-ixq_{x}) \, dx \int_{-B/2}^{B/2} \exp(-iyq_{y}) \, dy \right|$$
$$\times \left| \int_{-W/2}^{W/2} \exp(-izq_{z}) \, dz \right|^{2}$$

$$= \frac{1}{V} L^2 B^2 W^2 \left(\frac{\sin q_x L/2}{q_x L/2}\right)^2 \left(\frac{\sin q_y B/2}{q_y B/2}\right)^2$$
(sin q. W/2)²

$$\pm \left(\frac{\sin q_z W/2}{q_z W/2}\right). \tag{10}$$

Using the fact that the function $(L/2\pi)[(\sin q_x L/2)/(q_x L/2)]^2$ converges to the Dirac δ function when L gets very large, we obtain

X

$$S_1(\mathbf{q}) = 4\pi^2 \frac{L B W^2}{V} \left(\frac{\sin q_z W/2}{q_z W/2}\right)^2 \delta(q_x) \,\delta(q_y), \quad (11)$$

for L and B sufficiently large (in practical terms, larger than what SAXS would resolve in the relevant q range). Now, we call $N_{\mathcal{R}}$ the number of particles with their normal directions oriented within a small solid angle around \mathcal{R} and we denote $f_{\mathcal{R}}(W) dW$ the corresponding thickness distribution [normalized so that $f_{\mathcal{R}}(W) dW = 1$]. Then the total intensity pointing in the \mathcal{R} direction will be

$$\overline{S}_{\mathcal{R}}(q_{\mathcal{R}}) = N_{\mathcal{R}} \int f_{\mathcal{R}}(W) \, \mathrm{d}W \frac{1}{4\pi} \int_{0}^{2\pi} \mathrm{d}\varphi \int_{0}^{\pi} \mathrm{d}\chi \, \sin\chi \, S_{1}(\mathbf{q})$$
$$= \frac{\pi}{q_{\mathcal{R}}^{2}} \frac{N_{\mathcal{R}} \, L \, B}{V} \int W^{2} \left(\frac{\sin q_{\mathcal{R}} W/2}{q_{\mathcal{R}} W/2}\right)^{2} f_{\mathcal{R}}(W) \, \mathrm{d}W$$
$$(\text{for } |\mathbf{q}_{\mathcal{R}}| > 0) \tag{12}$$

581

586

587

590

591

594

596

598

600

601

602

603

604

605

606

607

608

609

610

611

613

614

615

616

617

618

619

571 The angles are defined such that $q_x = q \sin \chi \cos \varphi$, $q_{y} = q \sin \chi \sin \varphi$ and $q_{z} = q \cos \chi$, and we have neglected 572 573 possible interference effects between the particles. The volume fraction of all particles perpendicular to $\mathcal R$ is 574 $\phi_{\mathcal{R}} = N_{\mathcal{R}} L B \overline{W}_{\mathcal{R}} / V$, where $\overline{W}_{\mathcal{R}}$ is the average thickness of 575 the particles. It is also worth noting that the same family of 576 particles will also generate a similar scattering in the direction 577 related to \mathcal{R} by an inversion symmetry with respect to the origin (meaning in the $-\mathbf{q}_{\mathcal{R}}$ direction) but will not contribute 579 to the scattering in any other direction. 580

Equation (12) can be expanded at large $q_{\mathcal{R}}$ to give an analog to Porod's law,

$$\overline{S}_{\mathcal{R}}(q_{\mathcal{R}}) \simeq \frac{2\pi}{q_{\mathcal{R}}^4} \frac{N_{\mathcal{R}} L B}{V} = \frac{2\pi}{q_{\mathcal{R}}^4} \frac{\phi_{\mathcal{R}}}{\overline{W}_{\mathcal{R}}} = \frac{P_{\mathcal{R}}}{q_{\mathcal{R}}^4}.$$
(13)

Moreover, the integrated intensity along the \mathcal{R} direction (starting at the origin of q space) will be proportional to the total volume of particles perpendicular to this direction, that is, proportional to $\phi_{\mathcal{R}}$. The proportionality constant must be such that we recover equation (5) when summing over all possible directions. Here, we need to take into account that each plate-like particle scatters in two directions related by an inversion symmetry, so that we are counting each family of particles twice when we integrate over all directions (hence the factor 1/2 in the equation below) to get

$$\int_{0}^{\infty} q_{\mathcal{R}}^{2} \,\overline{S}(q_{\mathcal{R}}) \,\mathrm{d}q_{\mathcal{R}} = 2\pi^{2} \,\frac{\phi_{\mathcal{R}}}{2}(1-\phi). \tag{14}$$

Indeed, when we are summing this expression over all possible directions, and considering that $\phi_{\mathcal{R}}$ and $\phi_{-\mathcal{R}}$ refer to the same family of particles, $\phi_{\mathcal{R}}/2$ sums up to ϕ , recovering equation (5). Taking the ratio of the integral intensity above and the Porod constant in the \mathcal{R} direction, $P_{\mathcal{R}}$, we obtain in analogy to equation (9)

$$T_{\mathcal{R}} = \frac{4}{\pi P_{\mathcal{R}}} \int_{0}^{\infty} q_{\mathcal{R}}^2 \,\overline{S}(q_{\mathcal{R}}) \,\mathrm{d}q_{\mathcal{R}} \simeq 2(1-\phi)\overline{W}_{\mathcal{R}}.$$
 (15)

3.2. SAXS data evaluation

The acquired scattering intensity on the 2D detector, $I_a(q, \chi)$, was corrected considering X-ray attenuation and background subtraction according to

$$I(q,\chi) = \left[I_{\rm a}(q,\chi)\frac{I_0}{I_{\rm T}}\right] - I_{\rm BG}(q,\chi),\tag{16}$$

where I_0 is the intensity of the incoming beam, I_T the transmitted intensity and $I_{BG}(q, \chi)$ the background intensity. The angle χ is measured with respect to an axis \mathcal{R} . $I_{BG}(q, \chi)$ was obtained by averaging the scattering pattern of three measurement points of a scan which were outside the sample. Due to the fluctuation in the beam flux during the experiments at BESSY II, a further correction was applied to the measurements using background images for normalization.

In general, this intensity is not a voxel property, since scattering crucially depends on how the nanostructural elements, like the mineral particles in bone, are oriented with respect to the incoming beam. However, the scattering pattern does not change under sample rotation along the rotational axis, *i.e.* $I(q, \chi = -\pi/2)$ and $I(q, \chi = \pi/2)$ are independent of the specimen rotation around \mathcal{R} . Indeed, if the sample is rotated by an angle θ around \mathcal{R} , the reciprocal space is also rotated by the same angle around the same axis. An alternative explanation which considers the nanostructure of bone is that by focusing only on the scattered signal around the rotational axis \mathcal{R} , it is always the same subpopulation of mineral particles which contributes to the scattering signal. Taking into consideration the plate-like shape of the particles, this subpopulation consists of platelets with their normal oriented along the rotational axis [dark-gray particles in Fig. 2(b)].

For the calculation for the axial integrated intensity $I_{\mathcal{R}}$ and the axial Porod constant $P_{\mathcal{R}}$, an azimuthal integration of $I(q, \chi)$ in a very narrow sector around $\chi = -\pi/2, \pi/2$ with an opening angle of $\Delta \chi$ was performed. Our choice of $\Delta \chi = 6^{\circ}$ proved to be a good compromise to be, on the one hand, large enough to provide a robust value for $I_{\mathcal{R}}$ and, on the other hand, small enough to ensure that $I_{\mathcal{R}}$ is independent of the rotational angle φ . The q ranges in the evaluation were 0.2– 3 nm⁻¹/0.1–2.6 nm⁻¹.

Besides the reconstruction of the $T_{\mathcal{R}}$ parameter, a second parameter (ρ parameter), which describes the mutual alignment of the mineral particles (Pabisch *et al.*, 2013), was evaluated. To obtain the ρ parameter the SAXS patterns are radially integrated, yielding the scattered intensity as only a function of the azimuthal angle, $I(\chi)$. The function $I(\chi)$ displays two peaks separated by 180° on top of a constant background. The ρ parameter is defined as $\rho = A_1/(A_0 + A_1)$, where $A_0 + A_1$ denotes the area under the curve $I(\chi)$ including the constant background, and A_1 the area under the peaks only. Consequently, the ρ parameter takes values between 0 and 1, where 0 corresponds to a random mutual alignment and 1 to a perfect alignment of all the mineral particles.

All the SAXS data analysis was performed using the software packages *DAWN* (Basham *et al.*, 2015) and *DPDAK* (Benecke *et al.*, 2014). In order to visualize the ρ parameter as a function of the rotational angle (Fig. 3), the values of the ρ parameter acquired at a specific rotational angle were averaged.

3.3. Tomographic reconstructions

The essence of tomography is to reconstruct the bulk properties of a sample when only projection data are available. Typically, the different projections are obtained by scanning and rotating the sample. In the parallel-beam geometry applied in SAXS tomography experiments, reconstruction is a 2D problem defined by the slice of the sample which is scanned perpendicular to the rotational axis \mathcal{R} . In mathematical terms, projection data *p* are obtained at different scan-

628

629

630

631

632

633

634

635

636

637

638

Files: j/vg5128/vg5128.3d j/vg5128/vg5128.sgml VG5128 FA IU-2112/57(16)2 2112/56(16)2 ()

742

743

744

745

746

747

748

749

750

751

752

753

754

755

757

758

759

760

761

762

763

764

765

766

767

769

771

772

773

774

775

776

777

778

779

780

781

782

783

784

785

786

787

788

790

791

792

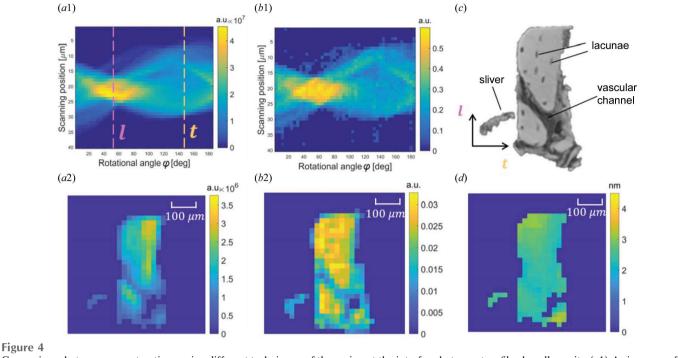
793

794

795

796

798



Comparisons between reconstructions, using different techniques, of the region at the interface between two fibrolamellar units. (*a*1) A sinogram of the integrated intensity achieved from SAXS measurements. Here the longitudinal and tangential directions are marked by dashed lines. (*b*1) A sinogram of the quantity $-\log(I_T/I_0)(r, \varphi)$. (*c*), (*a*2), (*b*2) Reconstructions from, respectively, the μ CT, SAXS and synchrotron CT measurements. (*d*) Reconstruction of the T_R parameter map. [Labels are rather fuzzy and/or small - is a revised figure available?]

ning positions r and different rotational angles φ and are usually plotted as a function of these two variables, $p(\varphi, r)$, as a sinogram [see *e.g.* Figs. 4(*a*1), 4(*b*1)]. From these input data a material property m(x, y, z) can be inferred, where x and y are coordinates in the coordinate system of the sample and z is assumed fixed, since we consider here only the reconstruction of a 2D slice of the sample.

The reconstruction problem is solved when two requirements are fulfilled:

(i) The material property is a scalar property of the 'voxel' representing a small material volume and, in particular, does not depend on the angle φ under which the voxel is measured. The X-ray attenuation coefficient, $\mu(x, y)$, is such a voxel property.

(ii) The material property is an additive quantity, *i.e.* $p(\theta, r) = \int_{\text{beam path}} m(x, y) \, ds. \ p(\varphi, r)$ is called the Radon transform of m(x, y). In the case of X-ray attenuation, the Beer-Lambert law ensures that the logarithm of the measured transmitted intensity is the Radon transform of the attenuation coefficient of the material.

For each synchrotron experiment, three reconstructions were performed. From the SAXS experiments, (i) the axial 733 integrated intensity $I_{\mathcal{R}}$ and (ii) the axial Porod constant $P_{\mathcal{R}}$ 734 were reconstructed (see Section 2.3). From the X-ray 735 attenuation experiments, (iii) the attenuation coefficient 736 $\mu(x, y, z)$ was reconstructed starting from measurements of 737 the transmitted intensity, specifically $-\log(I_T/I_0)(r,\varphi)$. For 739 the reconstruction, a filtered back-projection (FBP) algorithm was used (Thorsten, 2011) as implemented by the function 740 Iradon in MATLAB. Due to the virtually parallel-beam 741

geometry, the reconstruction could be performed in 'slices' of the sample, where the slice has a normal identical with the rotational axis \mathcal{R} and a thickness defined by the beam size.

To perform an FBP, the type of interpolation and high-pass filtering has to be specified, which accomplishes the mapping from the polar coordinate system of the sinogram to the Cartesian one after reconstruction. From the options offered by the Iradon function, a spline interpolation and a Ram-Lak filter yielded the most satisfying reconstruction results. Reconstruction quality was tested by projecting the result of reconstruction (i.e. by performing a Radon transform) and calculating the mean-squared error between these projected data and the original measurement data. To avoid spurious results for the T parameter $T_{\mathcal{R}}$ as a result of a division of two values close to zero, a threshold value for the axial integrated intensity I_R was introduced as 16% of the maximum value of $I_{\mathcal{R}}$ after reconstruction. All $I_{\mathcal{R}}$ values below this threshold were set to zero. Our choice of the threshold rendered the outer shape of the sample close to results from μCT measurements (Section 2.2). A sensitivity analysis showed that the exact value of the threshold has a negligible influence on the reported results.

The attenuation coefficients $\mu(x, y, z)$ were reconstructed after normalization of the data to correct for variations in the beam intensity. The data were first thresholded by setting all values of the sinogram data $(r, \varphi) = -\log(I_T/I_0)(r, \varphi)$, which are smaller than 6% of the maximum value equal to zero. For the normalization a factor was used which was obtained by averaging $s(r, \varphi)$ for a fixed rotational angle. The time for the reconstruction of each slice (*i.e.* fixed z coordinate) for inte-

685

687

690

691

604

697

698

700

701

702

704

705

706

708

709

710

711

719

720

721

722

723

724

725

Paolino De Falco et al. • Tomographic X-ray scattering from bone 7 of 12

grated intensity, Porod constant and attenuation coefficientwas below one second with a standard PC using FBP.

3.4. Spatial correlations

801

802

803

804

805

807

808

809

810

811

812

813

814

815

816

817

818

819

821

822

To allow a spatial correlation between the bone microstructure (vascular channels, osteocyte lacunae) and the mineral nanostructure, distance transforms were used. After binarizing the μ CT image and defining the voxels in the digital image that belonged to vascular channels/osteocyte lacunae, the distance transform assigns each bone voxel in the image the shortest distance value to the defined objects. Calculations were performed in MATLAB using the function *bwdistsc*. Image registration (using the MATLAB function *imregtform*) between the μ CT image and the 3D reconstruction of the attenuation coefficients from the synchrotron experiment was employed to map microstructure information (distance transforms) on nanostructure information (map of the *T* parameter).

4. Results

4.1. Correlation between mineral nanostructure and sample macrostructure

In a first step, we analyzed anisotropies of the mineral 823 nanostructure in relation to the macroscopic coordinate 824 system of the bovine femur defined by the longitudinal, radial and tangential directions [Fig. 1(a)]. The nanostructural 826 anisotropy was assessed by the ρ parameter (see *Methods* 827 section [p not actually mentioned until Section 3.2?]). 828 Analyzed sample slices (*i.e.* cross sections through the sample with a thickness of the beam size) had their normals (which 830 are identical to the rotational axis \mathcal{R}) in the direction of the 831 832 longitudinal, radial and tangential directions, respectively [Figs. 3(a), 3(b) and 3(c)]. For each rotational angle, a ρ 833 parameter was calculated as the average over all measurement 834 points in the slice [Figs. 3(a), 3(b) and 3(c)]. Different 835 dependencies as a function of the rotational angle are 836 837 observed: a rather constant curve for the longitudinal sample, while the radial and tangential samples each exhibit a 838 minimum. These minima correlate well in position with the 839 macroscopic directions, demonstrating that the preferred 840 orientations of the mineral nanostructure as quantified by the 841 ρ parameter align with the macroscopic coordinate system. 842 843 The majority of mineral particles are aligned with their long axis along the longitudinal direction. Fig. 3(a1) (longitudinal sample) shows that the scattering intensity $I(\chi)$ is a maximum 845 at an azimuthal angle χ of about 180°, independent of the 847 rotational angle φ , *i.e.* perpendicular to the rotation axis, and therefore $I_{\mathcal{R}}$ is low in the longitudinal sample. Radial samples are considered to exhibit the most effective scattering power 849 for our SAXS tomography approach, with a position of the maximally scattered intensity near $\chi = 90^{\circ}$ at most of the 851 rotational angles φ and, therefore, a large value for $I_{\mathcal{R}}$ 852 853 [Fig. 3(b2) [No such part?]]. The majority of particles in the radial sample are oriented such that their long axes are 854 perpendicular to the long axis of the sample and these parti-855

cles contribute to the scattering signal mainly along the direction of partial integration (vertical axis of the detector in our setup), as described in Section 2.3.

4.2. SAXS tomography and 3D T parameter map

The experimental setup allows one to obtain 3D maps by performing an independent reconstruction of sample slices (*i.e.* slices of a height equivalent to the beam size and oriented normal to the rotational axis \mathcal{R}) and piling these slices up after reconstruction to obtain the full 3D information. Fig. 4 shows representative data for a sample slice that includes a vascular channel. In the μ CT image of Fig. 4(c), not only can the vascular channel be clearly discerned, but osteocyte lacunae are also visible as small dark ellipsoids. A bone sliver close to the lower left corner of the sample, accidentally produced during cutting, additionally helped in the image registration of the different measurements.

Figs. 4(b1) and 4(a1) show $s(r, \varphi)$ from the absorption measurement and the projections of the axial integrated intensity from the scattering experiment as sinogram plots (*i.e.* as a function of the rotational angle φ and the scanning position r, see Section 2.3). The shape of the sample contributes substantially to the values in the sinogram, with the values being highest when the sample is viewed along its diagonal. The corresponding reconstructions for the attenuation coefficient $\mu(x, y)$ and the axial integrated intensity $I_{\mathcal{R}}(x, y)$ are shown in Figs. 4(b2) and 4(a2), respectively. A comparison between the μ CT image and the reconstructed attenuation coefficient shows that the synchrotron experiment allows a reliable reconstruction of the sample shape (including the sliver) and larger internal structures like vascular channels. However, the lacunae are too small to be visible.

Having reconstructed data not only for the axial integrated intensity $I_{\mathcal{R}}$ but also for the axial Porod constant allows the determination of the thickness $T_{\mathcal{R}}$ of mineral particles, which have their normal parallel to the rotational axis \mathcal{R} . The resulting map [Fig. 4(*d*)] shows spatial gradients in the particle thickness, with values above 3 nm in the upper left corner and values below 2 nm close to the vascular channel.

Repeating the evaluation for all measured sample slices and arranging the slices in the correct spatial order results in the 3D T_R parameter map shown in Fig. 5. Values of the T_R parameter vary between 1.7 and 4.5 nm, where regions close to the vascular channels display low values of T_R (region in light blue). The highest values of T_R were found within two band-like structures (regions in orange/reddish hue), with the thicknesses of the bands roughly 40 µm and a separation between them of about 15 µm.

4.3. Spatial correlation between micro- and nanostructure

The experimental design, with the same X-ray beam used to make both an absorption and a scattering experiment, allows a straightforward evaluation of spatial correlations between different structural quantities. In Fig. 6(c) the attenuation coefficient and the T_R parameter for identical voxels in the 3D reconstructions are plotted. While T_R reflects the nano-

Files: j/vg5128/vg5128.3d j/vg5128/vg5128.sgml VG5128 FA IU-2112/57(16)2 2112/56(16)2 ()

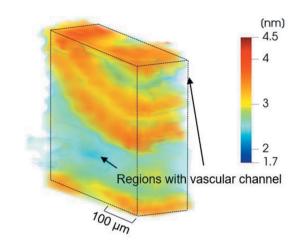


Figure 5

Structural characterization of the radial sample, showing a 3D reconstructed map of the $T_{\mathcal{R}}$ parameter for the sample with its main axis aligned to the radial direction of the bovine femur.

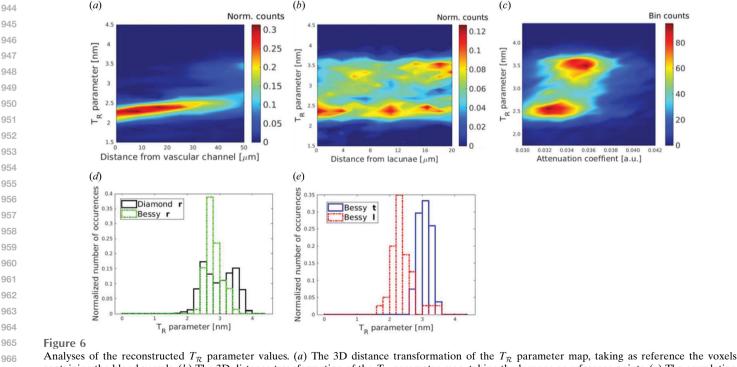
structural thickness of the mineral particles, the attenuation coefficient provides information about the local mineral content. The data points are concentrated in two regions of the plot: thin particles ($T_R \simeq 2.5$ nm) and low mineral content, and thicker particles ($T_{\mathcal{R}} \simeq 3.5 \text{ nm}$) and higher mineral content.

Next, the mineral nanostructure is correlated with microscopic features of the fibrolamellar bone. In Fig. 6(a) the $T_{\mathcal{R}}$ parameter is plotted as a function of distance from the vascular channel. Bone close to the vascular channel exhibits particularly low values of the $T_{\mathcal{R}}$ parameter. No clear spatial correlation can be detected when looking at the particle thickness as a function of the distance from the osteocyte lacunae [Fig. 6(b)].

In the frequency plots of Fig. 6(d) the normalized number of occurrences of the $T_{\mathcal{R}}$ parameter in both analyzed radial samples is represented. While data from the two samples with a smaller imaged volume show only one peak at relatively low values of the $T_{\mathcal{R}}$ parameter with a mean value of $T_{\mathcal{R}} = 2.81$ nm and a standard deviation (SD) of 0.23, the analogous plot of the other radial sample with the larger imaged volume exhibits two peaks of the T_R parameter, at 2.60 nm and 3.40 nm, as calculated with a Gaussian mixture model. Fig. 6(e) shows frequency plots for the longitudinal and tangential samples. For the longitudinal sample the obtained $T_{\mathcal{R}}$ parameter is relatively low (mean value 2.39 nm and SD 0.36) compared with the tangential sample (mean value 3.09 nm and SD 0.19).

5. Discussion and conclusions

In this proof-of-concept study on a mineralized tissue, we have demonstrated that 3D nanostructural information about mineral particle characteristics can be obtained using a new form of SAXS tomography. Our strategy was to define a quantity with contributions adding up from all the voxels that the X-ray beam passes through during the experiment, which is a voxel property independent of its orientation in relation to the X-ray beam. This strategy then allows the use of standard reconstruction methods (Thorsten, 2011) to transform the projected data into a 3D map. We have demonstrated the feasibility of this procedure for the T parameter, which is



containing the blood vessels. (b) The 3D distance transformation of the $T_{\mathcal{R}}$ parameter map, taking the lacunae as reference points. (c) The correlation between the $T_{\mathcal{R}}$ parameter and attenuation coefficients reconstructed from synchrotron CT measurements. (d) A normalized frequency plot of the $T_{\mathcal{R}}$ parameter distribution in the two radial samples. (e) A normalized frequency plot of the $T_{\mathcal{R}}$ parameter distribution in the tangential and longitudinal samples. [Labels are rather fuzzy and/or small - is a revised figure available?]

1027 calculated as the ratio of two scattering invariants, the integrated intensity and the Porod constant, and which is an 1028 1029 important indicator of tissue maturity and its mechanical performance. 1030

A prerequisite of our approach is that the evaluated scat-1031 tering signal remains unchanged during the rotation of the 1032 sample. This rotational invariance is only fulfilled for scat-1033 tering around the rotational axis, and consequently only 1034 slightly more than 3% of the detector information is used in 1035 the evaluation. This restriction of the evaluation in reciprocal 1036 space corresponds to selecting a subpopulation of the mineral 1037 particles in real space. Only mineral particles with a normal 1038 parallel to the rotational axis are considered in the evaluation 1039 and are, therefore, described by the reported $T_{\mathcal{P}}$ parameter 1040 (which is, for this reason, denoted with a subscript). This 1041 specificity of our method for the particle orientation carries 1042 implications for its usefulness. In tissues with a preferred 1043 1044 matrix orientation, the investigation can focus on the mineral particles, which are embedded in the matrix in conformance 1045 with its preferred orientation. The type of bone used in this 1046 study - fibrolamellar bone - falls into this category, with the 1047 fibrous collagenous matrix preferentially orientated along the 1048 axis of the long bone. Similar to methods like polarized light 1049 microscopy, Raman spectroscopy and second-harmonic 1050 generation microscopy, which exploit orientational interaction 1051 effects with collagen, our method is able to detect variations in 1052 the alignment of mineral particles by yielding a detectable 1053 scattering signal only if particles are 'in plane', i.e. if the 1054 normals to their plate-like surfaces are parallel to the rota-1055 tional axis. 1056

SAXS is an important technique to characterize the 1057 nanostructure of inorganic-organic hybrid materials (Herwig 1058 & Fratzl, 2006) and SAXS tomography may become a 1059 1060 powerful way of mapping nanostructure variations in three dimensions within macroscopic specimens. Indeed, our 1061 approach is not limited to biological materials like bone. While 1062 our equations were derived for plate-like inclusions, they are 1063 valid more generally, and $T_{\mathcal{R}}$ is then an average chord length 1064 of the inclusions measured in the direction \mathcal{R} . Unfortunately, 1065 Porod's chord-length measure does not directly describe a 1066 particle thickness, as in the case of platelet-shaped inclusions, 1067 but it still represents an interesting size characteristic of any 1068 nanostructured two-phase system. 1069

An issue that has to be considered when studying biological 1070 materials with synchrotron radiation is the damage caused to 1071 the sample by the radiation. While radiation damage in X-ray 1072 tomographic approaches is known to affect the mechanical 1073 properties of bone strongly, mostly by the degradation of 1074 1075 collagen (Barth *et al.*, 2010), the shape of mineral particles is less affected. Although we cannot exclude the influence of 1076 radiation damage, we did not observe a change in the SAXS 1077 intensity from the mineral over time scales corresponding to 1078 an experiment. 1079

Our experimental setup allows the combination of several 1080 1081 methods using the same X-ray beam as the probe. In the current study the beam was used to measure the absorption 1082 coefficient, which provides information about the local 1083

calcium content of the bone, together with the scattering signal. In a similar way, the combination of methods could be extended to include chemical analysis [e.g. using X-ray fluorescence (Lange et al., 2011)]. With measurements based on the same incoming beam, the registration of different image data and the evaluation of the spatial correlation between different physical quantities [like the $T_{\mathcal{R}}$ parameter and Ca content, see Fig. 6(c)] are straightforward.

An advantage of our approach is the strongly reduced effort required for both the experimental and computational work compared with tensor tomography, when a specific parameter like the mineral particle thickness is sought. In the experiment, a single rotation of the sample is sufficient to obtain reconstructable information about the nanostructural thickness. As a result, the size of the data set for reconstruction is reduced and is, therefore, less demanding on the RAM of the reconstruction computer. In addition, the reduction in the dimensionality of the reconstruction problem allows the use of standard reconstruction algorithms, which have been optimized over the last few decades. Computational time can be saved by performing the preparatory evaluation of SAXS parameters during the measurement itself.

We tested our method on samples of fibrolamellar bovine bone. The preparation of a stick-like sample oriented in the radial direction and an imaged volume of 250 µm in this radial direction allowed us to image all the different layers within a complete fibrolamellar unit. As described by Magal et al. (2014), starting from a vascular channel the succession of layers are first lamellar bone around the channel, then a parallel-fibered layer, the primary hypercalcified layer in the center of the fibrolamellar unit, and then again the succession of a parallel-fibered layer and lamellar bone, before reaching the next layer with vascular channels. Based on the μ CT image of our sample, we observed vascular channels close to the top of the sample and in the lower part approximately 180 µm apart. Due to its branching, the lower vascular channel occupies a substantial area within a cross section of the sample [Fig. 4(c)].

Using the vascular channels as spatial references leads to the following interpretation of the 3D $T_{\mathcal{R}}$ parameter map of Fig. 5: the lamellar bone around the vascular channels displays low values of the T parameter, also clearly observable in the spatial correlation plot [Fig. 6(a)]. The highest values of the $T_{\mathcal{R}}$ parameters are found in two curved layers with a thickness of 30 to 40 µm, with the lower layer thicker than the upper one. These two layers are interleaved with a thinner layer $(\sim 15 \,\mu m$ thick), which comprises thinner mineral particles. The location of the layers and their thicknesses correspond exactly to the core of the fibrolamellar unit, with the central hypercalcified layer with a lower $T_{\mathcal{R}}$ parameter separating the layers of parallel-fibered bone with thick particles. The correlation plot of Fig. 6(c) shows that the lamellar bone not only has thinner mineral particles, but also the mineral content is lower compared with the parallel-fibered bone.

The 3D map of the T parameter provides new insights into the arrangement of the fibrolamellar units. While the lamellar bone around the vascular channel at the bottom results in a

Files: j/vg5128/vg5128.3d j/vg5128/vg5128.sgml VG5128 FA IU-2112/57(16)2 2112/56(16)2 ()

1129

1130

1131

1132

1133

1137

1138

1139

1140

1084

1085

1086

1087

1088

1089

1090

1091

1092

1093

1094

1095

1096

1097

1098

1099

1100

1101

1102

1103

1104

1105

1106

1107

1108

1109

1110

1111

1112

1113

1114

1115

1116

1117

1118

1119

1120

1121

1141 clear separation between the fibrolamellar units, the lamellar bone on top is quite localized around the more isolated 1142 1143 vascular channel, which results in a more continuous transition between the two fibrolamellar units at the top. In plexiform 1144 (fibrolamellar) ovine bone, a similar correspondence of the T1145 parameter and structural organization was found, i.e. rela-1146 tively high T parameters in regions with a higher degree of 1147 structural organization and lower T parameters in the more 1148 woven-like areas (Kerschnitzki et al., 2013). Testing different 1149 subpopulations of mineral particles showed differences in the 1150 $T_{\mathcal{R}}$ parameter [Figs. 6(d) and 6(e)]. While these measurements 1151 were performed on differently oriented samples, it is 1152 straightforward to test different subpopulations in the same 1153 sample by rotating the sample around more than one rota-1154 tional axis \mathcal{R} or by reconstructing the full 3D scattering 1155 pattern (Liebi et al., 2015; Schaff et al., 2015). In addition, our 1156 study is limited by a low sample number. A systematic 1157 1158 investigation of several fibrolamellar units in different loca-1159 tions of bovine bone and in different individuals would allow a quantitative assessment of biological variability. 1160

The experimental approach proposed in this study provides 1161 much more information about the sample than is actually used 1162 for the reconstruction. One might also implement the idea of 1163 virtual rotation axes, originally proposed by Schaff et al. 1164 (2015), into this simplified treatment of SAXS data where only 1165 invariants are reconstructed. This would probably make the 1166 reconstruction of particle sizes with many orientations more 1167 effective. Furthermore, the recent past has proved that 1168 important progress in imaging techniques occurs on the 1169 computational side of the process. For our problem we see a 1170 substantial potential to exploit information from the beam 1171 profile and the availability of two data sets (scattering and 1172 attenuation) in order to improve the reconstruction result in 1173 1174 combination with algebraic reconstruction tools (Hansen & Jørgensen, 2018). 1175

Finally, the success of a new method is always linked to 1176 interesting applications. In investigating mineralized tissues, 1177 the most pressing problems encountered are posed by bone 1178 1179 diseases. While mineralization disorders are often characterized in terms of the amount of mineral incorporated in the 1180 bone (Roschger et al., 2008), our method would allow a 1181 mapping of structural disorders of the mineral nanostructure. 1182

Acknowledgements

1183

1184

1188

1189

1185 Open access funding enabled and organized by Projekt 1186 DEAL. 1187

Funding information

The authors wish to acknowledge support from BiGmax, the 1190 Max Planck Society's Research Network on Big-Data-Driven 1191 Materials Science. We also acknowledge Diamond Light 1192 Source for the generous award of beamtime (SM18524-2). H. S. 1193 Gupta thanks the Biotechnology and Biological Sciences 1194 1195 Research Council (grant No. BB/R003610/1) and UK Research and Innovation (grant No. MR/R025673/1) for 1196 research grant funding. 1197

Files: j/vg5128/vg5128.3d j/vg5128/vg5128.sgml VG5128 FA IU-2112/57(16)2 2112/56(16)2 ()

References

- Bar-On, B. & Wagner, H. D. (2013). J. Struct. Biol. 183, 149-164.
- Barth, H. D., Launey, M. E., MacDowell, A. A., Ager, J. W. III & Ritchie, R. O. (2010). Bone, 46, 1475-1485.
- Basham, M., Filik, J., Wharmby, M. T., Chang, P. C. Y., El Kassaby, B., Gerring, M., Aishima, J., Levik, K., Pulford, B. C. A., Sikharulidze, I., Sneddon, D., Webber, M., Dhesi, S. S., Maccherozzi, F., Svensson, O., Brockhauser, S., Náray, G. & Ashton, A. W. (2015). J. Synchrotron Rad. 22, 853-858.
- Benecke, G., Wagermaier, W., Li, C., Schwartzkopf, M., Flucke, G., Hoerth, R., Zizak, I., Burghammer, M., Metwalli, E., Müller-Buschbaum, P., Trebbin, M., Förster, S., Paris, O., Roth, S. V. & Fratzl, P. (2014). J. Appl. Cryst. 47, 1797-1803.
- Bouville, F., Maire, E., Meille, S., Van de Moortèle, B., Stevenson, A. J. & Deville, S. (2014). Nat. Mater. 13, 508-514.
- Buzug & Thorsten, M. (2011). Handbook of Medical Technology: Computed Tomography. Heidelberg; Springer.
- Feldkamp, J. M., Kuhlmann, M., Roth, S. V., Timmann, A., Gehrke, R., Shakhverdova, I., Paufler, P., Filatov, S. K., Bubnova, R. S. & Schroer, C. G. (2009). Phys. Status Solidi A, 206, 1723-1726.
- Filik, J., Ashton, A. W., Chang, P. C. Y., Chater, P. A., Day, S. J., Drakopoulos, M., Gerring, M. W., Hart, M. L., Magdysyuk, O. V., Michalik, S., Smith, A., Tang, C. C., Terrill, N. J., Wharmby, M. T. & Wilhelm, H. (2017). J. Appl. Cryst. 50, 959-966.
- Fratzl, P., Fratzl-Zelman, N., Klaushofer, K., Vogl, G. & Koller, K. (1991). Calcif. Tissue Int. 48, 407-413.
- Fratzl, P. & Weinkamer, R. (2007). Mater. Sci. 52, 1263-334.
- Fratzl-Zelman, N., Schmidt, I., Roschger, P., Glorieux, F. H., Klaushofer, K., Fratzl, P., Rauch, F. & Wagermaier, W. (2014). Bone, 60, 122-128.
- Gao, Z., Guizar-Sicairos, M., Lutz-Bueno, V., Schröter, A., Liebi, M., Rudin, M. & Georgiadis, M. (2019). Acta Cryst. App. 75.
- Georgiadis, M., Guizar-Sicairos, M., Gschwend, O., Hangartner, P., Bunk, O., Müller, R. & Schneider, P. (2016). PLoS One, 11, e0159838.
- Georgiadis, M., Guizar-Sicairos, M., Zwahlen, A., Trüssel, A. J., Bunk, O., Müller, R. & Schneider, P. (2015). Bone, 71, 42-52.
- Guinier, A. & Fournet, G. (1955). Small-angle Scattering of X-rays. New York: Wiley
- Hansen, P. C. & Jørgensen, J. S. (2018). Numer. Algor, 79, 107-137.
- He, F., Chiou, A. E., Loh, H. C., Lynch, M., Seo, B. R., Song, Y. H., Lee, M. J., Hoerth, R., Bortel, E. L., Willie, B. M., Duda, G. N., Estroff, L. A., Masic, A., Wagermaier, W., Fratzl, P. & Fischbach, C. (2017). Proc. Natl Acad. Sci. USA, 114, 10542-10547.
- Herwig, P. & Fratzl, P. (2006). Monatsh. Chem. 137, 529-543.
- Jäger, I. & Fratzl, P. (2000). Biophys. J. 79, 1737-1746.
- Jaschouz, D., Paris, O., Roschger, P., Hwang, H.-S. & Fratzl, P. (2003). J. Appl. Cryst. 36, 494-498.
- Jensen, T. H., Bech, M., Bunk, O., Thomsen, M., Menzel, A., Bouchet, A., Le Duc, G., Feidenhans'l, R. & Pfeiffer, F. (2011). Phys. Med. Biol. 56, 1717-1726.
- Kerschnitzki, M., Kollmannsberger, P., Burghammer, M., Duda, G. N., Weinkamer, R., Wagermaier, W. & Fratzl, P. (2013). J. Bone Miner. Res. 28, 1837–1845.
- Lange, C. C., Li, I., Manjubala, W., Wagermaier, J., Kühnisch, M., Kolanczyk, S., Mundlos, P., Knaus, P. & Fratzl, P. (2011). J. Struct. Biol. 176, 159-167.
- Liebi, M., Georgiadis, M., Kohlbrecher, J., Holler, M., Raabe, J., Usov, I., Menzel, A., Schneider, P., Bunk, O. & Guizar-Sicairos, M. (2018). Acta Cryst. A74, 12-24.
- Liebi, M., Georgiadis, M., Menzel, A., Schneider, P., Kohlbrecher, J., Bunk, O. & Guizar-Sicairos, M. (2015). Nature, 527, 349-352.
- Magal, Almany, R., Reznikov, N., Shahar, R. & Weiner, S. (2014). J. Struct. Biol. 186, 253-64.
- Meyers, M. A., Chen, P.-Y., Lin, A. Y.-M. & Seki, Y. (2008). Prog. Mater. Sci. 53, 1-206.

1198

1199

1200

1201

1202

1203

1204

1206

1219

1220

1225 1226

1227 1228

1229 1230 1231

1235 1236 1237

1238 1239

1240

1241 1242

1243 1244

1245 1246

1247

1248 1249

1250 1251 1252

1253

1254

11 of 12

Paolino De Falco et al. • Tomographic X-ray scattering from bone

1255	Milovanovic, P., Zimmermann, E. A., Riedel, C., vom Scheidt, A.,
1256	Herzog, L., Krause, M., Djonic, D., Djuric, M., Püschel, K. &
1257	Amling, M. (2015). Biomaterials, 45, 46-55.
	Pabisch, S., Wagermaier, W., Zander, T., Li, C. & Fratzl, P. (2013).
1258	Methods in Enzymology, Vol.?, Vol. title and editors? Chapter?,
1259	Page range?. Amsterdam; Elsevier. [Please complete reference]
1260	Paris, O., Li, C., Siegel, S., Weseloh, G., Emmerling, F., Riesemeier, H.,
1261	Erko, A. & Fratzl, P. (2007). J. Appl. Cryst. 40, s466-s470.
1262	Paris, O. I., Zizak, H., Lichtenegger, P., Roschger, K., Klaushofer, K.

1262 & Fratzl, P. (2000). Cell. Mol. Biol. 46, 993–1004.
 1263 Bathi S. P. Lin Dohn D. W. Dorne, L. P. Estroff, L. A.

Pathi, S. P., Lin, Debra D. W., Dorvee, J. R., Estroff, L. A. &
 Fischbach, C. (2011). *Biomaterials*, 32, 5112–5122.

- Rinnerthaler, S. P., Roschger, P., Jakob, H. F., Nader, K., Klaushofer,
 K. & Fratzl, P. (1999). *Calcif. Tissue Int.* 64, 422–429.
- 1266
 K. & Fratzl, P. (1999). Calcif. Tissue Int. 64,

 1267
 Ritchie, R. O. (2011). Nat. Mater. 10, 817–822.

Roschger, P., Paschalis, E. P., Fratzl, P. & Klaushofer, K. (2008). Bone,
 42, 456–466.

Roschger, P., Grabner, B. M., Rinnerthaler, W., Tesch, M., Kneissel,
 A., Berzlanovich, K., Klaushofer, K. & Fratzl, P. (2001). J. Struct.
 Biol. 136, 126–136.

Schaff, F., Bech, M., Zaslansky, P., Jud, C., Liebi, M., Guizar-Sicairos, M. & Pfeiffer, F. (2015). *Nature*, **527**, 353–356.

Schroer, C. G., Kuhlmann, M., Roth, S. V., Gehrke, R., Stribeck, N., Almendarez-Camarillo, A. & Lengeler, B. (2006). *Appl. Phys. Lett.* **88**, 164102.

Seidel, R., Gourrier, A., Kerschnitzki, M., Burghammer, M., Fratzl, P.,

- Gupta, H. S. & Wagermaier, W. (2012). *Biointerphases* 1, 123–131.
 Seto, J., Gupta, H. S., Zaslansky, P., Wagner, H. D. & Fratzl, P. (2008). *Adv. Funct. Mater.* 18, 1905–1911.
- Smith, A. J., Davidson, L. S., Emmins, J. H., Bardsley, J. C. P., Holloway, M., Marshall, M. A. R., Pizzey, C. L., Rogers, S. E. & Shebanova, O. (2019). arXiv:1903.05405.
- Studart, A. R. (2013). Adv. Funct. Mater. 23, 4423-4436.
- Wagermaier, W. H. S., Gupta, A., Gourrier, M., Burghammer, P., Roschger & Fratzl, P. (2006). *Biointerphases*, 1, 1–5.
- Weiner, S., Traub, W. & Wagner, H. D. (1999). J. Struct. Biol. 126, 241–255.
- Weiner, S. & Wagner, H. D. (1998). Annu. Rev. Mater. Sci. 28, 271–298.
- Weinkamer, R. & Fratzl, P. (2016). MRS Bull. 41, 667-671.
- Xi, L., Paolino De Falco, E., Barbieri, A., Karunaratne, L., Bentley, C. T. E., Terrill, N. J., Brown, S. D. M., Cox, R. D. & Davis, G. R. (2018). *Acta Biomater.* **76**, 295–307.
- Zizak, I. P., Roschger, O., Paris, O., Misof, B. M., Berzlanovich, S., Bernstorff, H., Amenitsch, K., Klaushofer, K. & Fratzl, P. (2003). J. Struct. Biol. 141, 208–217.



ISSN: 1600-5767

ORDER FORM

YOU WILL AUTOMATICALLY BE SENT DETAILS OF HOW TO DOWNLOAD AN ELECTRONIC REPRINT OF YOUR PAPER, FREE OF CHARGE. PRINTED REPRINTS MAY BE PURCHASED USING THIS FORM.

Please scan your order and send to Is@iucr.org

INTERNATIONAL UNION OF CRYSTALLOGRAPHY

5 Abbey Square

Chester CH1 2HU, England.

VAT No. GB 161 9034 76

Article No.: J210088-VG5128

Title of article Tomographic X-ray scattering based on invariant reconstruction – analysis of the 3D nanostructure of bovine bone

Name Wolfgang Wagermaier

Address Department of Biomaterials, Max Planck Institute of Colloids and Interfaces, Potsdam 14476, Germany

E-mail address (for electronic reprints) wolfgang.wagermaier@mpikg.mpg.de

OPEN ACCESS

Your article appears to qualify for open-access publication at no charge under a transformative deal with the institution of your corresponding author. For more information on open-access arrangements for this journal, please go to https://journals.iucr.org/j/services/openaccess.html.

DIGITAL PRINTED REPRINTS

I wish to order paid reprints

These reprints will be sent to the address given above. If the above address or e-mail address is not correct, please indicate an alternative:

PAYMENT (REPRINTS ONLY)

Charge for reprints USD							
An official purchase order made out to INTERNATIONAL UNION OF CRYSTALLOGRAPHY	is enclosed	will follow					
Purchase order No.							
Please invoice me							
I wish to pay by credit card							
EU authors only: VAT No:							
EU authors only: VAT No:							

Date Sig	ignature
----------	----------

DIGITAL PRINTED REPRINTS

An electronic reprint is supplied free of charge.

Printed reprints without limit of number may be purchased at the prices given in the table below. The requirements of all joint authors, if any, and of their laboratories should be included in a single order, specifically ordered on the form overleaf. All orders for reprints must be submitted promptly.

Prices for reprints are given below in United States dollars and include postage.

	Size of paper (in printed pages)				
Number of reprints required	1–2	3–4	5–8	9–16	Additional 8's
50	184	268	372	560	246
100	278	402	556	842	370
150	368	534	740	1122	490
200	456	664	920	1400	610
Additional 50's	86	128	178	276	116

PAYMENT AND ORDERING

Official purchase orders should be made out to INTERNATIONAL UNION OF CRYSTALLOGRAPHY.

Orders should be returned by email to Is@iucr.org

ENQUIRIES

Enquiries concerning reprints should be sent to support@iucr.org.