Sensitive periods for social development: Interactions between predisposed and learned mechanisms

3

4 Orsola Rosa-Salva^a, Uwe Mayer^a, Elisabetta Versace^{b,a}, Marie Hébert^a, Bastien S.
5 Lemaire^a, Giorgio Vallortigara^{a §}

6

^a Center for Mind/Brain Sciences, University of Trento, Piazza Manifattura 1, 38068
Rovereto (TN), Italy

9

^b Department of Biological and Experimental Psychology, School of Biological and
 Chemical Sciences, Queen Mary University of London, 327 Mile End Road, London E1
 4NS, United Kingdom

13

[§] Corresponding author, <u>giorgio.vallortigara@unitn.it</u>

15

16

19 Abstract

20 We analysed research that makes use of precocial species as animal models to describe the interaction of predisposed mechanisms and environmental factors in early learning, 21 in particular for the development of social cognition. We also highlight the role of 22 23 sensitive periods in this interaction, focusing on domestic chicks as one of the main animal models for this field. In the first section of the review, we focus on the 24 emergence of early predispositions to attend to social partners. These attentional 25 26 biases appear before any learning experience about social stimuli. However, non-27 specific experiences occurring during critical periods of the early post-natal life determine the emergence of these predisposed mechanisms for the detection of social 28 29 partners. Social predispositions have an important role for the development learningbased social cognitive functions, showing the interdependence of predisposed and 30 learned mechanisms in shaping social development. In the second part of the review 31 we concentrate on the reciprocal interactions between filial imprinting and 32 spontaneous (not learned) social predispositions. Reciprocal influences between these 33 two sets of mechanisms ensure that, in the natural environment, filial imprinting will 34 target appropriate social objects. Neural and physiological mechanisms regulating the 35 36 sensitive periods for the emergence of social predispositions and for filial imprinting learning are also described. 37

Keywords: sensitive periods, predispositions, learning, social cognition, domestic chicks

44 Glossary

45 Altricial: animal born or hatched in a very immature condition so that the young
46 require intense parental care.

47 **c-Fos**: protein product of the corresponding Immediate Early Gene *c-fos* that is acting
48 as a transcription factor and is commonly used as a marker to quantify neural activity
49 and plasticity.

50 **Deprivation experiment**: involves keeping an animal in an environment that has been 51 systematically impoverished of a given type of stimuli. The rationale of these 52 experiments is usually to deprive animals of a certain experience, to see whether the 53 behaviour or trait of interest is nonetheless displayed.

Filial imprinting: rapid form of learning by exposure, through which the young of precocial bird species learn to recognize the object(s) to which they are exposed during a sensitive period. By this process, the young chick restricts its affiliative behaviour towards the first salient object(s) it encounters. In the natural environment, filial imprinting usually results in social attachment towards the mother hen and/or the brood mates.

Immediate Early Genes (IEG): a class of genes that are rapidly expressed in response
 to neural activation. Their products are often used as neural activity markers.

62 **IMM (Intermediate Medial Mesopallium)**: associative brain region of the avian 63 pallium (homolog to mammalian cortexes), involved in the memory storage for filial 64 imprinting learning in domestic chicks.

65 **Mesotocin:** this hormone, which regulates many social behaviours, is the oxytocin-66 equivalent of nonmammalian tetrapods. Receptors for these neuropeptides are present 67 in areas of the Social Behaviour Network.

Point-light display: animation created by placing light-points on the major joints of a walking body and obscuring the rest of the image. This sort of stimulus conveys many of the dynamic properties of the portrayed motion while removing most information on the configuration and appearance of the moving body.

Precocial: animal born or hatched in a very mature condition so that the young do not need intensive care, being able to perform autonomously many behavioural and physiological functions.

Predispositions: unlearned tendencies to react in predetermined ways (e.g., with approach or avoidance) to stimuli featuring specific characteristics (motion, colour, configuration, etc.). Predispositions are displayed by newborn or newly-hatched animals or by naïve animals, deprived of any experience with a given category of stimuli. Usually, the features that elicit predispositions are typical of objects of great biological significance, such as potential social companions, preys or predators.

Self-propulsion: propulsion powered by an energy source internal to the moving object. It is one of the main features that distinguish animate creatures from inanimate objects, which can be set in motion only by the action of external forces.

Sensitive period: limited periods during which specific mechanisms are more likely to take place and the effects of experience on developing organisms are stronger than in the rest of the lifespan.

Semi-rigid motion: Kind of movement, typically associated with biological motion. Some points maintain a fixed distance from each other, while their distance to other points varies. This motion pattern is exhibited by vertebrates and other legged animals and differs from the cinematics of rigid translation and random movement shown by most inanimate objects.

Social Behaviour Network: a set of interconnected subcortical areas, rich of sex steroid hormone receptors, highly conserved across vertebrates and in charge of many
 social behaviours.

95 **T3 (triiodothyronine):** tyrosine-based hormone synthesized by the thyroid gland 96 involved in the regulation of many physiological processes in the body. T3 represents 97 the primary metabolically active thyroid hormone in avian species and has been 98 recently implicated in the control of the sensitive window for filial imprinting.

99

101 **1. General introduction**

Throughout the tradition of developmental psychology, the role of predispositions 102 (spontaneous preferences and behaviours) and experience-based learning mechanisms 103 104 in determining cognitive development has been a central topic. The writings of Jacques 105 Melher pay a special attention towards the way evolution equipped infants with specialised mechanisms, predisposed to perform specific tasks of great adaptive value. 106 107 These mechanisms provide pre-representations at the basis of all subsequent knowledge (Mehler & Dupoux, 1994). Interestingly, although the early predisposed 108 109 mechanisms envisioned by Mehler and Dupoux do not depend on direct learning 110 experience, they are not experience-independent. On the contrary, environmental influences act by selecting, triggering and regulating them. Here, we provide an 111 112 overview of the research on the dynamic interplay of predisposed and learned 113 mechanisms in the development of social cognition during the early post-natal life. In doing so, we focus on the temporal dynamics of these interactions. 114

Experiences occurring during specific ontogenetic stages are particularly influential on subsequent cognitive, social and neural development (Bateson & Gluckman, 2012; Chan, 2014; Charil et al., 2010; Hubel & Wiesel, 1970; Kalcher-Sommersguter et al., 2015; Lorenz, 1937; Zeanah et al., 2009), as shown by the influential notion of sensitive periods. These are time windows in which the effects of environmental stimuli on developing organisms are stronger than in the rest of the lifespan ((Bodin, Yeates, & Cass, 2011) for the distinction between sensitive and critical periods, Dehorter & Del

Pino, 2020; Knudsen, 2004). We describe the sensitive periods in which different 122 environmental factors are particularly effective and how they are modulated by genetic 123 or environmental influences. This shows how the timing of events occurring in the 124 125 species-typical environment can shape social development. As model systems, we will 126 mostly refer to domestic chicks and their social development (Di Giorgio, Loveland, et al., 2017; Lorenzi et al., 2020; Rosa-Salva, Mayer, & Vallortigara, 2015). Precocial birds 127 offer advantages such as the presence of a mature sensory-motor system and of fast 128 129 learning mechanisms already in hatchlings (e.g., filial imprinting). For these reasons, they have been central to understand the interface between predisposed and learned 130 mechanisms at the beginning of life. 131

Research in precocial birds revealed that, even in the pre-hatching phase, experience 132 133 and genetically determined mechanisms interact in the development of socio-cognitive skills. Thanks to the in ovo development and the possibility to test behavioural 134 135 responses already shortly after hatching, precocial birds (Versace, 2017) provide 136 optimal models for the investigation of pre-natal influences on behavioural outcomes. This has been exploited by studies on the development of neuroanatomical and 137 functional lateralization in domestic chicks, revealing how the interaction of genetic 138 and experience-based mechanism within specific critical periods of the pre-natal life 139 can modulate cognitive functions ((Rogers, 1982, 1997); (Deng & Rogers, 1997; Rogers 140 & Bolden, 1991; Rogers & Deng, 1999; Rogers & Sink, 1988); (Deng & Rogers, 2002a); 141 (Rogers, 1990); (Dharmaretnam & Rogers, 2005) (Lorenzi et al., 2019); (Chiandetti, 142 2011; Chiandetti & Vallortigara, 2019); (Chiandetti et al., 2013)). 143

In this review, however, we analyse how social and cognitive development are affected 144 145 by the interplay between predisposed and learning mechanisms during the first stages of post-natal life. The review is divided into two main sections that follow an 146 ontogenetic order, describing mechanisms occurring in the early postnatal life and then 147 148 in the filial imprinting phase. In the first section, we focus on the mechanisms underlying the emergence of early predispositions to attend to social partners. These 149 150 predispositions develop in the earliest stages of the postnatal life, in visually naïve 151 chicks. To clarify whether a behaviour is spontaneous or acquired, researchers have used deprivation experiments. This method consists in depriving animals of a certain 152 153 experience to see whether the behaviour of interest is displayed nonetheless (Lorenz, 1965). Animals are either tested before they have performed the activity, at birth or 154 hatching, or reared in isolation to eliminate opportunities for learning through 155 observation. These procedures are facilitated when embryos can be directly controlled 156 and animals hatch individually, making deprivation experiments particularly suitable for 157 158 oviparous species. Hence, it comes as no surprise that avian species have been central 159 to investigate the ontogenetic origins of behaviour. The ethological tradition has emphasized stereotypical and species-specific fixed action patterns that do not depend 160 161 on experience (Schleidt, 2010). Originally, behavioural biologists focused on the notion of instinct and complex sequences of relatively fixed adult behaviour (Burghardt & 162 Bowers, 2017; Eibl-Eibesfeldt, 1975). More recently, scientists have identified early 163 164 cognitive traits and spontaneous preferences (predispositions) that orient the behaviour of neonate, inexperienced animals, towards particular stimuli. As detailed below (see 165 2.1), converging evidence across neonates of different amniote species (e.g. domestic 166

chicks, human infants, tortoise hatchlings) tested with the deprivation method suggests 167 168 the presence of predisposed preferences to attend to stimuli associated with the presence of animate, living beings, the so-called "animacy detectors" (Di Giorgio et al., 169 2017a; Rosa-Salva et al., 2015; Vallortigara, 2012; Vallortigara, in press). These 170 171 preferences, in the literature on domestic chicks and human infants, are often called "social predispositions". This is because, in the natural environment of social species, 172 these early predispositions will result in preferential attention towards appropriate 173 174 social partners. Differently from the fully structured, species-specific behaviours targeted by classical behavioural biologists, early predispositions appear as building 175 blocks of the developing cognitive abilities of the vertebrate mind. These early social 176 predispositions, whose neural basis we are starting to uncover in chicks (2.2), appear to 177 contribute to the development and specialization of the brain circuits that, in adults, 178 carry out sophisticated social information processing (Johnson, 2005). This may be 179 achieved even by simply directing visual attention towards appropriate social stimuli, 180 181 biasing the visual input received by the subject during a critical period of the postnatal development. Indeed, anomalies of early social predispositions have been associated 182 with abnormalities in the development of social cognition, such as those associated 183 184 with disorders of the autistic spectrums (Di Giorgio et al., 2016). However, even though social predispositions do not require any specific learning experience about social 185 stimuli, their emergence unfolds also through the action of environmental influences. In 186 187 the first section of the review, we thus describe how non-specific experiences occurring during well-defined critical periods of the early post-natal life determine the 188 189 emergence of predisposed mechanisms for the detection of social partners (and

animate creatures in general) (see 2.2). These predispositions have an important role in shaping the future development of more sophisticated, learning-based social cognitive functions. This literature beautifully shows the crucial interdependence of predisposed and learned mechanisms in shaping social development.

In the second section of the review (section 3), we will focus on filial imprinting 194 195 (Vallortigara & Versace, 2018), a very prominent form of learning, which is crucial for the survival and subsequent social adaptation of young chicks. Through filial 196 imprinting, young precocial birds learn to recognize the object(s) to which they are 197 exposed during a sensitive period. By this process, in the absence of any overt 198 reinforcement, the young chick restricts its affiliative behaviour towards the first salient 199 200 object(s) it encounters. In the natural environment, filial imprinting usually results in 201 social attachment towards the mother hen and/or the brood mates (Bateson, 1966; Bolhuis, 1991; McCabe, 2019). This has, of course, crucial implications for social 202 203 development, allowing chicks not only to receive parental care from the mother hen, 204 but also to maintain group cohesion. Filial imprinting can thus provide the foundations for the formation and maintenance of stable social groups, whose internal hierarchy 205 (pecking order) regulates "chicken societies". By being exposed to conspecifics during 206 207 development, chickens develop sophisticated social skills. These include the ability to recognize familiar conspecifics and their dominance hierarchy (Deng & Rogers, 2002b; 208 Rogers & Workman, 1989; Vallortigara & Andrew, 1994; Vallortigara & Andrew, 1991; 209 Vallortigara, 1992), the capacity to acquire information about food sources by 210 observing others' behaviour (Nicol, 2004) and even forms of perspective-taking and 211 social deception (Marino, 2017). In the last part of the review, we will describe how 212

experience-driven mechanisms and social predispositions can influence the course of 213 214 filial imprinting, directing it towards appropriate objects. We will discuss how the emergence of social predispositions facilitates imprinting also towards non-predisposed 215 features of the stimuli. Moreover, we will see that even after an initial imprinting 216 217 towards an inanimate object, predispositions can emerge, which then direct subsequent secondary imprinting to appropriate social companions. We will see how 218 experience occurring in specific time windows of the perinatal life in turn affects the 219 220 duration of the sensitive period for imprinting. At least some of the brain regions involved in filial imprinting have been identified with a good degree of certainty (e.g., 221 the Intermediate Medial Mesopallium, or IMM, see Horn, 2004). Some of the 222 physiological mechanisms that control the opening and closing of the sensitive period 223 for filial imprinting, though the actions of specific hormones and neurotransmitters, 224 have been recently identified also (Aoki et al., 2018; Yamaguchi et al., 2012). This 225 literature reveals the presence of interdependent relationships between experience-226 227 based and predisposed mechanisms.

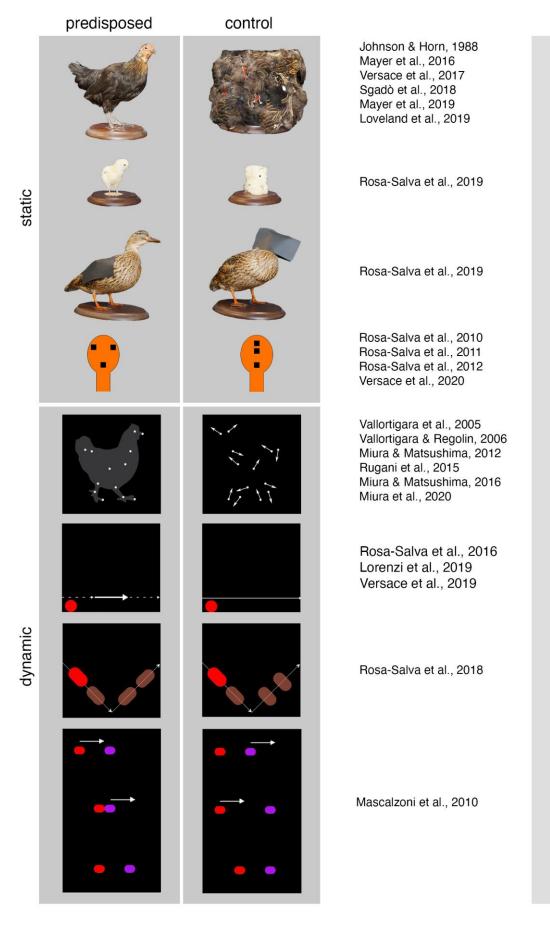
228

229 2. Early post-natal life: sensitive periods for the emergence of predispositions

230 2.1 The study of early predispositions as adaptive responses

In this section, we analyse the evidence of preparedness to attend to social stimuli in the early post-natal life, as revealed by social predispositions in domestic chicks (the main model considered in this review). Early predispositions to orient towards and

engage with particular stimuli have been observed for both static features and motion 234 dynamics. Interestingly, the preferred features are associated with the presence of 235 animate creatures (see Figure 1). Among static features, preferences have been 236 observed for particular colours (e.g. red and blue are preferred to yellow and green 237 238 (Kovach, 1971; Miura, Nishi, & Matsushima, 2020; Salzen, Lily, & McKeown, 1971; Salzen & Meyer, 1968; Taylor, Sluckin, & Hewitt, 1969), shapes (Hess & Goodwin, 1969; 239 Salzen & Meyer, 1968; Schulman, Hale, & Graves, 1970), for hollow objects (Versace et 240 241 al., 2016), for faces and for face-like configurations (in newborns, Johnson & Morton, 1991; Morton & Johnson, 1991; chicks, Rosa-Salva, Regolin, & Vallortigara, 2010, 2012; 242 Rosa Salva et al., 2011; monkeys, Sugita, 2008; and tortoises Versace, Damini, & 243 Stancher, 2020). A seminal work by Johnson and Horn (Johnson & Horn, 1988) has 244 shown that, in the first hours after hatching, dark-reared chicks deprived from 245 experience with conspecifics exhibit a preference to orient towards a stuffed fowl 246 compared to a disassembled version of a similar fowl (Bolhuis, Johnson, & Horn, 1989; 247 Egorova & Anokhin, 2003; Rosa-Salva et al., 2015; Versace et al., 2017). Subsequent 248 experiments have shown that this predisposition is not specifically tuned to hens or 249 fowls, but is based on low-level cues present in the area of the neck and face (Johnson 250 251 & Horn, 1988; Rosa-Salva, Mayer & Vallortigara, 2019).





255 Fig. 1. Schematic representation of the main classes of stimuli used to test social predispositions for

256 static and dynamic features typical animate objects. For each pair of stimuli, the preferred object 257 featuring the predisposed trait and a control stimulus are presented side by side, followed by a selection 258 of papers reporting a behavioural preference for the predisposed object in naïve animals. From above, 259 the first two images of the first panel represent the stimuli used to test the preference for hen-like 260 objects (a stuffed junglefowl-like hen versus a scrambled version of a similar specimen). In the following 261 rows: a pair of similar stimuli obtained from stuffed chick models; a stuffed duck with her wings 262 occluded compared to a similar exemplar with the head region occluded; a schematic face-like stimulus 263 and a non-face control image. The first two images of the second panel represent a point light display 264 of a walking hen and a control stimulus with random motion of the same dots (the silhouette of the hen 265 has been added for illustrative purposes). In the following two rows: a schematic representation of a 266 speed changing stimulus and its speed-constant control; an object that always moves in the direction of 267 its main body axis and its control stimulus. In the last row, we represented on the left the sequence of 268 movement of a self-propelled red object hitting and putting in motion a non-self-propelled purple 269 object (the sequence has to be read from above to below). In this case, chicks preferentially imprint on 270 the red objects. On the right, both objects appear self-propelled and chicks display no preferences 271 between the two. In all dynamic stimuli arrows have been added for illustrative purposes.

272 Deprivation studies on social predispositions have been conveniently conducted on domestic chicks and other precocial species, thanks to the ease of testing precocial 273 274 animals that are born with a mature sensory and motor system (Versace, 2017). However, the preference for faces and face-like configurations has been extensively 275 observed also in human infants (Buiatti et al., 2019; Di Giorgio et al., 2016; Goren, Sarty 276 277 & Wu, 1975; Morton & Johnson, 1991; Simion & Di Giorgio, 2015), human fetuses (Reid et al., 2017), monkeys (Sugita, 2008) and recently in tortoise hatchlings (Versace 278 et al., 2020). This suggests the presence of an ancient mechanism for the detection of 279 280 faces, which is tuned to low-level features associated with the presence of faces. This

281 mechanism, active soon after birth or hatching, does not specify the species-specific or 282 individual details of faces, but increases the chances that newborn animals orient 283 towards other animals by directing their attention toward any face-like configuration.

Dynamic patterns are also very effective in attracting the attention of newborn animals. 284 Although precocial avian species can exhibit affiliative responses to still objects, the 285 phenomenon of filial imprinting is enhanced by the presence of moving objects (ten 286 Cate, 1986). This suggests that the first orienting responses of animals, before they 287 developed affiliative responses to particular objects they have experienced, might be 288 driven by cues of animacy, the property of "being alive". In line with this idea, we have 289 observed early preferences of visually inexperienced animals for motion dynamics 290 associated with the presence of animate, living beings. This supports the idea of an 291 292 animacy-detector tuned to some of the features that distinguish the biological motion of animate creatures (Mascalzoni, Regolin, & Vallortigara, 2010; Rosa-Salva et al., 2016; 293 294 Rosa-Salva et al., 2018; Vallortigara, Regolin, & Marconato, 2005). Vallortigara and 295 colleagues (2005) have initially discovered a spontaneous preference for point-light displays that move according to semi-rigid biological motion. In this kind of 296 movement, points located on different parts of the body move relative to one another 297 within an elastic, constrained range. This is different from the cinematics of rigid 298 translation and random movement shown by most inanimate objects. The preference 299 for biological motion has been documented in human neonates too (Simion, Regolin, 300 & Bulf, 2008). Social predispositions for animate motion can be observed using very 301 simplified visual stimuli. For instance, visually naïve chicks prefer to imprint on objects 302 that start to move on their own, rather than being pushed in motion by a collision with 303

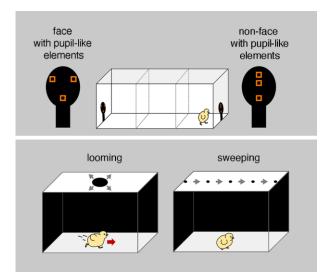
another object (Mascalzoni et al., 2010; Simion et al., 2008). Likewise, the ability to 304 spontaneously start to move from a resting state ("start from rest") elicits visual 305 preferences also in human newborns (Di Giorgio et al., 2017b). Similarly, both chicks 306 and human infants prefer to approach objects that change in speed and, in the case of 307 308 chicks, rotate autonomously (Frankenhuis et al., 2013; Hernik, Fearon, & Csibra, 2014; Rosa-Salva et al., 2016, 2018). These are all features that signal the presence of an 309 internal energy source in self-propelled objects (a defining feature for the 310 311 discrimination of animate creatures). Moreover, a predisposition for objects that move in the direction of their main body-symmetry axis, as most bilaterians, is present in 312 both infants (Hernik et al., 2014) and visually-inexperienced chicks (Rosa-Salva et al., 313 2018). Overall, compelling evidence suggests that early approach responses are 314 facilitated by cues associated with animate objects. 315

An important function of early preferences might be to direct the attention of the 316 young animals towards the animate objects that will provide care, and towards which 317 318 young animals should develop affiliative responses and sexual responses later on (Morton & Johnson, 1991; Versace et al., 2018). As mentioned in the introduction, this 319 would also enable the subsequent development of neural mechanisms specialised for 320 321 the processing of various aspects of social information (Di Giorgio et al., 2016; Johnson, 2005). This idea is supported by evidence of enhanced imprinting responses elicited by 322 predisposed stimuli, such as red colour on the head of a creature that moves 323 according to biological motion (Lemaire et al., 2020; Miura et al., 2020; Miura & 324 Matsushima, 2016) or such as "start from rest" (Mascalzoni et al., 2010). However, 325 similar early predispositions have been found in solitary animals with no parental care, 326

such as land tortoises (Versace et al., 2020). This suggests that mechanisms to direct 327 attention towards animate objects might respond to a general need of identifying 328 living agents to gain information about important aspects of life, including the 329 presence of resources, preys and predators (Lorenzi & Vallortigara, 2020; Vallortigara, 330 331 2012; Versace et al., 2020). The predispositions for some stimuli exhibited at the beginning of life, though, are not rigidly prescriptive, as revealed by the fact that both 332 chicks (Versace et al., 2017) and young tortoises (Versace et al., 2018) can show 333 334 exploration of alternative stimuli. The propensity to focus only on the predisposed stimuli or explore alternative stimuli has a genetic basis in chicks, as shown by the 335 differences in perseverance in approaching a stuffed hen in visually naïve chicks 336 337 (Versace et al., 2017).

338 In line with the idea that predispositions direct attention to stimuli relevant for an individual's fitness, we have recently shown that spontaneous responses that do not 339 require experience include anti-predator defensive behaviours. Although this topic has 340 341 received less attention than social predispositions, to date studies in chicks (and mallard ducklings, Dessborn et al., 2012) have shown that precocial birds are able to 342 recognize and appropriately react to various threats, and this in the absence of 343 previous experience. A few decades ago, Schiff reported that dark-hatched and -reared 344 chicks escape a stimulus rapidly looming in front of them (Schiff, 1965). Recently, we 345 have shown that young chicks reared with no experience with moving stimuli 346 spontaneously assess the difference between distant and approaching threats posed by 347 stimuli moving overhead. Being presented with stimuli that mimicked either an 348 approaching threat (a 'looming' stimulus increasing in size like an approaching 349

350 predator) or a distant threat (a 'sweeping' stimulus moving at a constant distance like a cruising prey bird), naïve chicks modulated their defensive responses accordingly 351 352 (Hébert, Versace, & Vallortigara, 2019). They ran away from an approaching stimulus and froze in the presence of a distant threat (see Fig. 2). Static visual stimuli can also 353 354 trigger innate anti-predatory responses: while chicks are attracted by 'face-like' stimuli 355 (a silhouette containing three dark blobs organized in a triangular fashion) over a control stimulus (Rosa-Salva et al., 2009) (Fig. 1), they avoid stimuli with a black 'pupil-356 like' feature (Rosa-Salva, Regolin, & Vallortigara, 2012) (see Fig. 2) (see also (Gagliardi, 357 Gallup, & Boren, 1976; Jones, 1980; Scaife, 1976) and (Rosa-Salva, Regolin, & 358 Vallortigara, 2007)). 359



360

Fig. 2 Illustration of stimuli and setups used to test predisposed anti-predator responses in naïve chicks. In the upper panel, a face-like and a non-face-like schematic stimulus are shown, both featuring three internal features with a central pupil-like element. Only in the face-like configuration these resemble a pair of predator eyes. Naïve chicks tend to avoid the face-like stimulus in this test. In the lower panel, prototypical chicks' reactions to looming and sweeping stimuli (fleeing and freezing, respectively) are shown. 367 2.2 Neural and physiological mechanisms underlying early predispositions

368 Our recent research started to shed some light on the neural mechanism underlying early predispositions for social stimuli. We did this visualising c-Fos to map neural 369 activation. In one of our first studies on this regard we found higher activation of IMM 370 in chicks that preferred a scrambled version of a stuffed fowl hen compared to chicks 371 372 that preferred the normal fowl. It was already known that the IMM (Bolhuis & Honey, 1998; Horn, 1986, 1990; Horn & McCabe, 1984; McCabe & Horn, 1994; McCabe, Horn, 373 & Bateson, 1981) is not required for the preference for hen-like objects. Bilateral 374 lesions of the IMM impair the recognition of imprinting objects, but do not suppress 375 the predisposed preference for hen-like objects (Horn & McCabe, 1984). However, 376 although this region does not cause the expression of the predisposition, IMM 377 378 responds differently to naturalistic and artificial stimuli in inexperienced chicks (Mayer et al., 2016). At least three explanations are possible for this finding. The mismatch 379 380 between the template provided by the predisposition for hen-like objects and the 381 artificial object experienced may require increased plasticity to imprint on the scrambled stimulus. This could cause increased c-Fos expression in the chicks that 382 choose the non-predisposed stimulus, since c-Fos is a marker of learning and memory 383 related plasticity (Lanahan & Worley, 1998; Okuno, 2011; Sauvage, Kitsukawa, & 384 385 Atucha, 2019). Another possibility is that the choice to approach the scrambled fowl reflects the lack of a predisposed preference for hen-like objects and that c-Fos 386 expression in IMM could be systematically higher individuals with this abnormal 387 developmental outcome. Last but not least, it has been proposed that the neural 388

389 circuits responsible for the expression of predispositions in chicks with hen preference390 suppress the neuronal activity in the IMM (McCabe, 2019).

391 Other studies have investigated the involvement of the social behaviour network in chicks' predispositions. This network includes interconnected brain regions that are rich 392 in sex-steroid receptors and modulate various social behaviours in adult vertebrates 393 394 (Goodson, 2005; Newman, 1999; O'Connell & Hofmann, 2011). Until recently, however, it was unclear whether areas of this network participate in the expression of early social 395 behaviours in newborn animals too. We found activation of important social behaviour 396 397 network nodes, such as septum, preoptic area and amygdaloid areas (arcopallium and nucleus taeniae of the amygdala) in visually naïve chicks exposed for the first time to 398 social stimuli (Lorenzi et al., 2017; Mayer et al., 2017, 2019; Mayer, Rosa-Salva, & 399 400 Vallortigara, 2017). Among the amygdaloid nuclei, nucleus taeniae of the amygdala and parts of arcopallium selectively responded to the static features of predisposed stimuli 401 402 (e.g., hen-like objects) (Mayer et al., 2019). This is similar to what happens in humans, 403 where amygdala has been implicated in early orienting responses towards face-like configurations (Johnson, 2005). An interesting dissociation appeared when we tested 404 the response to the animate motion of social companions. In this case, the amygdaloid 405 406 nuclei were not responsive, while the activation of septum and preoptic area was increased. In a first study, we found that septum was responsive to the motion of a 407 living conspecific compared to the rigid motion of a similarly stimulus (a stuffed chick 408 409 rotating at a constant speed) (Mayer et al., 2017). In a follow-up work, we found that both septum and preoptic area were also activated by a highly controlled stimulus 410 showing speed changes in the motion of a simple object (Lorenzi et al., 2017). Overall, 411

these nodes of the social behaviour network show selective responses to features of animate objects already in visually naïve animals. Previous visual experience and specific learning events do not seem to be necessary to establish these functions, in line with what has been recently reported for cortical areas specialised for face processing in humans (Ratan Murty et al., 2020). Areas of the social behaviour network might be involved in processing the social valence of, and bonding to, social stimuli in the absence of previous social experience.

In mammals and birds, social behaviours are mediated by neuropeptide signalling 419 based on vasopressin and oxytocin receptors, present in medial amygdala and lateral 420 septum (Goodson et al., 2009). A recent study investigated the role of vasotocin and 421 mesotocin (the avian homologs of vasopressin and oxytocin) in mediating the social 422 423 predisposition for hen-like objects. This revealed that intracranial mesotocin administration increased the level of preference for the stuffed fowl model compared 424 425 to saline-injected controls (Loveland, Stewart, & Vallortigara, 2019). This suggests that 426 mesotocin signalling may regulate the expression of social predispositions, probably acting on areas of the social behaviour network. 427

Intriguingly, in two of our works investigating the physiological substrate of chicks' predispositions for social stimuli, we found that their development can be disrupted exposing embryos to valproic acid during the last week of incubation (Lorenzi et al., 2019; Sgadò et al., 2018). In humans, prenatal exposure to this drug increases the risk to develop disorders of the autistic spectrum (Christensen et al., 2013). For this reason, embryonic exposure to valproic acid is used to develop models of autism in rodents

(Nicolini & Fahnestock, 2018) and chicks (Lorenzi et al., 2019; Nishigori et al., 2013; 434 Sgadò et al., 2018). The embryonic administration of valproic acid to chicks induces 435 selective anomalies in social aggregation behaviours (Nishigori et al., 2013), impairment 436 in the recognition of familiar conspecifics (Zachar et al., 2019), a loss of the 437 438 predisposition for hen-like objects (Sgadò et al., 2018) and speed-changing stimuli (Lorenzi et al., 2019). These results support the idea that, in chicks like in human 439 newborns (Di Giorgio et al., 2016), disturbances of early social predisposition may derail 440 441 the normal development of social cognition, causing abnormal developmental outcomes such as those observed in autistic disorders. The mechanisms subtending to 442 the effects of valproic acid in chicks are currently unknown. However, in mammals, 443 valproic acid seems to act on an inhibitory pathway (GABAergic parvalbumine cells) 444 that controls critical periods for cortical plasticity (eq., Gervain et al., 2013; Gogolla et 445 al., 2009; Lauber, Filice, & Schwaller, 2016; Reh et al., 2020; Silingardi et al., 2010). This 446 opens the way for future studies investigating how valproic acid may affect similar 447 pathways in birds, deepening our understanding of the mechanisms that control 448 sensitive periods in non-mammalian species. 449

450 Overall, evidence suggests that the neural mechanisms to detect living animals are 451 widespread across species and might fulfill a range of adaptive functions from the first 452 stages of life.

453 2.3 Transient time windows for the emergence of early predispositions and their454 hormonal correlates

455 Early predispositions are not fixed and crystallised responses that are present 456 throughout life (differently from the fixed-action patterns investigated by classical 457 ethologists) (see (Shultz, Klin, & Jones, 2018 for a review on human neonates).

The preference for face-like stimuli, for instance, changes during the first months of 458 459 life. This preference is apparent in human neonates in the first hours after birth, declines at around two months of age before reappearing again at five months for 460 more complex stimuli (Buiatti et al., 2019; Johnson et al., 1991; Shultz et al., 2018; 461 Simion & Di Giorgio, 2015) and being detectable up to adulthood (Tomalski, Csibra, & 462 Johnson, 2009). Likewise, at hatching, chicks of both sexes have a spontaneous 463 preference to approach other females (Pallante, Rucco, & Versace, in preparation). This 464 preference rapidly fades when chicks are expose to other chicks. The loss of this 465 466 preference occurs at the age in which in the will chicks leave the nest. This dynamic 467 may be important to promote social cohesion with the flock, that is composed of both male and female chicks. 468

Transient time windows in the appearance of predispositions have been well 469 470 documented for chicks' preferences to approach stuffed hens (Bolhuis, Johnson, & Horn, 1985; Egorova & Anokhin, 2003; Horn, Bolhuis, & Hampton, 1995; Johnson, 471 Bolhuis, & Horn, 1985). This preference emerges only after the animals have been 472 stimulated through some activating experience, such as motoric activity, exposure to 473 474 unrelated visual patterns, handling or acoustical stimulation. These activating experiences do not provide any specific information about the predisposed visual 475 stimuli. Indeed, the activating experiences can even involve modalities other than 476

vision. To have an effect, the activating experiences that trigger the appearance of the 477 predisposition for hen-like objects must happen within a specific time window in the 478 early post-natal life (between 24 and 36 hours after hatching) (Bolhuis & Horn, 1997; 479 Bolhuis et al., 1989; Davies, Johnson, & Horn, 1992; Johnson, Davies, & Horn, 1989). 480 481 Moreover, the ensuing predisposition will be detectable only at precise time points after the activating experiences (between 5 and 24h after the stimulation, Davies et al., 482 1992). This timing, however, is modulated by the type of activating experience to which 483 484 chicks are exposed. For instance, if chicks also receive visual stimulation while performing motoric activity, the preference for hen-like objects is detectable already 485 after 2 hours (Bolhuis et al., 1985). However, it is unclear whether this acceleration 486 depends on the level of arousal or the sensory modality stimulated by the activating 487 experience (see also Lickliter, 2000; Rosa-Salva et al., 2015). Studies on the role of 488 stimulating experiences during sensitive developmental periods also revealed 489 information on the physiological mechanisms involved. An intriguing case is that of 490 491 noradrenaline, which is not directly involved in the expression of the preference for 492 hen-like objects (Bolhuis, McCabe, & Horn, 1986; Davies, Horn, & McCabe, 1985; Davies et al., 1992; see Rosa-Salva et al., 2015 for a review). This neurotransmitter, 493 however, seems to play a role in the opening of the sensitive period related to its 494 emergence. Indeed, the administration of a noradrenaline antagonist delays the onset 495 of the sensitive period during which activating experiences cause the subsequent 496 497 emergence of the predisposition for hen-like objects (Davies et al., 1992; for similar evidence after the administration of the anaesthetic equithisin see Bolhuis & Horn, 498 499 1997).

Similar developmental properties also characterise the predispositions for self-500 501 propelled objects and biological motion. The preference for objects that change in speed is present 24 hours after hatching, but fades two days later (Versace, Ragusa, & 502 503 Vallortigara, 2019). The development of a preference for the semi-rigid biological 504 motion typical of legged animals may require previous exposure to moving visual stimuli (Miura & Matsushima, 2012) or motoric activity such as walking on a treadmill 505 506 (Vallortigara et al., 2005). In the case of biological motion, the behavioural preference is 507 detectable already thirty minutes after the exposure to moving visual stimuli and 508 persists one day late. A sensitive period was also present for the preference for biological motion, which was visible in two- but not in five-day old chicks (Miura et al., 509 510 2020).

In some studies, the development of biological motion preferences were limited to males (Miura & Matsushima, 2012). This is in line with the evidence of the involvement of sex hormones, such as testosterone, in the expression of the predisposed preference for hen-like objects (e.g., Bolhuis et al., 1986; see Rosa-Salva et al., 2015 for a review). This may indicate that at least some of the underlying physiological mechanisms may be shared between different social predispositions.

517 Thyroid hormones too have been implicated in the development of social 518 predispositions for animate motion. These hormones play essential roles in the brain 519 development of vertebrate species (McNabb & King, 1993). Among thyroid hormones, 520 3,5,3'-triiodothyronine (T3) has the highest affinity with avian thyroid receptors and is, 521 therefore, the primary metabolically active thyroid hormone (Bellabarba et al., 1988;

Weirich & McNabb, 1984). We recently obtained preliminary evidence that T3 affects 522 the emergence of chicks' predisposition for speed-changing objects, by modulating its 523 timing (Lorenzi et al., 2020). Blocking the thyroid hormone receptors with an 524 antagonist, at the age when domestics chicks normally show a preference for speed 525 526 changing objects (day 1 post-hatching), leads to the disappearance of the preference. Contrariwise, injecting T3 when the sensitive period normally terminates (day 3 post-527 hatching), reinstates the preference for self-propelled objects, at least in females 528 529 (Lorenzi et al., 2020). Whether T3 only controls the emergence of the predisposition for self-propelled objects or it acts on a more general brain mechanism common to most 530 531 predispositions, remains however to be univocally determined.

532

533 **3. Experience-driven effects and the control of sensitive periods for imprinting**

The predisposed preference for biological motion has facilitating effects on filial 534 imprinting for non-predisposed features, such as the colour of the moving stimulus. 535 Chicks that developed a stronger preference for a biological motion stimulus are better 536 able to imprint on the colour in which this stimulus was depicted (Miura & 537 Matsushima, 2016). Miura and Matsushima (2020) found evidence of a complex 538 interaction between the predisposition for biological motion, the predisposition for the 539 540 red colour (typical of hens' combs and junglefowl faces) and filial imprinting. This work revealed a predisposed preference for approaching and imprinting on stimuli in which 541 the biological motion of a walking hen is associated with the colour red. On the 542 contrary, the same stimulus presented in yellow or performing a simple linear motion is 543

a less effective imprinting stimulus. This might reflect a predisposed preference for a 544 walking animal with a red comb/crest or a reddish face. Notably, this effect seemed 545 driven by the movement of the hen's head, since presenting only the head of the 546 stimulus was as effective as presenting the whole body, whereas the tail region was not 547 548 effective. Crucially, however, the capacity to imprint on biological motion stimuli presented in a yellow colour could be rescued, if chicks were pre-exposed to visual 549 moving stimuli. When chicks had been stimulated by exposure to moving objects 550 551 before imprinting, they could also imprint on a biological motion stimulus with the less predisposed yellow colour. Similarly, previous visual exposure to a group of 552 conspecifics enhances imprinting preferences (Deng & Rogers, 2002b). Only chicks that 553 were exposed to a group of conspecifics, in a later test showed a preference for the 554 individual chick with whom they had been subsequently reared. This effect was due to 555 an improvement of the performance of the right-eye system (left hemisphere), in line 556 with the dominance of the right hemisphere in biological motion processing (Rugani et 557 558 al., 2015) or in familiarity recognition (Vallortigara & Rogers, 2005). On the contrary, the left eye-system showed a stably superior performance, also in the absence of this 559 priming experience. See also (Lickliter, Dyer, & McBride, 1993; Lickliter & Gottlieb, 1985; 560 561 Lickliter & Gottlieb, 1988) for similar effects in ducklings.

562 Overall, the studies reviewed so far suggest a dynamic and potentially enhancing 563 interplay between experience-based and predisposed mechanisms. On the one hand, 564 as we saw in the previous paragraph, non-specific experiences, occurring within specific 565 sensitive periods, drive the emergence of predispositions for animate stimuli. On the

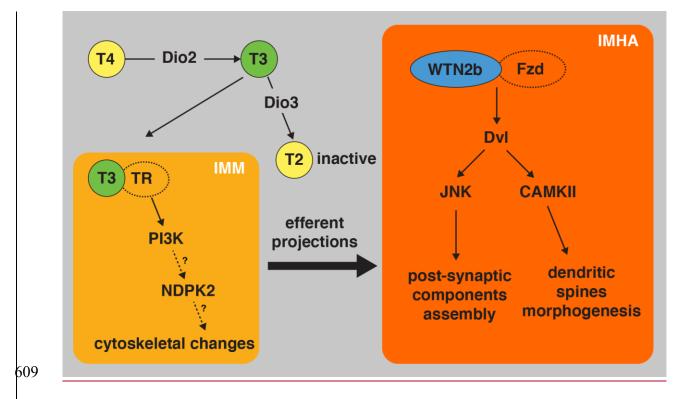
566 other hand, predispositions are associated with enhanced learning of additional 567 features of the stimulus.

A well-known feature of filial imprinting is the presence of a limited sensitive period 568 during which this form of learning can occur. Behaviourally, the sensitive period for 569 imprinting begins as soon as precocial birds start following an object and terminates 570 571 once the birds start to avoid unfamiliar ones (Bolhuis, 1991). The sensitive period for imprinting usually lasts a couple of days after hatching; for instance, Yamaguchi et al. 572 (2012) reported that 4 day-old, dark-reared chicks can no longer be imprinted (but see 573 Case & Graves, 1978; Sluckin, 1972 for reports of imprintability up to 7 days in dark 574 reared chicks, indicating potential inter-breed variability). However, the duration of the 575 sensitive period can substantially vary, being influenced by different factors (Bateson, 576 577 1966; Bolhuis, 1991; Sluckin, 1972). Both pre-hatch and post-hatch experiences influence the determination of the sensitive period (Dimond, 1968; Landsberg, 1976; 578 579 Simner, 1973). For instance, rearing ducklings in diffuse, non-patterned light 580 (preventing structured visual experiences) extends the sensitive period in this species (Moltz & Stettner, 1961). 581

At the neurophysiological level, an increase of the concentration of the neurotransmitter glutamate extends the imprinting period, suggesting a possible role of NMDA glutamate receptors (Parsons & Rogers, 1997, 2000). The sensitive period for filial imprinting may be influenced by hormones too (Bateson, 1983; Knudsen, 2004). For instance, in ducklings a rise of corticosterone has been described around the end of the sensitive period for imprinting (Weiss, Köhler, & Landsberg, 1977). This

hormonal change may be the source for the increasing fear observed when imprinting
 terminates (Bolhuis, 1991).

590 Recently, a role of thyroid hormones has been identified in domestic chicks. In 591 precocial galliform birds, the thyroid hormones T4 and T3 peak around hatching and then gradually decrease (Lu, McMurtry, & Coon, 2007; McNabb, 2006; Yamaguchi et al., 592 593 2012). The level of T3 in the brain correlates with the strength of imprinting and 594 artificially injecting T3 facilitates it. The action of T3 appears to be quick, being detectable already in 30 minutes, similarly to what reported above for the induction of 595 596 the predisposition for biological motion. Importantly, T3 is strongly implicated in the control of the sensitive period for imprinting in chicks. Inhibiting it (either systemically 597 or via brain injections) impairs visual imprinting, whereas injecting it extends and even 598 599 re-opens its sensitive period up to 8 days after hatching. After this age, the capability to form imprinting attachment cannot be rescued by T3, indicating the presence of 600 601 further, yet unknown, closing mechanisms (Yamaguchi et al., 2012). Recent studies have 602 also elucidated the neural mechanisms subtending to the action of T3, which seem to be specifically localised to IMM (a region classically implicated in imprinting, Horn, 603 2004) (summarised in Fig. 3). Blockage of the nucleotide diphosphate kinase 2 in IMM 604 impairs the reopening of the sensitive window by T3 (Yamaguchi et al., 2016). Likewise, 605 blocking the Wnt-2b glycoprotein, related to neuronal growth, in an area connected to 606 IMM (IMHA, intermediate medial hyperpallium apicale) also prevents T3 action (Aoki et 607 608 al., 2015; Yamaguchi et al., 2018) (see Fig. 3).



610 Fig. 3. Known pathways for the action of T3 on IMM (Intermediate Medial Mesopallium) and its efferent 611 IMHA (Intermediate Medial Hyperpallium Apicale). The concentration of the thyroid hormone T3 (3,5,3'-612 triiodothyronine) is balanced in the brain by the enzyme Dio2 (type 2 iodothyronine deiodinase), which 613 converts T4 into T3 (the active form) and by Dio3, which converts T3 into its inactive metabolite T2 (3,3'-614 diiodothyronine). In IMM, T3 binds a thyroid hormone receptor (TR), which activates the enzyme 615 phosphatidylinositol 3'-kinase (PI3K). PI3K is hypothesised (dotted arrow) to act on another enzyme, 616 NDPK2 (Nucleoside diphosphate kinase II), which would in turn interact with proteins that regulate the 617 actin cytoskeleton, potentially leading to changes in the dendritic spines, supporting learning related 618 plasticity. In IMHA, the molecular mechanisms that regulate the sensitive period for imprinting may 619 involve the Wnt2b protein, which binds to a Fzd (frizzled) receptor, causing the activation of Dvl 620 (Dishvelled) protein. This in turn modulates the activity of two enzymes, JNK (c-Jun N-terminal kinase) 621 and CAMKII (Ca2+/calmodulin-dependent protein kinase II). The activation of this pathway induces 622 changes in actin and reorganization of the microtubules, modulating the morphogenesis of dendritic 623 spines and post-synaptic components assembly. Overall, the activation of the Wtn pathway in IMHA, 624 concurrently with the input that this structure receives from the IMM that has been activated by T3

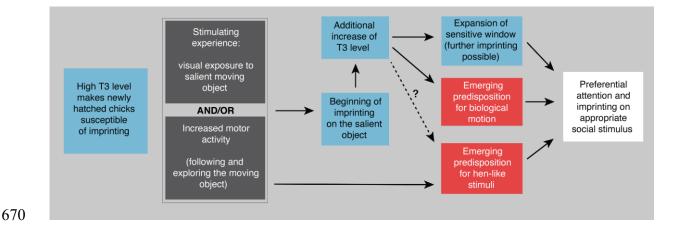
625 influx, may strengthen the connectivity between these two areas, contributing to the opening of the 626 critical period for imprinting.

627

Moreover, it has been recently shown that the expression of GABA_A-GABA_B receptors 628 within IMM defines the opening and closing of the sensitive period downstream to T3. 629 While GABA_B initiates imprinting, GABA_A contributes to its termination (Aoki et al., 2018; 630 see also McCabe, 2019 for a comprehensive review of this literature). In mammals, the 631 632 brain-region specific maturation parvalbumin-positive GABAergic inhibitory interneurons have a crucial role in the neurobiology of sensitive periods. This allows 633 634 the refinement of cortical circuits that build up advanced cognitive functions (reviewed 635 in Reh et al., 2020). Once again, similar mechanisms might be present in avian species (e.g., Aoki et al., 2018, see above), opening the way for exciting comparative 636 investigations. 637

Similar to what described for the behavioural effects of the biological motion 638 639 preference, the relationship between imprinting and T3 hormonal levels seems to be a circular one. The process of imprinting increases the forebrain concentration of T3 640 641 hormone, by increasing the expression of Dio2, the enzyme that converts T4 to T3 (the active form) (see also Takemura et al., 2018). This "primes" the mechanisms for the 642 development of further secondary imprinting learning. As a consequence, after this 643 644 initial priming even chicks as old as 4-8 days became susceptible to further imprinting (Yamaguchi et al., 2012). The similarity with the mechanisms described above for the 645 646 action of the biological motion preference is not only a superficial one. Indeed, the

induction of the predisposition for biological motion is linked to an increase in the 647 648 gene expression of Dio2 (Miura et al., 2018; Takemura et al., 2018). The level of preference for biological motion developed by chicks primed with exposure to moving 649 650 stimuli correlates with the level of telencephalic Dio2 expression (Takemura et al., 651 2018). Moreover, injections of IOP (iopanoic acid, that inhibits Dio2) reduced the level of biological motion preference developed by newly-hatched chicks after priming with 652 653 motion stimuli (Miura et al., 2018). This reveals a direct causal link between the action 654 of thyroid hormones and predispositions for biological motion. According to the model proposed by Miura and Matsushima (2020), the first encounter of an animal with a 655 salient moving object would cause a surge of telencephalic T3 concentration. This 656 would have consequences both at the level of inborn predispositions (causing the 657 rapid emergence of the predisposition for biological motion), and at the level of 658 learning mechanisms (causing the acute activation of imprinting learning and priming 659 the memory system for further imprinting learning in the later days). Thanks to the 660 661 induction of the predisposition for biological motion, even if the first moving object seen by the chick would be an inanimate object, the subsequent imprinting would be 662 directed towards more biologically plausible objects. Please note that secondary 663 664 imprinting towards naturalistic stimuli overrides previous imprinting learning on artificial ones, but not vice versa (Boakes & Panter, 1985; Bolhuis & Trooster, 1988). 665 Moreover, the first "wave" of imprinting will determine at least some degree of 666 667 following behaviour (i.e., motoric and visual stimulation). Thus, this first visual experience with any moving object is also likely to activate the emergence of the 668 669 predispositions for hen-like objects, and specifically for their red face region (Fig. 4).



671 Fig. 4. Schematic illustration of a model representing the emergence of multiple social predispositions as 672 a consequence of the exposure to any salient moving stimulus, in newly hatched chicks. T3 level peak at 673 hatching, making these animals susceptible to imprinting. The exposure to the stimulus and the 674 associated motoric activity will cause the emergence of the predisposition for the head region of hen-675 like stimuli. Moreover, by being exposed to a salient stimulus the animals will start to learn its features 676 through filial imprinting. This process will cause an additional increase in the concentration of T3. This 677 will have two consequences: extending the sensitive window for imprinting and causing the emergence 678 of the predisposition for biological motion. (It is unknown whether T3 has any effect on the 679 predisposition for hen-like object, dotted arrow). The combined presence of the two predispositions will 680 direct chicks' attention towards appropriate social stimuli (e.g., the mother hen) if they are at all available 681 in the environment. Since the animals are still susceptible to (secondary) imprinting learning, this will 682 ensure filial attachment towards the mother hen.

It seems that multiple mechanisms evolved to ensure this crucial step of social development (Versace et al., 2018). Filial attachment towards the mother hen has important short-term beneficial consequences for the chicks, that gain heat, protection and guidance from the hen. Thanks to their notable social-learning abilities, chicks can also acquire crucial information (e.g., on the position and palatability of various food sources) from the hen and other brood-mates (e.g., Marino, 2017; Nicol, 2004).

Unfortunately, compared to the earlier developmental stages, less is known about how 689 690 these interactive processes extend to later stages of chicks' life. For instance, there is relatively little evidence of the impact of these early environmental influences on the 691 692 social adaptation of adult chickens (but see Rogers & Workman, 1989 for longitudinal 693 data on lateralization and social interaction in chicks up to 16 days of age; see also Leonard et al., 1993; Leonard, Zanette, & Wayne Fairfull, 1993; Widowski, Lo Fo Wong, 694 & Duncan, 1998 for the long-term effect of group composition on adult mating and 695 696 agonistic behaviours). This aspect has been perhaps better studied in other bird species such as zebra finches, where sexual imprinting has been the object of 697 numerous studies (see Bischof, 1994, 2018; ten Cate & Vos, 1999 for reviews). In sexual 698 imprinting, the early experience the birds have with social companions, within a specific 699 700 sensitive period of the post-natal life, affects their mate choice later in life. After this 701 early acquisition phase, the formation of stable mate-preferences by sexual imprinting involves a second sensitive period, at the time when the young bird performs its first 702 703 courtship. The availability of an appropriate partner to court can modify or consolidate 704 the preference acquired in the earlier phase, crystallising it in its definitive form. It has been proposed that the first stage of sexual imprinting in zebra finches could 705 706 correspond to filial imprinting as described for altricial birds. In this phase, the young bird would acquire general information that allow it to recognise parents, siblings and 707 708 other members of its social group. During the subsequent consolidation phase, its first 709 courtship attempts are directed towards individuals resembling this template, if at all 710 available, and the tendency is further consolidated. Similar to what we saw for filial 711 imprinting, also in sexual imprinting the preferences of young birds are not completely

unconstrained, revealing the presence of genetic biases or unlearned predispositions. 712 713 Indeed, cross-fostering experiments with birds of different species revealed that sexual imprinting is usually easier to achieve for the own species. This two-phase process, 714 715 involving the interaction of predisposed and learned mechanisms, thus increases the 716 chances to obtain sexual imprinting towards the bird's own species, while allowing still sufficient behavioural flexibility to cope with different environmental circumstances 717 718 (Bischof, 2018). The neural correlates of learning occurring in sexual imprinting during 719 the consolidation phase have been also well investigated (Bischof, 2007). In the case of visual sexual imprinting, changes occurring during the second sensitive period involved 720 mostly an irreversible reduction of dendritic spines in the Lateral Nido-Mesopallium 721 (LNM). Similarities with the processes observed for filial imprinting include a clear 722 involvement of hormonal signalling (testosterone, in this case) in the opening of the 723 sensitive window for the consolidation phase, and of GABAergic inhibition for its 724 closing. Future studies should be devoted to compare how predispositions and 725 726 learning mechanisms interact during the earlier stages of life and later in development (see Vidal, 1980 for a behavioural study of the relationship between sexual and filial 727 in chicks). It would thus be important to 728 imprinting perform longitudinal 729 developmental studies, targeting how early environmental influences shape adult social behaviour. 730

731 **4.** Conclusions

In a world rich in stimuli, social predispositions help young animals to direct theirattention and learning toward social partners, which provide protection and guidance,

and to ignore less relevant stimuli. Predispositions are not fixed and immutable mechanisms, though, but respond in different ways to the environmental and social stimulation present at specific time-points during the ontogenetic development. We have described how the relation between predisposed and environmental factors in the first phases of life has profound influences on subsequent development (see also Versace et al., 2018; Versace & Vallortigara, 2015). However, long term effects have just started to be elucidated.

We have shown how distinct sensitive periods shape the development of social 741 behaviour via complex interactions of environmental and genetic influences. This is 742 apparent, for instance, in the development of affiliative responses for the mother 743 through predispositions that orient the newborn towards animate objects (reviewed in 744 745 Di Giorgio et al., 2017a) and enhance learning of the particular features of the social partners. The latter mechanism is shown for example by the enhancement of 746 747 imprinting in chicks whose predispositions for biological motion have been activated 748 by previous exposure to moving objects (Miura & Matsushima, 2016; Miura et al., 2020). Importantly, we have shown how environmental events can modulate the timing 749 and duration of sensitive periods, maximising the chances of optimal developmental 750 outcomes. This is well-exemplified by in the surge of T3 elicited by the first wave of 751 imprinting learning after encountering a salient object, which allows subsequent 752 753 secondary imprinting to take place (Yamaguchi et al., 2012).

Domestic chicks are currently an elective model system to investigate connectionbetween predispositions and learning at the behavioural and neurobiological level. This

is mostly due to the advantages of precocial animals that respond to behavioural tests 756 immediately after birth, as well as to the opportunity to investigate the fast learning 757 mechanism of filial imprinting. These advantages allowed researchers to shed some 758 759 light on the neural and physiological bases of the early mechanisms (e.g., Horn, 2004; 760 Lorenzi et al., 2017; Loveland et al., 2019; Mayer et al., 2016, 2017a, 2017b, 2019), including those for the opening and closing of sensitive periods (Aoki et al., 2018; 761 762 Yamaguchi et al., 2012). While the notion of sensitive period has been traditionally 763 applied to filial imprinting, mounting evidence shows that specific time windows regulate the emergence of predispositions too (e.g., (Bolhuis & Horn, 1997; Bolhuis et 764 al., 1989; Davies, Johnson, & Horn, 1992; Johnson, Davies & Horn, 1989; Miura et al., 765 766 2020; Versace, Ragusa & Vallortigara, 2019).

767 More recently, chicks have been used for the study of how predispositions influence typical and pathological development, as they are being proposed as a model for the 768 social impairment of autism spectrum disorders (Sgadò et al., 2018; Lorenzi al., 2019; 769 770 see also Di Giorgio et al., 2016; Zachar et al., 2019). This is facilitated by the fact that 771 predispositions appear to be very similar across taxa. In fact, organisms as different as human babies, chicks and tortoises display similar social predispositions (e.g., Johnson, 772 2005; Rosa-Salva et al., 2010; Versace et al., 2020). This further points at the adaptive 773 value of mechanisms that enable young animals to cope with their environment from 774 775 the early stages of life, as already envisioned in the pre-representations conceptualised 776 by Mehler and Dupoux for human infants (Mehler & Dupoux, 1994).

777

779 Acknowledgements

This work was supported by a grant from the European Research Council under the European Union's Seventh Framework Programme (FP7/2007-2013) Grant ERC-2011-ADG_20110406, Project No: 461 295517, PREMESOR, by Fondazione Caritro Grant Biomarker DSA [40102839] and PRIN 2015 (Neural bases of animacy detection, and their relevance to the typical and atypical development of the brain) to G.V.

- 786 References
- Aoki, N., Yamaguchi, S., Fujita, T., Mori, C., Fujita, E., Matsushima, T., & Homma, K. J.
- 788 (2018). GABA-A and GABA-B receptors in filial imprinting linked with opening and
- closing of the sensitive period in domestic chicks (Gallus gallus domesticus).
- 790 Frontiers in Physiology, 9. https://doi.org/10.3389/fphys.2018.01837
- 791 Aoki, N., Yamaguchi, S., Kitajima, T., Takehara, A., Katagiri-Nakagawa, S., Matsui, R.,
- 792 Watanabe, D., Matsushima, T., & Homma, K. J. (2015). Critical role of the neural
- pathway from the intermediate medial mesopallium to the intermediate
- hyperpallium apicale in filial imprinting of domestic chicks (Gallus gallus
- 795 domesticus). *Neuroscience*, *308*, 115–124.
- 796 https://doi.org/10.1016/j.neuroscience.2015.09.014
- 797 Bateson, P. P. G. (1966). The characteristics and context of imprinting. *Biological*

- 798 *Reviews*, *41*(2), 177–217. https://doi.org/10.1111/j.1469-185X.1966.tb01489.x
- Bateson, P. P. G. (1983). Sensitive periods in behavioural development. *Archives of Disease in Childhood*, *58*(2), 85–86. https://doi.org/10.1136/adc.58.2.85
- 801 Bateson, P. P. G., & Gluckman, P. (2012). Plasticity and robustness in development and
- evolution. *International Journal of Epidemiology*, *41*(1), 219–223.
- 803 https://doi.org/10.1093/ije/dyr240
- 804 Bellabarba, D., Belisle, S., Gallo-Payet, N., & Lehoux, J.-G. (1988). Mechanism of action
- 805 of thyroid hormones during chick embryogensis. American Zoologist, 28(2), 389-
- 806 399. https://doi.org/10.1093/icb/28.2.389
- Bischof, H.-J. (1994). Sexual imprinting as a two stage process. In *Causal Mechanisms of Behavioural development* (pp. 82–87). Cambridge University Press.
- 809 Bischof, H.-J. (2007). Behavioral and neuronal aspects of developmental sensitive
- 810 periods. *NeuroReport*, *18*(5), 461–465.
- 811 https://doi.org/10.1097/WNR.0b013e328014204e
- 812 Bischof, H.-J. (2018). Sexual imprinting. In *Encyclopedia of Reproduction* (Second Edi,
- 813 pp. 267–271). Academic Press.
- 814 Boakes, R., & Panter, D. (1985). Secondary imprinting in the domestic chick blocked by
- previous exposure to a live hen. *Animal Behaviour*, *33*(2), 353–365.
- 816 https://doi.org/10.1016/S0003-3472(85)80059-2
- Bodin, D., Yeates, K. O., & Cass, J. (2011). Sensitive periods. In *Encyclopedia of Clinical*

818	Neuropsychology (pp. 2255–225	56). New York, NY: Springe	er New York.
-----	-------------------------------	----------------------------	--------------

819 https://doi.org/10.1007/978-0-387-79948-3_1593

- Bolhuis, J. J. (1991). Mechanisms of avian imprinting: a review. *Biological Reviews*, *66*(4),
 303–345. https://doi.org/10.1111/j.1469-185X.1991.tb01145.x
- 822 Bolhuis, J. J., & Honey, R. C. (1998). Imprinting, learning and development: from
- behaviour to brain and back. *Trends in Neurosciences, 21*(7), 306–311.
- 824 https://doi.org/10.1016/S0166-2236(98)01258-2
- 825 Bolhuis, J. J., & Horn, G. (1997). Delayed induction of a filial predisposition in the chick

after anaesthesia. *Physiology & Behavior, 62*(6), 1235–1239.

827 https://doi.org/10.1016/S0031-9384(97)00231-X

828 Bolhuis, J. J., Johnson, M. H., & Horn, G. (1985). Effects of early experience on the

829 development of filial preferences in the domestic chick. *Developmental*

830 *Psychobiology*, *18*(4), 299–308. https://doi.org/10.1002/dev.420180403

- 831 Bolhuis, J. J., Johnson, M. H., & Horn, G. (1989). Interacting mechanisms during the
- formation of filial preferences: The development of a predisposition does not
- 833 prevent learning. Journal of Experimental Psychology: Animal Behavior Processes,
- 834 *15*(4), 376–382. https://doi.org/10.1037/0097-7403.15.4.376
- 835 Bolhuis, J. J., McCabe, B. J., & Horn, G. (1986). Androgens and imprinting: differential
- 836 effects of testosterone on filial preference in the domestic chick. *Behavioral*
- 837 Neuroscience, 100(1), 51–56. https://doi.org/10.1037/0735-7044.100.1.51

838	Bolhuis, J. J., & Trooster, W. J. (1988). Reversibility revisited: stimulus-dependent
839	stability of filial preference in the chick. Animal Behaviour, 36(3), 668–674.
840	https://doi.org/10.1016/S0003-3472(88)80149-0

- 841 Buiatti, M., Di Giorgio, E., Piazza, M., Polloni, C., Menna, G., Taddei, F., Baldo, E., &
- Vallortigara, G. (2019). Cortical route for facelike pattern processing in human
- newborns. *Proceedings of the National Academy of Sciences, 116*(10), 4625–4630.
- 844 https://doi.org/10.1073/pnas.1812419116
- 845 Burghardt, G. M., & Bowers, R. I. (2017). From instinct to behavior systems: an
- 846 integrated approach to ethological psychology. In APA handbook of comparative
- 847 psychology: Basic concepts, methods, neural substrate, and behavior. (pp. 333–
- 848 364). Washington: American Psychological Association.
- 849 https://doi.org/10.1037/0000011-017
- 850 Case, V. J., & Graves, H. B. (1978). Functional versus other types of imprinting and
- sensitive periods in Gallus chicks. *Behavioral Biology*, *23*(4), 433–445.
- 852 https://doi.org/10.1016/S0091-6773(78)91495-5
- 853 Cate, C. T. (1986). Does behavior contingent stimulus movement enhance filial
- imprinting in Japanese quail? *Developmental Psychobiology*, *19*(6), 607–614.
- 855 https://doi.org/10.1002/dev.420190611
- 856 Chan, K. P. (2014). Prenatal meditation influences infant behaviors. *Infant Behavior and*
- 857 Development, 37(4), 556–561. https://doi.org/10.1016/j.infbeh.2014.06.011
- 858 Charil, A., Laplante, D. P., Vaillancourt, C., & King, S. (2010). Prenatal stress and brain

859	devel	opment.	Brain	Research	n Reviews,	<i>65</i> (1),	56–79.
-----	-------	---------	-------	----------	------------	----------------	--------

- 860 https://doi.org/10.1016/j.brainresrev.2010.06.002
- Chiandetti, C. (2011). Pseudoneglect and embryonic light stimulation in the avian brain. *Behavioral Neuroscience*, *125*(5), 775–782. https://doi.org/10.1037/a0024721
- 863 Chiandetti, C., Galliussi, J., Andrew, R. J., & Vallortigara, G. (2013). Early-light embryonic
- stimulation suggests a second route, via gene activation, to cerebral lateralization
- in vertebrates. *Scientific Reports, 3*(1), 2701. https://doi.org/10.1038/srep02701
- 866 Chiandetti, C., & Vallortigara, G. (2019). Distinct effect of early and late embryonic
- 867 light-stimulation on chicks' lateralization. *Neuroscience*, 414, 1–7.
- 868 https://doi.org/10.1016/j.neuroscience.2019.06.036
- 869 Christensen, J., Grønborg, T. K., Sørensen, M. J., Schendel, D., Parner, E. T., Pedersen, L.
- 870 H., & Vestergaard, M. (2013). Prenatal valproate exposure and risk of autism
- spectrum disorders and childhood autism. *JAMA*, *309*(16), 1696.
- 872 https://doi.org/10.1001/jama.2013.2270
- 873 Davies, D. C., Horn, G., & McCabe, B. J. (1985). Noradrenaline and learning: effects of
- 874 the noradrenergic neurotoxin DSP4 on imprinting in the domestic chick. *Behavioral*
- 875 *Neuroscience*, *99*(4), 652–660. https://doi.org/10.1037/0735-7044.99.4.652
- 876 Davies, D. C., Johnson, M. H., & Horn, G. (1992). The effect of the neurotoxin DSP4 on
- the development of a predisposition in the domestic chick. *Developmental*
- 878 *Psychobiology*, *25*(4), 251–259. https://doi.org/10.1002/dev.420250403

- Dehorter, N., & Del Pino, I. (2020). Shifting developmental trajectories during critical
 periods of brain formation. *Frontiers in Cellular Neuroscience*, *14*.
- 881 https://doi.org/10.3389/fncel.2020.00283
- 882 Deng, C., & Rogers, L. J. J. (1997). Differential contributions of the two visual pathways
- to functional lateralization in chicks. *Behavioural Brain Research*, *87*(2), 173–182.
- 884 https://doi.org/10.1016/S0166-4328(97)02276-6
- 885 Deng, C., & Rogers, L. J. J. (2002a). Factors affecting the development of lateralization
- in chicks. In *Comparative vertebrate lateralization*. Cambridge University Press.
- 887 Deng, C., & Rogers, L. J. J. (2002b). Social recognition and approach in the chick:
- lateralization and effect of visual experience. *Animal Behaviour, 63*(4), 697–706.
- 889 https://doi.org/10.1006/anbe.2001.1942
- 890 Dessborn, L., Englund, G., Elmberg, J., & Arzél, C. (2012). Innate responses of mallard
- 891 ducklings towards aerial, aquatic and terrestrial predators. *Behaviour*, *149*(13–14),
- 892 1299–1317. https://doi.org/10.1163/1568539X-00003014
- 893 Dharmaretnam, M., & Rogers, L. J. J. (2005). Hemispheric specialization and dual
- 894 processing in strongly versus weakly lateralized chicks. *Behavioural Brain Research*,
- 895 *162*(1), 62–70. https://doi.org/10.1016/j.bbr.2005.03.012
- B96 Di Giorgio, E., Frasnelli, E., Rosa-Salva, O., Luisa Scattoni, M., Puopolo, M., Tosoni, D.,
- 897 Simion, F., & Vallortigara, G. (2016). Difference in visual social predispositions
- between newborns at low- and high-risk for autism. *Scientific Reports, 6*(1), 26395.
- 899 https://doi.org/10.1038/srep26395

900	Di Giorgio, E.,	Loveland, J. L., I	Mayer, U.,	Rosa-Salva, O.,	Versace, E., &	Vallortigara, G

- 901 (2017). Filial responses as predisposed and learned preferences: Early attachment in
- 902 chicks and babies. *Behavioural Brain Research*, *325*, 90–104.
- 903 https://doi.org/10.1016/j.bbr.2016.09.018
- 904 Di Giorgio, E., Lunghi, M., Simion, F., & Vallortigara, G. (2017). Visual cues of motion
- 905 that trigger animacy perception at birth: the case of self-propulsion.

906 Developmental Science, 20(4), e12394. https://doi.org/10.1111/desc.12394

- 907 Dimond, S. J. (1968). Effects of photic stimulation before hatching on the development
- 908 of fear in chicks. Journal of Comparative and Physiological Psychology, 65(2), 320-
- 909 324. https://doi.org/10.1037/h0025550
- 910 Egorova, O. V., & Anokhin, K. V. (2003). Experimental analysis of the processes of
- 911 systems genesis: expression of the c-fos gene in the chick brain during treatments
- 912 inducing the development of the species-specific results-of-action acceptor.
- 913 *Neuroscience and Behavioral Physiology*, *33*(3), 209–216.
- 914 https://doi.org/10.1023/A:1022186911789
- 915 Eibl-Eibesfeldt, I. (1975). Ethology, the biology of behavior. New York: Holt, Rinehart,
- 916 Winston.
- 917 Frankenhuis, W. E., House, B., Clark Barrett, H., & Johnson, S. P. (2013). Infants'
- 918 perception of chasing. *Cognition*, *126*(2), 224–233.
- 919 https://doi.org/10.1016/j.cognition.2012.10.001
- 920 Gagliardi, G. J., Gallup, G. G., & Boren, J. L. (1976). Effect of different pupil to eye size

- 921 ratios on tonic immobility in chickens. *Bulletin of the Psychonomic Society*, 8(1),
- 922 58–60. https://doi.org/10.3758/BF03337075
- 923 Gervain, J., Vines, B. W., Chen, L. M., Seo, R. J., Hensch, T. K., Werker, J. F., & Young, A.
- 924 H. (2013). Valproate reopens critical-period learning of absolute pitch. *Frontiers in*
- 925 Systems Neuroscience, 7. https://doi.org/10.3389/fnsys.2013.00102
- 926 Gogolla, N., LeBlanc, J. J., Quast, K. B., Südhof, T. C., Fagiolini, M., & Hensch, T. K.
- 927 (2009). Common circuit defect of excitatory-inhibitory balance in mouse models of
- 928 autism. Journal of Neurodevelopmental Disorders, 1(2), 172–181.
- 929 https://doi.org/10.1007/s11689-009-9023-x
- Goodson, J. L. (2005). The vertebrate social behavior network: evolutionary themes and
 variations. *Hormones and Behavior*, *48*(1), 11–22.
- 932 https://doi.org/10.1016/j.yhbeh.2005.02.003
- 933 Goodson, J. L., Schrock, S. E., Klatt, J. D., Kabelik, D., & Kingsbury, M. A. (2009).
- 934 Mesotocin and nonapeptide receptors promote estrildid flocking behavior. Science,
- 935 *325*(5942), 862–866. https://doi.org/10.1126/science.1174929
- 936 Goren, C. C., Sarty, M., & Wu, P. Y. K. (1975). Visual following and pattern discrimination
- 937 of face-like stimuli by newborn infants. *Pediatrics*, *56*(4), 544–549.
- 938 Hébert, M., Versace, E., & Vallortigara, G. (2019). Inexperienced preys know when to
- 939 flee or to freeze in front of a threat. *Proceedings of the National Academy of*
- 940 Sciences, 116(46), 22918–22920. https://doi.org/10.1073/pnas.1915504116

- 941 Hernik, M., Fearon, P., & Csibra, G. (2014). Action anticipation in human infants reveals
- 942 assumptions about anteroposterior body-structure and action. *Proceedings of the*
- 943 *Royal Society B: Biological Sciences, 281*(1781), 20133205.
- 944 https://doi.org/10.1098/rspb.2013.3205
- Hess, E. H., & Goodwin, E. B. (1969). Innate visual form preferences in the imprinting
 behavior of hatchling chicks. *Behaviour*, *34*(4), 238–254.
- 947 https://doi.org/10.1163/156853969X00143
- 948 Horn, G. (1986). Imprinting, learning, and memory. *Behavioral Neuroscience*, 100(6),
- 949 825–832. https://doi.org/10.1037/0735-7044.100.6.825
- 950 Horn, G. (1990). Neural bases of recognition memory investigated through an analysis
- 951 of imprinting. *Philosophical Transactions of the Royal Society of London. Series B:*
- 952 Biological Sciences, 329(1253), 133–142. https://doi.org/10.1098/rstb.1990.0158
- 953 Horn, G. (2004). Pathways of the past: the imprint of memory. *Nature Reviews*
- 954 *Neuroscience*, *5*(2), 108–120. https://doi.org/10.1038/nrn1324
- Horn, G., Bolhuis, J. J., & Hampton, N. G. (1995). Induction and development of a filial
- predisposition in the chick. *Behaviour*, *132*(5–6), 451–477.
- 957 https://doi.org/10.1163/156853995X00667
- 958 Horn, G., & McCabe, B. J. (1984). Predispositions and preferences. Effects on imprinting
- of lesions to the chick brain. *Animal Behaviour*, *32*(1), 288–292.
- 960 https://doi.org/10.1016/S0003-3472(84)80349-8

961	Hubel, D. H., & Wiesel, T. N. (1970). The period of susceptibility to the physiological
962	effects of unilateral eye closure in kittens. The Journal of Physiology, 206(2), 419–
963	436. https://doi.org/10.1113/jphysiol.1970.sp009022
964	Johnson, M. H. (2005). Subcortical face processing. Nature Reviews Neuroscience, 6(10),
965	766–774. https://doi.org/10.1038/nrn1766
966	Johnson, M. H., Bolhuis, J. J., & Horn, G. (1985). Interaction between acquired
967	preferences and developing predispositions during imprinting. Animal Behaviour,
968	<i>33</i> (3), 1000–1006. https://doi.org/10.1016/S0003-3472(85)80034-8
969	Johnson, M. H., Davies, D. C., & Horn, G. (1989). A sensitive period for the development
970	of a predisposition in dark-reared chicks. Animal Behaviour, 37, 1044–1046.
971	https://doi.org/10.1016/0003-3472(89)90148-6
972	Johnson, M. H., Dziurawiec, S., Ellis, H., & Morton, J. (1991). Newborns' preferential
973	tracking of face-like stimuli and its subsequent decline. <i>Cognition, 40</i> (1–2), 1–19.
974	https://doi.org/10.1016/0010-0277(91)90045-6
975	Johnson, M. H., & Horn, G. (1988). Development of filial preferences in dark-reared
976	chicks. <i>Animal Behaviour, 36</i> (3), 675–683. https://doi.org/10.1016/S0003-
977	3472(88)80150-7
978	Johnson, M. H., & Morton, J. (1991). <i>Biology and cognitive development: the case of</i>
979	face recognition. Early Development and Parenting. Oxford: Wiley Blackwell.
980	Retrieved from http://doi.wiley.com/10.1002/edp.2430020408

981	Jones, R. B. (1980). Reactions of male domestic chicks to two-dimensional eye-like
982	shapes. Animal Behaviour, 28(1), 212–218. https://doi.org/10.1016/S0003-
983	3472(80)80025-X

- 984 Kalcher-Sommersguter, E., Preuschoft, S., Franz-Schaider, C., Hemelrijk, C. K., Crailsheim,
- 985 K., & Massen, J. J. M. (2015). Early maternal loss affects social integration of
- 986 chimpanzees throughout their lifetime. *Scientific Reports, 5*(1), 16439.
- 987 https://doi.org/10.1038/srep16439
- 988 Knudsen, E. I. (2004). Sensitive periods in the development of the brain and behavior.
- 989 Journal of Cognitive Neuroscience, 16(8), 1412–1425.
- 990 https://doi.org/10.1162/0898929042304796
- 991 Kovach, J. K. (1971). Effectiveness of different colors in the elicitation and development
- 992 of approach behavior in chicks. *Behaviour*, *38*(1–2), 154–168.
- 993 https://doi.org/10.1163/156853971X00069
- Lanahan, A., & Worley, P. (1998). Immediate-early genes and synaptic function.
- 995 *Neurobiology of Learning and Memory, 70*(1–2), 37–43.
- 996 https://doi.org/10.1006/nlme.1998.3836
- 997 Landsberg, J.-W. (1976). Posthatch age and developmental age as a baseline for
- 998 determination of the sensitive period for imprinting. *Journal of Comparative and*
- 999 *Physiological Psychology*, *90*(1), 47–52. https://doi.org/10.1037/h0077253
- 1000 Lauber, E., Filice, F., & Schwaller, B. (2016). Prenatal palproate exposure differentially
- 1001 affects parvalbumin-expressing neurons and related Circuits in the cortex and

- 1002 striatum of mice. *Frontiers in Molecular Neuroscience, 9*.
- 1003 https://doi.org/10.3389/fnmol.2016.00150
- 1004 Lemaire, B. S., Rucco, D., Josserand, M., Vallortigara, G., & Versace, E. (2020). Stability
- and individual variability of social attachment in imprinting. *BioRxiv*.
- 1006 https://doi.org/10.1101/2020.04.04.025072.
- Leonard, M. L., Zanette, L., Thompson, B. K., & Wayne Fairfull, R. (1993). Early exposure
 to the opposite sex affects mating behaviour in White Leghorn chickens. *Applied*
- 1009 Animal Behaviour Science, 37(1), 57–67. https://doi.org/10.1016/0168-
- 1010 1591(93)90070-6
- 1011 Leonard, M. L., Zanette, L., & Wayne Fairfull, R. (1993). Early exposure to females affects
- 1012 interactions between male White Leghorn chickens. *Applied Animal Behaviour*
- 1013 Science, 36(1), 29–38. https://doi.org/10.1016/0168-1591(93)90096-8
- 1014 Lickliter, R. (2000). An ecological approach to behavioral development: insights from
- 1015 comparative psychology. *Ecological Psychology*, *12*(4), 319–334.
- 1016 https://doi.org/10.1207/S15326969ECO1204_06
- 1017 Lickliter, R., Dyer, A. B., & McBride, T. (1993). Perceptual consequences of early social
- 1018 experience in precocial birds. *Behavioural Processes*, *30*(3), 185–200.
- 1019 https://doi.org/10.1016/0376-6357(93)90132-B
- 1020 Lickliter, R., & Gottlieb, G. (1985). Social interaction with siblings is necessary for visual
- 1021 imprinting of species-specific maternal preferences in ducklings (Anas
- 1022 platyrhynchos). *Journal of Comparative Psychology*, *99*(4), 371–379.

- 1023 https://doi.org/10.1037/0735-7036.99.4.371
- 1024 Lickliter, R., & Gottlieb, G. (1988). Social specificity: interaction with own species is
- 1025 necessary to foster species-specific maternal preference in ducklings.
- 1026 Developmental Psychobiology, 21(4), 311–321.
- 1027 https://doi.org/10.1002/dev.420210403
- Lorenz, K. Z. (1937). The companion in the bird's world. *The Auk, 54*(3), 245–273.
 https://doi.org/10.2307/4078077
- 1030 Lorenz, K. Z. (1965). *Evolution and modification of behavior*. University of Chicago
- 1031 Press.
- 1032 Lorenzi, E., Lemaire, B. S., Versace, E., Matsushima, T., & Vallortigara, G. (2020).
- 1033 Resurgence of a perinatal attraction for animate objects via thyroid hormone T3.
- 1034 *BioRxiv*. https://doi.org/10.1101/2020.11.16.384289
- 1035 Lorenzi, E., Mayer, U., Rosa-Salva, O., Morandi-Raikova, A., & Vallortigara, G. (2019).
- 1036 Spontaneous and light-induced lateralization of immediate early genes expression
- 1037 in domestic chicks. *Behavioural Brain Research, 368*, 111905.
- 1038 https://doi.org/10.1016/j.bbr.2019.111905
- 1039 Lorenzi, E., Mayer, U., Rosa-Salva, O., & Vallortigara, G. (2017). Dynamic features of
- animate motion activate septal and preoptic areas in visually naïve chicks (Gallus
- 1041 gallus). *Neuroscience*, *354*, 54–68.
- 1042 https://doi.org/10.1016/j.neuroscience.2017.04.022

- 1043 Lorenzi, E., Pross, A., Rosa-Salva, O., Versace, E., Sgadò, P., & Vallortigara, G. (2019).
- 1044 Embryonic exposure to valproic acid affects social predispositions for dynamic cues
- 1045 of animate motion in newly-hatched chicks. *Frontiers in Physiology*, 10.
- 1046 https://doi.org/10.3389/fphys.2019.00501
- 1047 Lorenzi, E., & Vallortigara, G. (2020). Evolutionary and neural bases of the sense of
- 1048 *animacy*. (A. Kaufman, J. Call, & J. Kaufman, Eds.). Cambridge University Press.
- 1049 Loveland, J. L., Stewart, M. G., & Vallortigara, G. (2019). Effects of oxytocin-family
- 1050 peptides and substance P on locomotor activity and filial preferences in visually
- naïve chicks. *European Journal of Neuroscience*, *50*(10), 3674–3687.
- 1052 https://doi.org/10.1111/ejn.14520
- 1053 Lu, J. W., McMurtry, J. P., & Coon, C. N. (2007). Developmental changes of plasma
- insulin, glucagon, insulin-like growth factors, thyroid hormones, and glucose
- 1055 concentrations in chick embryos and hatched chicks. *Poultry Science*, 86(4), 673–
- 1056 683. https://doi.org/10.1093/ps/86.4.673
- 1057 Marino, L. (2017). Thinking chickens: a review of cognition, emotion, and behavior in
- the domestic chicken. *Animal Cognition*, *20*(2), 127–147.
- 1059 https://doi.org/10.1007/s10071-016-1064-4
- 1060 Mascalzoni, E., Regolin, L., & Vallortigara, G. (2010). Innate sensitivity for self-propelled
- 1061 causal agency in newly hatched chicks. *Proceedings of the National Academy of*
- 1062 Sciences, 107(9), 4483–4485. https://doi.org/10.1073/pnas.0908792107
- 1063 Mayer, U., Rosa-Salva, O., Lorenzi, E., & Vallortigara, G. (2016). Social predisposition

- 1064 dependent neuronal activity in the intermediate medial mesopallium of domestic
- 1065 chicks (Gallus gallus domesticus). *Behavioural Brain Research*, *310*, 93–102.

1066 https://doi.org/10.1016/j.bbr.2016.05.019

- 1067 Mayer, U., Rosa-Salva, O., Loveland, J. L., & Vallortigara, G. (2019). Selective response of
- 1068 the nucleus taeniae of the amygdala to a naturalistic social stimulus in visually
- naive domestic chicks. *Scientific Reports, 9*(1), 9849.
- 1070 https://doi.org/10.1038/s41598-019-46322-5
- 1071 Mayer, U., Rosa-Salva, O., Morbioli, F., & Vallortigara, G. (2017). The motion of a living
- 1072 conspecific activates septal and preoptic areas in naive domestic chicks (Gallus
- 1073 gallus). *European Journal of Neuroscience*, *45*(3), 423–432.
- 1074 https://doi.org/10.1111/ejn.13484
- 1075 Mayer, U., Rosa-Salva, O., & Vallortigara, G. (2017). First exposure to an alive
- 1076 conspecific activates septal and amygdaloid nuclei in visually-naïve domestic chicks
- 1077 (Gallus gallus). *Behavioural Brain Research*, *317*, 71–81.
- 1078 https://doi.org/10.1016/j.bbr.2016.09.031
- 1079 McCabe, B. J. (2019). Visual imprinting in birds: behavior, models, and neural
- 1080 mechanisms. *Frontiers in Physiology*, *10*. https://doi.org/10.3389/fphys.2019.00658
- 1081 McCabe, B. J., & Horn, G. (1994). Learning-related changes in Fos-like immunoreactivity
- 1082 in the chick forebrain after imprinting. *Proceedings of the National Academy of*
- 1083 Sciences, 91(24), 11417–11421. https://doi.org/10.1073/pnas.91.24.11417
- 1084 McCabe, B. J., Horn, G., & Bateson, P. P. G. (1981). Effects of restricted lesions of the

1085 chick forebrain on the acquisition of filial preferences during imprinting. *Brain*

1086 Research, 205(1), 29–37. https://doi.org/10.1016/0006-8993(81)90717-4

- 1087 McNabb, F. A., & King, D. B. (1993). Thyroid hormone effects on growth, development,
- and metabolism. In *The Endocrinology of Growth, Development, and Metabolism*
- 1089 *in Vertebrates* (pp. 873–885). Academic Press.
- 1090 McNabb, F. A. (2006). Avian thyroid development and adaptive plasticity. *General and*
- 1091 *Comparative Endocrinology*, *147*(2), 93–101.
- 1092 https://doi.org/10.1016/j.ygcen.2005.12.011
- 1093 Mehler, J., & Dupoux, E. (1994). What infants know: the new cognitive science of early
- 1094 *development*. Blackwell Publishers. Retrieved from
- 1095 https://eric.ed.gov/?id=ED379059
- 1096 Miura, M., Aoki, N., Yamaguchi, S., Homma, K. J., & Matsushima, T. (2018). Thyroid
- 1097 hormone sensitizes the imprinting-associated induction of biological motion
- 1098 preference in domestic chicks. *Frontiers in Physiology*, 9.
- 1099 https://doi.org/10.3389/fphys.2018.01740
- 1100 Miura, M., & Matsushima, T. (2012). Preference for biological motion in domestic
- 1101 chicks: sex-dependent effect of early visual experience. *Animal Cognition*, 15(5),
- 1102 871–879. https://doi.org/10.1007/s10071-012-0514-x
- 1103 Miura, M., & Matsushima, T. (2016). Biological motion facilitates filial imprinting. Animal
- 1104 Behaviour, 116, 171–180. https://doi.org/10.1016/j.anbehav.2016.03.025

- 1105 Miura, M., Nishi, D., & Matsushima, T. (2020). Combined predisposed preferences for
- 1106 colour and biological motion make robust development of social attachment
- 1107 through imprinting. *Animal Cognition*, *23*(1), 169–188.
- 1108 https://doi.org/10.1007/s10071-019-01327-5
- 1109 Moltz, H., & Stettner, L. J. (1961). The influence of patterned-light deprivation on the
- 1110 critical period for imprinting. Journal of Comparative and Physiological Psychology,
- 1111 54(3), 279–283. https://doi.org/10.1037/h0046991
- 1112 Morton, J., & Johnson, M. H. (1991). CONSPEC and CONLERN: a two-process theory of
- 1113 infant face recognition. *Psychological Review*, *98*(2), 164–181.
- 1114 https://doi.org/10.1037/0033-295X.98.2.164
- 1115 Newman, S. W. (1999). The medial extended amygdala in male reproductive behavior.
- 1116 A node in the mammalian social behavior network. *Annals of the New York*
- 1117 Academy of Sciences, 877, 242–257. https://doi.org/10.1111/j.1749-
- 1118 6632.1999.tb09271.x
- 1119 Nicol, C. J. (2004). Development, direction, and damage limitation: Social learning in
- domestic fowl. *Animal Learning & Behavior, 32*(1), 72–81.
- 1121 https://doi.org/10.3758/BF03196008
- 1122 Nicolini, C., & Fahnestock, M. (2018). The valproic acid-induced rodent model of
- autism. *Experimental Neurology*, *299*, 217–227.
- 1124 https://doi.org/10.1016/j.expneurol.2017.04.017
- 1125 Nishigori, H., Kagami, K., Takahashi, A., Tezuka, Y., Sanbe, A., & Nishigori, H. (2013).

- 1126 Impaired social behavior in chicks exposed to sodium valproate during the last
- 1127 week of embryogenesis. *Psychopharmacology*, *227*(3), 393–402.
- 1128 https://doi.org/10.1007/s00213-013-2979-y
- 1129 O'Connell, L. A., & Hofmann, H. A. (2011). The vertebrate mesolimbic reward system
- and social behavior network: a comparative synthesis. *The Journal of Comparative*
- 1131 *Neurology*, *519*(18), 3599–3639. https://doi.org/10.1002/cne.22735
- 1132 Okuno, H. (2011). Regulation and function of immediate-early genes in the brain:
- 1133 beyond neuronal activity markers. *Neuroscience Research, 69*(3), 175–186.
- 1134 https://doi.org/10.1016/j.neures.2010.12.007
- 1135 Pallante, V., Rucco, D., & Versace, E. (2020). Young chicks quickly lose their
- spontaneous preference to aggregate with females.
- 1137 Parsons, C. H., & Rogers, L. J. (1997). Pharmacological extension of the sensitive period
- 1138 for imprinting in Gallus domesticus. *Physiology & Behavior, 62*(6), 1303–1310.
- 1139 https://doi.org/10.1016/S0031-9384(97)00342-9
- 1140 Parsons, C. H., & Rogers, L. J. (2000). NMDA receptor antagonists extend the sensitive
- 1141 period for imprinting. *Physiology & Behavior, 68*(5), 749–753.
- 1142 https://doi.org/10.1016/S0031-9384(99)00238-3
- 1143 Ratan Murty, N. A., Teng, S., Beeler, D., Mynick, A., Oliva, A., & Kanwisher, N. (2020).
- 1144 Visual experience is not necessary for the development of face selectivity in the
- 1145 lateral fusiform gyrus. *BioRxiv*. https://doi.org/10.1101/2020.02.25.964890

1146	Reh, R. K., Dias, B. G., Nelson, C. A., Kaufer, D., Werker, J. F., Kolb, B., Levine, J. D., &
1147	Hensch, T. K. (2020). Critical period regulation across multiple timescales.
1148	Proceedings of the National Academy of Sciences, 117(38), 23242–23251.
1149	https://doi.org/10.1073/pnas.1820836117
1150	Reid, V. M., Dunn, K., Young, R. J., Amu, J., Donovan, T., & Reissland, N. (2017). The
1151	human fetus preferentially engages with face-like visual stimuli. Current Biology,
1152	<i>27</i> (12), 1825-1828.e3. https://doi.org/10.1016/j.cub.2017.05.044
1153	Rogers, L. J. J. (1997). Early experiential effects on laterality: research on chicks has
1154	relevance to other species. Laterality: Asymmetries of Body, Brain and Cognition,
1155	2(3–4), 199–219. https://doi.org/10.1080/713754277
1156	Rogers, L. J. J., & Deng, C. (1999). Light experience and lateralization of the two visual
1157	pathways in the chick. <i>Behavioural Brain Research, 98</i> (2), 277–287.
1158	https://doi.org/10.1016/S0166-4328(98)00094-1
1159	Rogers, L. J. J. (1990). Light input and the reversal of functional lateralization in the
1160	chicken brain. <i>Behavioural Brain Research, 38</i> (3), 211–221.
1161	https://doi.org/10.1016/0166-4328(90)90176-F
1162	Rogers, L. J. J., & Bolden, S. W. W. (1991). Light-dependent development and
1163	asymmetry of visual projections. Neuroscience Letters, 121(1–2), 63–67.

- 1164 https://doi.org/10.1016/0304-3940(91)90650-I
- 1165 Rogers, L. J. J., & Sink, H. S. . S. (1988). Transient asymmetry in the projections of the
- 1166 rostral thalamus to the visual hyperstriatum of the chicken, and reversal of its

1167 direction by light exposure. *Experimental Brain Research*, 70(2).

1168 https://doi.org/10.1007/BF00248362

- 1169 Rogers, L. J. J., & Workman, L. (1989). Light exposure during incubation affects
- 1170 competitive behaviour in domestic chicks. Applied Animal Behaviour Science, 23(3),
- 1171 187–198. https://doi.org/10.1016/0168-1591(89)90109-3
- 1172 Rogers, L. J. J. (1982). Light experience and asymmetry of brain function in chickens.
- 1173 Nature, 297(5863), 223–225. https://doi.org/10.1038/297223a0
- 1174 Rosa-Salva, O., Grassi, M., Lorenzi, E., Regolin, L., & Vallortigara, G. (2016). Spontaneous
- 1175 preference for visual cues of animacy in naïve domestic chicks: The case of speed
- 1176 changes. *Cognition*, *157*, 49–60. https://doi.org/10.1016/j.cognition.2016.08.014
- 1177 Rosa-Salva, O., Hernik, M., Broseghini, A., & Vallortigara, G. (2018). Visually-naïve chicks
- 1178 prefer agents that move as if constrained by a bilateral body-plan. *Cognition*, *173*,
- 1179 106–114. https://doi.org/10.1016/j.cognition.2018.01.004
- 1180 Rosa-Salva, O., Mayer, U., & Vallortigara, G. (2015). Roots of a social brain:
- developmental models of emerging animacy-detection mechanisms. *Neuroscience*
- 1182 & Biobehavioral Reviews, 50, 150–168.
- 1183 https://doi.org/10.1016/j.neubiorev.2014.12.015
- 1184 Rosa-Salva, O., Mayer, U., & Vallortigara, G. (2019). Unlearned visual preferences for the
- head region in domestic chicks. *PLOS ONE*, *14*(9), e0222079.
- 1186 https://doi.org/10.1371/journal.pone.0222079

1187 Rosa-Salva, O., Regolin, L., & Vallortigara, G. (2007). Chicks discriminate human gaze

1188 with their right hemisphere. *Behavioural Brain Research*, *177*(1), 15–21.

1189 https://doi.org/10.1016/j.bbr.2006.11.020

- 1190 Rosa-Salva, O., Regolin, L., & Vallortigara, G. (2010). Faces are special for newly hatched
- 1191 chicks: evidence for inborn domain-specific mechanisms underlying spontaneous
- 1192 preferences for face-like stimuli. *Developmental Science*, *13*(4), 565–577.

1193 https://doi.org/10.1111/j.1467-7687.2009.00914.x

- 1194 Rosa-Salva, O., Regolin, L., & Vallortigara, G. (2012). Inversion of contrast polarity
- abolishes spontaneous preferences for face-like stimuli in newborn chicks.

1196 Behavioural Brain Research, 228(1), 133–143.

- 1197 https://doi.org/10.1016/j.bbr.2011.11.025
- 1198 Rosa-Salva, O., Farroni, T., Regolin, L., Vallortigara, G., & Johnson, M. H. (2011). The
- evolution of social orienting: evidence from chicks (Gallus gallus) and human
- 1200 newborns. *PLoS ONE*, *6*(4), e18802. https://doi.org/10.1371/journal.pone.0018802
- 1201 Rugani, R., Rosa-Salva, O., Regolin, L., & Vallortigara, G. (2015). Brain asymmetry

1202 modulates perception of biological motion in newborn chicks (Gallus gallus).

- 1203 Behavioural Brain Research, 290, 1–7. https://doi.org/10.1016/j.bbr.2015.04.032
- 1204 Salzen, E. A., Lily, R. E., & McKeown, J. R. (1971). Colour preference and imprinting in
- domestic chicks. *Animal Behaviour*, *19*(3), 542–547. https://doi.org/10.1016/S0003 3472(71)80109-4
- 1207 Salzen, E. A., & Meyer, C. C. (1968). Reversibility of imprinting. *Journal of Comparative*

1208 and Physiological Psychology, 66(2), 269–275. https://doi.org/10.1037/h0026349

1209	Sauvage, M., Kitsukawa, T., & Atucha, E. (2019). Single-cell memory trace imaging with
1210	immediate-early genes. Journal of Neuroscience Methods, 326, 108368.
1211	https://doi.org/10.1016/j.jneumeth.2019.108368
1212	Scaife, M. (1976). The response to eye-like shapes by birds II. The importance of
1213	staring, pairedness and shape. Animal Behaviour, 24(1), 200–206.
1214	https://doi.org/10.1016/S0003-3472(76)80116-9
1215	Schiff, W. (1965). Perception of impending collision: A study of visually directed
1216	avoidant behavior. <i>Psychological Monographs: General and Applied, 79</i> (11), 1–26.
1217	https://doi.org/10.1037/h0093887
1218	Schleidt, W. M. (2010). How "fixed" is the fixed action pattern? Zeitschrift Für
1219	<i>Tierpsychologie, 36</i> (1–5), 184–211. https://doi.org/10.1111/j.1439-
1220	0310.1974.tb02131.x
1221	Schulman, A. H., Hale, E. B., & Graves, H. B. (1970). Visual stimulus characteristics for
1222	initial approach response in chicks (Gallus domesticus). Animal Behaviour, 18, 461–
1223	466. https://doi.org/10.1016/0003-3472(70)90040-0
1224	Sgadò, P., Rosa-Salva, O., Versace, E., & Vallortigara, G. (2018). Embryonic exposure to
1225	valproic acid impairs social predispositions of newly-hatched chicks. Scientific
1226	<i>Reports, 8</i> (1), 5919. https://doi.org/10.1038/s41598-018-24202-8
1227	Shultz, S., Klin, A., & Jones, W. (2018). Neonatal transitions in social behavior and their

- implications for autism. *Trends in Cognitive Sciences*, *22*(5), 452–469.
- 1229 https://doi.org/10.1016/j.tics.2018.02.012
- 1230 Silingardi, D., Scali, M., Belluomini, G., & Pizzorusso, T. (2010). Epigenetic treatments of
- adult rats promote recovery from visual acuity deficits induced by long-term
- 1232 monocular deprivation. *European Journal of Neuroscience*, *31*(12), 2185–2192.
- 1233 https://doi.org/10.1111/j.1460-9568.2010.07261.x
- 1234 Simion, F., & Di Giorgio, E. (2015). Face perception and processing in early infancy:
- inborn predispositions and developmental changes. *Frontiers in Psychology, 6.*
- 1236 https://doi.org/10.3389/fpsyg.2015.00969
- 1237 Simion, F., Regolin, L., & Bulf, H. (2008). A predisposition for biological motion in the
- newborn baby. *Proceedings of the National Academy of Sciences, 105*(2), 809–813.
- 1239 https://doi.org/10.1073/pnas.0707021105
- 1240 Simner, M. L. (1973). The development of visual flicker rate preference in the newly
- 1241 hatched chick. *Developmental Psychobiology*, *6*(4), 377–384.
- 1242 https://doi.org/10.1002/dev.420060411
- 1243 Sluckin, W. (1972). *Imprinting and early Learning*. Methuen young books.
- 1244 Sugita, Y. (2008). Face perception in monkeys reared with no exposure to faces.
- 1245 Proceedings of the National Academy of Sciences of the United States of America,
- 1246 *105*(1), 394–398. https://doi.org/10.1073/pnas.0706079105
- 1247 Takemura, Y., Yamaguchi, S., Aoki, N., Miura, M., Homma, K. J., & Matsushima, T. (2018).

1248	Gene expression of Dio2 (thyroid hormone converting enzyme) in telencephalon is
1249	linked with predisposed biological motion preference in domestic chicks.
1250	<i>Behavioural Brain Research, 349</i> , 25–30. https://doi.org/10.1016/j.bbr.2018.04.039
1251	Taylor, A., Sluckin, W., & Hewitt, R. (1969). Changing colour preferences of chicks.
1252	Animal Behaviour, 17, 3–8. https://doi.org/10.1016/0003-3472(69)90105-5
1253	Ten Cate, C., & Vos, D. R. (1999). Sexual imprinting and evolutionary processes in birds:
1254	a reassessment (pp. 1–31). https://doi.org/10.1016/S0065-3454(08)60214-4
1255	Tomalski, P., Csibra, G., & Johnson, M. H. (2009). Rapid orienting toward face-like
1256	stimuli with gaze-relevant contrast information. Perception, 38(4), 569–578.
1257	https://doi.org/10.1068/p6137
1258	Vallortigara, G. (i <i>n press</i>). <i>Born knowing. The origins of knowledge</i> . MIT press.
1259	Vallortigara, G. (1992). Right hemisphere advantage for social recognition in the chick.
1260	<i>Neuropsychologia</i> , <i>30</i> (9), 761–768. https://doi.org/10.1016/0028-3932(92)90080-6
1261	Vallortigara, G. (2012). Aristotle and the chicken: animacy and the origins of beliefs. In
1262	The Theory of Evolution and Its Impact (pp. 189–199). Milano: Springer Milan.
1263	https://doi.org/10.1007/978-88-470-1974-4_12
1264	Vallortigara, G., & Andrew, R. J. (1991). Lateralization of response by chicks to change
1265	in a model partner. Animal Behaviour, 41(2), 187–194.
1266	https://doi.org/10.1016/S0003-3472(05)80470-1

1267 Vallortigara, G., & Andrew, R. J. (1994). Differential involvement of right and left

1268 hemisphere in individual recognition in the domestic chick. *Behavioural Processes*,

1269 *33*(1–2), 41–57. https://doi.org/10.1016/0376-6357(94)90059-0

- 1270 Vallortigara, G., Regolin, L., & Marconato, F. (2005). Visually inexperienced chicks exhibit
- 1271 spontaneous preference for biological motion patterns. *PLoS Biology*, *3*(7), e208.
- 1272 https://doi.org/10.1371/journal.pbio.0030208
- 1273 Vallortigara, G., & Rogers, L. J. J. (2005). Survival with an asymmetrical brain:
- advantages and disadvantages of cerebral lateralization. *Behavioral and Brain*
- 1275 Sciences, 28(4), 575–589. https://doi.org/10.1017/S0140525X05000105
- 1276 Vallortigara, G., & Versace, E. (2018). Filial imprinting. In *Encyclopedia of Animal*
- 1277 *Cognition and Behavior* (pp. 1–4). Cham: Springer International Publishing.

1278 https://doi.org/10.1007/978-3-319-47829-6_1989-1

- 1279 Versace, E. (2017). Precocial. In Encyclopedia of Animal Cognition and Behavior (pp. 1–
- 1280 3). Cham: Springer International Publishing. https://doi.org/10.1007/978-3-319-
- 1281 47829-6_459-1
- 1282 Versace, E., Damini, S., Caffini, M., & Stancher, G. (2018). Born to be asocial: newly
- 1283 hatched tortoises avoid unfamiliar individuals. *Animal Behaviour, 138*, 187–192.
- 1284 https://doi.org/10.1016/j.anbehav.2018.02.012
- 1285 Versace, E., Damini, S., & Stancher, G. (2020). Early preference for face-like stimuli in
- 1286 solitary species as revealed by tortoise hatchlings. *Proceedings of the National*
- 1287 Academy of Sciences.

1288	Versace, E., Fracasso, I., Baldan, G., Dalle Zotte, A., & Vallortigara, G. (2017). Newborn
1289	chicks show inherited variability in early social predispositions for hen-like stimuli.

1290 Scientific Reports, 7(1), 40296. https://doi.org/10.1038/srep40296

- 1291 Versace, E., Martinho-Truswell, A., Kacelnik, A., & Vallortigara, G. (2018). Priors in animal
- and artificial intelligence: where does learning begin? *Trends in Cognitive Sciences*,
- 1293 22(11), 963–965. https://doi.org/10.1016/j.tics.2018.07.005
- 1294 Versace, E., Ragusa, M., & Vallortigara, G. (2019). A transient time window for early
- 1295 predispositions in newborn chicks. *Scientific Reports*, *9*(1), 18767.
- 1296 https://doi.org/10.1038/s41598-019-55255-y
- 1297 Versace, E., Schill, J., Nencini, A. M. M., & Vallortigara, G. (2016). Naïve chicks prefer
- 1298 hollow objects. *PLOS ONE*, *11*(11), e0166425.
- 1299 https://doi.org/10.1371/journal.pone.0166425
- 1300 Versace, E., & Vallortigara, G. (2015). Origins of Knowledge: insights from precocial
- 1301 species. Frontiers in Behavioral Neuroscience, 9.
- 1302 https://doi.org/10.3389/fnbeh.2015.00338
- 1303 Vidal, J.-M. (1980). The relations between filial and sexual imprinting in the domestic
- 1304 fowl: effects of age and social experience. *Animal Behaviour, 28*(3), 880–891.
- 1305 https://doi.org/10.1016/S0003-3472(80)80148-5
- 1306 Weirich, R. T., & McNabb, F. M. A. (1984). Nuclear receptors for I-triiodothyronine in
- 1307 quail liver. *General and Comparative Endocrinology*, *53*(1), 90–99.
- 1308 https://doi.org/10.1016/0016-6480(84)90228-4

1309	Weiss, J., Köhler, W., & Landsberg, JW. (1977). Increase of the corticosterone level in
1310	ducklings during the sensitive period of the following response. Developmental
1311	<i>Psychobiology, 10</i> (1), 59–64. https://doi.org/10.1002/dev.420100109
1312	Widowski, T. M., Lo Fo Wong, D. M., & Duncan, I. J. (1998). Rearing with males
1313	accelerates onset of sexual maturity in female domestic fowl. Poultry Science, 77(1),
1314	150–155. https://doi.org/10.1093/ps/77.1.150
1315	Yamaguchi, S., Aoki, N., Kitajima, T., Iikubo, E., Katagiri, S., Matsushima, T., & Homma, K.
1316	J. (2012). Thyroid hormone determines the start of the sensitive period of
1317	imprinting and primes later learning. Nature Communications, 3(1), 1081.
1318	https://doi.org/10.1038/ncomms2088
1319	Yamaguchi, S., Aoki, N., Matsushima, T., & Homma, K. J. (2018). Wnt-2b in the
1320	intermediate hyperpallium apicale of the telencephalon is critical for the thyroid
1321	hormone-mediated opening of the sensitive period for filial imprinting in domestic
1322	chicks (Gallus gallus domesticus). <i>Hormones and Behavior</i> , 102, 120–128.
1323	https://doi.org/10.1016/j.yhbeh.2018.05.011
1324	Yamaguchi, S., Aoki, N., Takehara, A., Mori, M., Kanai, A., Matsushima, T., & Homma, K.

1325 J. (2016). Involvement of nucleotide diphosphate kinase 2 in the reopening of the

- 1326 sensitive period of filial imprinting of domestic chicks (Gallus gallus domesticus).
- 1327 Neuroscience Letters, 612, 32–37. https://doi.org/10.1016/j.neulet.2015.12.004
- 1328 Zachar, G., Tóth, A. S., Gerecsei, L. I., Zsebők, S., Ádám, Á., & Csillag, A. (2019).
- 1329 Valproate exposure in ovo attenuates the acquisition of social preferences of

- 1330 young post-hatch domestic chicks. *Frontiers in Physiology*, 10.
- 1331 https://doi.org/10.3389/fphys.2019.00881
- 1332 Zeanah, C. H., Egger, H. L., Smyke, A. T., Nelson, C. A., Fox, N. A., Marshall, P. J., &
- 1333 Guthrie, D. (2009). Institutional rearing and psychiatric disorders in Romanian
- 1334 preschool children. *American Journal of Psychiatry*, *166*(7), 777–785.
- 1335 https://doi.org/10.1176/appi.ajp.2009.08091438

- precocial avian species are optimal models to study social cognition development
- predisposed and environmental factors interact in the development of domestic chicks
- social predispositions require activating experiences, occurring in critical periods
- social predispositions and filial imprinting are reciprocally interacting mechanisms
- hormonal and neural mechanisms control critical periods for social development

