



**BIOMATERIALS AND REGENERATIVE TECHNOLOGIES USED IN BONE REGENERATION IN THE CRANEO-MAXILLO-FACIAL REGION. Consensus report of group 2 of the 15th European Workshop on Periodontology on Bone Regeneration**

Journal:	<i>Journal of Clinical Periodontology</i>
Manuscript ID	Draft
Manuscript Type:	Supplement Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Sanz, Mariano; Universidad Complutense de Madrid, Periodoncia
Topic:	Treatment
Main Methodology:	Systematic Review
Keywords:	bone regeneration, biomaterials, barrier membrane, bioactive agents, cell therapies

SCHOLARONE™  
Manuscripts

1  
2  
3  
4  
5  
6 **BIOMATERIALS AND REGENERATIVE TECHNOLOGIES USED IN**  
7 **BONE REGENERATION IN THE CRANEO-MAXILLO-FACIAL**  
8 **REGION. Consensus report of group 2 of the 15th European**  
9 **Workshop on Periodontology on Bone Regeneration**  
10  
11  
12  
13  
14

15 *Sanz M<sup>1</sup>, Dahlin C<sup>2</sup>, Apatzidou D<sup>3</sup>, Artzi Z<sup>4</sup>, Bozic D<sup>5</sup>, Calciolari E<sup>6</sup>, De Bruyn H<sup>7</sup>,*  
16 *Dommisch H<sup>8</sup>, Donos N<sup>6</sup>, Eickholz P<sup>9</sup>, Ellingsen JE<sup>10</sup>, Haugen HJ<sup>11</sup>, Herrera D<sup>1</sup>,*  
17 *Lambert F<sup>12</sup>, Layrolle P<sup>13</sup>, Montero E<sup>1</sup>, Mustafa K<sup>14</sup>, Omar O<sup>2</sup> and Schliephake H<sup>15</sup>*  
18  
19  
20  
21

22  
23 *1. Department of Dental Clinical Specialties and ETEP Research Group, Faculty of*  
24 *Odontology, University Complutense of Madrid, Plaza Ramon y Cajal, E-28040*  
25 *Madrid, Spain*  
26

27  
28 *2. Department of Biomaterials, Institute of Clinical Sciences, Sahlgrenska Academy,*  
29 *University of Gothenburg, Box 412, SE-405 30 Gothenburg, Sweden*  
30

31  
32 *3. Department of Preventive Dentistry, Periodontology and Implant Biology, School*  
33 *of Dentistry, Aristotle University of Thessaloniki, Thessaloniki, Greece*  
34

35  
36 *4. Department of Periodontology and Oral Implantology, School of Dental Medicine,*  
37 *Tel Aviv University, Ramat Aviv, Tel Aviv 69978, Israel*  
38

39  
40 *5. Department of Periodontology, School of Dental Medicine, University of Zagreb,*  
41 *Gunduliceva 5, 10000 Zagreb, Croatia*  
42

43  
44 *6. Centre for Immunobiology & Regenerative Medicine & Centre for Oral Clinical*  
45 *Research, Institute of Dentistry, Barts & The London School of Medicine and*  
46 *Dentistry, Queen Mary University of London (QMUL), Turner Street E12AD, London,*  
47 *UK*  
48

49  
50 *7. Department Periodontology & Implantology. College of Dental Science, Radboud*  
51 *University Medical Center. Nijmegen, The Netherland*  
52

53  
54 *8. Department of Periodontology and Synoptic Dentistry Charité -*  
55 *Universitätsmedizin Berlin Assmannshausen Strasse 4-6, 14197 Berlin, Germany*  
56

57  
58 *9. Department of Periodontology, Johann Wolfgang Goethe-University, Frankfurt,*  
59 *Germany*  
60

1  
2  
3  
4 *10. Department of Prosthetics an Oral Function, Institute of Clinical Dentistry, Faculty*  
5 *of Dentistry, University of Oslo, PO Box 1109 Blindern, 0317 Oslo, Norway*

6  
7 *11. Department of Biomaterials, Institute of Clinical Dentistry, Faculty of Dentistry,*  
8 *University of Oslo, PO Box 1109 Blindern, 0317 Oslo, Norway*

9  
10  
11 *12 Dental Biomaterials Research Unit (d-BRU), University of Liège (ULiège),*  
12 *Belgium. Dpt. of Periodontology and Oral Surgery, University of Liège (ULiège),*  
13 *Belgium*

14  
15  
16 *13 Inserm, U791, Laboratory for Osteoarticular and Dental Tissue Engineering,*  
17 *Faculty of Dental Surgery, University of Nantes, 1 Place Alexis Ricordeau, 44042*  
18 *Nantes cedex 1, France*

19  
20  
21 *14. Department of Clinical Dentistry, Center for Clinical Dental Research, University*  
22 *of Bergen, Norway*

23  
24  
25 *15. Department of Oral and Maxillofacial Surgery, George-Augusta-University,*  
26 *Gottingen, Germany*

### 27 28 29 30 **Key Words**

31 Bone regeneration, biomaterials, bone replacement graft, osteoconductive,  
32 osteoinductive, barrier membrane, guided bone regeneration, bio-absorbable,  
33 bioactive agent, cell therapies  
34  
35  
36

### 37 38 **Abstract**

39 **Background and Aims:** to review the regenerative technologies used in bone  
40 regeneration: bone grafts, barrier membranes, bioactive factors and cell therapies

41 **Material and Methods:** four background review publications served to elaborate this  
42 consensus report  
43  
44

45 **Results and Conclusions:** Biomaterials used as bone grafts must meet specific  
46 requirements: biocompatibility, porosity, osteoconductivity, osteoinductivity, surface  
47 properties, biodegradability, mechanical properties, angiogenicity, handling and  
48 manufacturing processes. Currently used biomaterials have demonstrated  
49 advantages and limitations based on the fulfillment of these requirements. Similarly,  
50 membranes for guided bone regeneration (GBR) must fulfill specific properties and  
51 potential biological mechanisms to improve their clinical applicability. Pre-clinical and  
52 clinical studies have evaluated the added effect of Bone Morphogenetic Proteins  
53 (mainly BMP-2) and Autologous Platelet Concentrates (APCs) when used as  
54  
55  
56  
57  
58  
59  
60

1  
2  
3  
4 bioactive agents to enhance bone regeneration. Three main approaches using cell  
5 therapies to enhance bone regeneration have been evaluated: a) “minimally  
6 manipulated” whole tissue fractions; b) ex-vivo expanded “uncommitted”  
7 stem/progenitor cells, and c) ex-vivo expanded “committed” bone-/periosteum-  
8 derived cells. Based on the evidence from clinical trials, transplantation of cells, most  
9 commonly whole BMA or BMAC, in combination with biomaterial scaffolds has  
10 demonstrated an additional effect in sinus augmentation and horizontal ridge  
11 augmentation, and comparable bone regeneration to autogenous bone in alveolar  
12 cleft repair.  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

## Introduction

This consensus report aims to describe the regenerative technologies currently used in bone regenerative interventions in the craniomaxillofacial region. The relevant scientific evidence that served to elaborate this report came from four background publications, being three narrative reviews using a systematic search approach and one systematic review, respectively:

1. Bone grafts: which is the ideal biomaterial? Haugen HJ, Lyngstadaas SP, Rossi, F and Perale G
2. Barrier membranes: more than the barrier effect? Omar O, Elgali I, Dahlin C and Thomsen P
3. The use of bioactive factors to enhance bone regeneration. A narrative review Donos N, Dereka X and Calciolari E
4. Cell therapy for orofacial bone regeneration: a systematic review and meta-analysis Shanbhag S, Suliman S, Pandis N, Stavropoulos A, Sanz M and Mustafa K

## Biomaterials used as bone replacement grafts in regenerative interventions in the craniomaxillo-facial region

Biomaterials used as bone replacement grafts must meet specific requirements to achieve the goal of developing a new and healthy bone tissue formation:

- *Biocompatibility*. The interaction between the material and the tissues should not adversely affect the surrounding tissues, the intended healing result or the safety of the patient. (Williams, 2017). Ideally, biomaterials should be inherently bioactive in promoting the bone regeneration process (e.g. ion release, surface characteristics, etc.).
- *Porosity*. An adequate pore size, morphology and inter-connectivity is needed to allow for diffusion throughout the whole scaffold of bone cells, nutrients and exchange of waste products. It is important to distinguish between micro-porosity and macro-porosity (Hutmacher, 2000). Micro-porosity is defined as pores  $\leq 10 \mu\text{m}$  to improve cell adhesion, to allow fluids and nutrients flow (permeability) and thus to enhance the bioactivity. Macro-porosity is defined as pores  $\geq 100 \mu\text{m}$  to allow

1  
2  
3  
4 for angiogenesis and to bone cell ingrowth, thus mimicking the porosity of  
5 trabecular bone, which has a mean value 250  $\mu\text{m}$ , although it is highly variable.  
6 Inter-connectivity (the connection between pores) is also an important property to  
7 allow for permeability, vascularization and bone ingrowth.  
8  
9

- 10  
11 - *Osteoconductivity/ Osteoinductivity*. All biomaterials used for bone regeneration  
12 should allow for bone growth directly in contact with the biomaterial surface from  
13 the surrounding bone (*osteoconduction*), but ideally it should also be able to  
14 promote *osteoinduction* (Albrektsson, 2001). An *osteoinductive* biomaterial should  
15 first be capable of recruiting mesenchymal-type osteoprogenitor cells. Secondly, it  
16 should be capable of transforming an undifferentiated mesenchymal cell into a  
17 mature, bone-forming osteoblast. Lastly, it should be capable of inducing in-growth  
18 of ectopic bone formation when implanted into extra-skeletal locations. This  
19 capacity may be related to its micro-porosity and surface properties.  
20  
21  
22 - *Surface properties*. Surface topography at the nano and micro level as well as  
23 surface physico-chemistry are important characteristics for protein adsorption,  
24 extracellular matrix deposition, cell adhesion, differentiation, migration and finally  
25 bone formation.  
26  
27  
28 - *Biodegradability*. The capacity of the biomaterial to bio-absorb during the tissue  
29 healing and remodeling process. The ideal bone graft substitute is expected to be  
30 fully replaced by bone, preferably at a predictable absorption rate, without losing  
31 tissue volume and without interfering with the healing and regeneration process. In  
32 case of biomaterials with a slow bio-absorbability rate, these should assure a  
33 process of new bone formation with sufficient volume in contact with the  
34 biomaterial.  
35  
36  
37 - *Mechanical properties*. Compressive strength and elasticity should be high enough  
38 to absorb the load from the surrounding hard and soft tissues in non-contained  
39 defects. Ideally the compressive strength and elasticity of the biomaterial should  
40 be at least those of the natural bone at the site of regeneration. These mechanical  
41 properties are also influenced by pore morphology and size.  
42  
43  
44 - *Angiogenicity*. The inherent biomaterial properties (porosity, surface, etc.) should  
45 promote angiogenesis and the appropriate vascularization of the graft volume.  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

- 1  
2  
3  
4 - *Handling*. The biomaterial should be cohesive and dimensionally stable, easy for  
5  
6 chairside use to adapt to the defect. When used in non-contained defects, it  
7  
8 should allow for three-dimensional build up. Biomaterials for craniomaxillo-facial  
9  
10 bone regeneration are usually available in the form of granules or blocks.  
11  
12 Depending on the clinical needs, it is desirable to have a wide variability of sizes  
13  
14 and forms, ranging from 0.1 mm-2.0 mm in the particulate form. For certain  
15  
16 indications an injectable mode of application would be desired to fill the defect  
17  
18 volume through its plasticity.
- 19 - *Manufacturing processes*. The biomaterial should be provided with certification or  
20  
21 documentation of the appropriate manufacturing and sterilization processes and  
22  
23 assure long shelf time and reduced production costs.  
24  
25  
26

27 ***Which are the advantages and limitations of the currently used bone grafts for***  
28 ***craniomaxillo-facial bone regeneration?***  
29

30  
31 *Autologous*  
32

33 Even though autologous bone is not a biomaterial per se, it is considered the gold  
34  
35 standard graft material for bone regeneration and it has the following advantages: it  
36  
37 contains the patient's own cells, growth factors and bio-molecules needed for  
38  
39 osteogenesis, it has the highest degree of biological safety, biocompatibility,  
40  
41 matched mechanical properties and scaffolding effect.  
42

43 In regard to limitations, autologous grafts may need a second surgical site for its  
44  
45 harvesting, which increases patient's morbidity, pain or discomfort and other  
46  
47 complications related to increased surgical time and invasiveness. It has been  
48  
49 reported that the resorption of these bone replacement grafts is higher, and their rate  
50  
51 of resorption is not predictable. Depending on the source of the graft (cortical versus  
52  
53 cancellous bone) the vascularization may be slowed, mainly in highly cortical bone  
54  
55 grafts. It has also limitations in terms of volume availability, mainly when harvesting  
56  
57 from intraoral sources and the resulting grafts, mainly in a block form may be difficult  
58  
59 to adapt to the anatomy of the defect.  
60

1  
2  
3  
4 The application of particulate dentin has been recently suggested as another  
5 autologous source for minor ridge augmentation or socket site preservation.  
6  
7 However, there is no clinical documentation to substantiate its clinical use.  
8

### 9 10 *Allogenic*

11  
12 There are different ways of processing allogenic bone replacement grafts (freeze  
13 dried, fresh frozen, etc.), which may change their biological properties. These  
14 allografts can be produced as particulate or blocks. In principle, the general  
15 advantage of this biomaterial is that it provides similar mechanical properties as the  
16 autologous bone and it may contain the collagenous matrix and proteins of natural  
17 bone, although it lacks viable cells. Similarly, handling properties are comparable to  
18 autologous bone, although the reduced surgical time needed for their implantation, in  
19 addition to their increased availability are clear advantages when compared with  
20 autologous bone.  
21  
22

23  
24 Its biological safety due to possible disease transmission and potential unwanted  
25 immune reactions are clear disadvantages. Furthermore, the sources of donor  
26 material are heterogeneous, what may influence their biological activity and similarly,  
27 resorption rate are highly variable. Other drawbacks could be possible impairment to  
28 achieve vascularization of the grafted site. In the future its availability for clinical use  
29 may be reduced in light of the regulatory changes in Europe.  
30  
31

### 32 33 *Xenogeneic*

34  
35 This is the biomaterial with the most documentation in the scientific literature for  
36 bone grafting in the cranio-maxillo-facial area. Its main advantages are that since it is  
37 derived from both natural cancellous and cortical bone its architecture and geometric  
38 structure resemble bone, although highly depending on the tissue source and  
39 manufacturing process. Its slow bio-absorbability may be a clinical advantage for  
40 preserving the augmented bone volume.  
41  
42

43  
44 In regard to limitations, it lacks biological components thus limiting its biological  
45 activity. Similar to allogenic materials, its use implies a potential biological risk of  
46 disease transmission (e.g. prions and retroviruses) and/or immunogenic host tissue  
47 response, although these risks can be diminished through the manufacturing  
48 process (de-proteinization). In spite of this inherent risk, however, transmission of  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60



1  
2  
3  
4 bovine spongiform encephalitis (BSE) has not yet been reported associated with the  
5  
6 implantation of this biomaterial.

7  
8 Mechanical properties (brittleness) may vary depending on the source and  
9  
10 manufacturing process. Since these biomaterials are mainly available for use in  
11  
12 particulate form, they may have limitations in large defect regeneration interventions.

### 13 14 *Synthetic Bio Ceramics*

15  
16 Calcium sulphate, calcium phosphate (CaP), bioactive glass and combinations are  
17  
18 the most commonly used bio ceramics available at present. Their main advantage is  
19  
20 the controlled manufacturing process which may assure biocompatibility,  
21  
22 biodegradability and similarity in structure and inorganic composition to natural bone  
23  
24 minerals. The most investigated CaP bone graft substitutes are hydroxyapatite (HA),  
25  
26  $\beta$ -tricalcium phosphate ( $\beta$ -TCP) and their combination, also called biphasic calcium  
27  
28 phosphate (BCF). Bio ceramics have shown osteoinductive properties through the  
29  
30 stimulation of inorganic matrix deposition, osteoblast differentiation, osteoblast  
31  
32 growth and bone promotion. By modulating their chemical composition and sintering  
33  
34 temperature their bioactivity and degradation time can be controlled to a certain  
35  
36 extent.

37  
38 Their main disadvantage is associated with their limited mechanical properties (load  
39  
40 bearing resistance) and unpredictable bio-absorption rates. They are mainly  
41  
42 delivered as particulates, which may limit their use in large bone defects.

43  
44 Glass-based ceramics share the same problems of lower mechanical strength  
45  
46 despite excellent material-bone interactions. Similarly, their degradation times can be  
47  
48 unpredictable.

49  
50 To improve their mechanical properties (brittleness), bio ceramics have been mixed  
51  
52 with polymers developing composite materials (Laurencin et al.,2014).

### 53 54 *Synthetic Polymers*

55  
56 The most studied synthetic polymers used as biomaterials for bone tissue  
57  
58 regeneration are aliphatic polyesters like poly (lactic-acid) (PLA), poly ( $\epsilon$ -  
59  
60 caprolactone) (PCL), and poly (glycolic-acid) (PGA) and their copolymers and  
61  
62 derivatives. They share the advantage that their manufacture is controllable and

1  
2  
3  
4 tunable in terms of adjusting their physiochemical structure, porosity and hence their  
5 biodegradability and shape, size and biomechanical properties can be customized.  
6  
7

8 Their most important limitation is that they have not demonstrated osteoconductivity  
9 and hence, their use as bone replacement grafts requires their combination with bio  
10 ceramics as composite materials or they can be functionalized, for example with  
11 different coatings. Their process of bio-absorbability usually leads to the release of  
12 acid compounds that may interfere with wound healing, although this limitation can  
13 be controlled by the manufacturing process as composite materials can be combined  
14 with bio ceramics. Furthermore, the bio absorbability of synthetic polymers is highly  
15 variable, which may impair their mechanical strength *in vivo*.  
16  
17  
18  
19  
20  
21

### 22 *Composite bio-materials*

23 Composite biomaterials are biomaterials generated by combining bio ceramics with  
24 polymers or xenogeneic biomaterials together with bio ceramics or polymers. Their  
25 properties will vary depending on their composition and manufacturing processes.  
26  
27  
28  
29  
30

### 31 *Other synthetic biomaterials*

32 Granules made of titanium particles were marketed for use in bone regeneration,  
33 although their use is no longer available in the market.  
34  
35  
36  
37

### 38 ***Recommendations for Future Research***

39 The future of cranio-maxillo-facial bone regeneration will probably entail the  
40 manufacturing of personalized biomaterials from 3-D digital data obtained from  
41 patients. Additive manufacturing (e.g. 3-D printing) of different biomaterials (e.g. bio  
42 ceramics) will allow rapid production of these customized scaffolds that will perfectly  
43 fit the bone defect anatomy. The addition of synthetic polymers in the design of  
44 composite biomaterials may mechanically reinforce these 3D constructed  
45 biomaterials. Similarly, the addition of cells (bio-printing) may add biological activity  
46 to the 3-D printed constructs  
47  
48  
49  
50  
51  
52  
53  
54

55 Future biomaterials should have optimized surface characteristics, pore size and  
56 interconnection. These characteristics will be adjusted to control of their bio-  
57 absorbability, promote osteoinduction and ensure ideal mechanical properties.  
58  
59  
60

1  
2  
3  
4 Biomimetic biomaterials should be developed at ambient temperatures by hydrolysis  
5 and precipitation of calcium deficient apatite, what will result in similar composition  
6 and crystallinity as natural bone. These biomaterials should be completely replaced  
7 by new bone through controlled processes of bio-absorbability and osteoinduction.  
8  
9  
10

11  
12 There is a need of standardized and validated pre-clinical models, with the use of  
13 small animal models for screening and large-animal models for comparing new  
14 biomaterials using established standards. In concordance with the ARRIVE  
15 guidelines to reduce animal research, there is a need for standardized pre-clinical  
16 models, such as *in silico* modeling and *ex-vivo* tissue engineering testing to reduce  
17 animal research.  
18  
19  
20  
21  
22  
23

## 24 25 **Membranes for Guided Bone Regeneration used in regenerative** 26 **interventions in the cariomaxillo-facial region** 27 28

29 This report addresses the scientific evidence on the effects of membranes used for  
30 guided bone regeneration (GBR), focusing on their properties and potential biological  
31 mechanisms, regardless of the clinical applicability.  
32  
33  
34

35 Ideally, a new developed membrane should pass the cascades of evaluations from  
36 *in vitro* to clinical testing until being approved as medical devices according to  
37 current ISO standards and specifications. In addition, the new European Medical  
38 Device Regulation (EU-MDR) for implantable medical devices requires confirmation  
39 of the product claims through prospective clinical studies.  
40  
41  
42  
43

44 Beside their inherent barrier effect, membranes for guided bone regeneration should  
45 have certain properties:  
46  
47

- 48 - *Biocompatibility*. The biomaterial shall perform with an appropriate tissue  
49 response. Hence, the interaction between the material and the tissues should not  
50 adversely affect the surrounding tissues, the intended healing result or the safety  
51 of the patient. (Williams, 2017).  
52
- 53 - *Biological activity*. There is increasing evidence that membranes not only function  
54 due to their occlusive properties, but they actively promote bone regeneration  
55 within the osseous defect below the membrane. Specifically, this biological activity  
56  
57  
58  
59  
60

1  
2  
3  
4 may include recruitment of cells, angiogenesis, bone formation and bone  
5 remodeling leading to bone fill of the defect with mature bone. There is  
6 experimental evidence that collagen membranes allow inward migration of cells  
7 that express and secrete osteogenic and angiogenic factors. It is not yet  
8 established whether similar biological processes are shared by membranes of  
9 different composition.  
10  
11  
12  
13

- 14  
15 - *Porosity / occlusive properties.* A defined porosity or a certain degree of barrier  
16 effect of the membrane are not prerequisites for their use in guided bone  
17 regeneration, although these properties may affect the regenerative outcomes.  
18 There is a wide variability in the pore size and degree of permeability in the  
19 commercially available membranes for GBR, ranging from: micro-porosity (5 – 20  
20  $\mu\text{m}$ ), which may limit the passage of cells, but allows the passage of chemicals,  
21 biomolecules, viruses; moderate porosity (non-resorbable materials  $\leq 100 \mu\text{m}$ ) that  
22 also allows the passage of bacteria, cells and tissue integration/migration (tissue  
23 integration occurs  $\geq 30 - 40 \mu\text{m}$ ); macro-porosity (non-resorbable materials  $> 100$   
24  $\mu\text{m}$ ) which allows unrestricted passage of chemicals, biomolecules, viruses,  
25 bacteria, cells and allows tissue integration and migration. The pore size can  
26 increase during the process of membrane degradation within the tissue, which  
27 may in turn influence its bioactivity, passage of nutrient and cells and the ingrowth  
28 of nutrients and soft and hard tissue cells. The optimal membrane porosity has not  
29 been defined yet.  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42 - *Mechanical properties.* The ideal GBR membrane should be sufficiently rigid for an  
43 adequate space-making capacity and able to withstand the pressure of the  
44 overlying soft tissues during function in order to prevent its collapse. At the same  
45 time. It should possess certain degree of plasticity and elasticity to be easily  
46 contoured and adapted to the anatomy of the defect. In situations where the  
47 membrane does not possess the required mechanical properties, it should be  
48 combined with a bone replacement biomaterial/graft that serves as a scaffold, to  
49 attain the desire volume of regenerated bone.  
50  
51  
52  
53  
54  
55  
56  
57 - *Integration with the tissues.* The integration of the membrane with the adjacent  
58 connective tissues is essential for optimal primary wound closure and healing. In  
59 fact, lack of integration and membrane exposure is associated with inferior  
60

1  
2  
3  
4 regenerative outcomes. There is evidence that mobility of the membrane and lack  
5 of hydrophilicity will impair connective tissue integration and bone formation. In  
6 case of non-resorbable membranes, the degree of tissue integration should be  
7 coupled with an easy and atraumatic removal. There is lack of information on the  
8 optimal degree of membrane tissue integration during healing.  
9  
10  
11

- 12  
13 - *Exposure tolerance.* Membrane exposure and its subsequent bacterial  
14 contamination may hamper the regenerative outcomes irrespectively whether the  
15 membrane is biodegradable or non-resorbable. In case of exposure, the exposed  
16 membrane should be kept in situ and continue to function during the during the  
17 regenerative process, although in case of overt infections, its removal should be  
18 considered. When combined with biomaterials, the incidence of infections might  
19 increase.  
20  
21  
22 - *Biodegradability.* Systematic reviews have shown comparable clinical outcomes  
23 between resorbable and non-resorbable membranes. However, there is clear  
24 evidence that the membrane must retain its function for a certain amount of time to  
25 achieve a predictable regenerative outcome. In fact, the longer the membrane  
26 maintains its function the greater the maturity of the bone is, although in spite of 30  
27 years of GBR research, the ideal membrane bio absorption time has not yet been  
28 established. Moreover, the inflammatory response elicited by the degradation of  
29 the membrane should not adversely affect the regenerative outcome.  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40

### 41 ***What are advantages and limitations of currently available membranes used in*** 42 ***GBR?*** 43

#### 44 *PTFE and modifications*

45

46  
47 Due to its synthetic nature, PTFE membranes have the advantage of not eliciting any  
48 immunological reaction and being resistant to breakdown by the host tissues.

49 Compared with biodegradable membranes, they have superior space making  
50 capability, mainly when these membranes have titanium reinforcement, which makes  
51 them the ideal membranes for vertical bone regeneration. Their main limitation is the  
52 increased frequency of membrane exposure with a subsequent risk for bacterial  
53 contamination and infection, in fact superior regenerative outcomes with these  
54 membranes are associated with a closed uneventful healing. Other limitation is the  
55  
56  
57  
58  
59  
60

1  
2  
3  
4 difficulty in their removal due to their soft tissue integration. Moreover, the cost of  
5 PTFE membranes is higher compared to biodegradable membranes.  
6  
7

### 8 *Synthetic polymers*

9

10 The main advantages of polymeric membranes are their manageability, process  
11 ability, tuned biodegradation, and drug-encapsulating ability. However, their  
12 degradation might elicit a strong inflammatory response, leading to resorption of the  
13 regenerated bone. The resorption rate of these types of membranes is largely  
14 dependent on the type of polymer used.  
15  
16  
17  
18

### 19 *Naturally derived membranes:*

20

#### 21 *Collagen (non-crosslinked)*

22

23 Collagen-based membranes are the most commonly used naturally derived  
24 membranes for GBR and their degradation does not exert any potential deleterious  
25 effect to the tissues. Their use has not been associated with relevant adverse  
26 effects since collagen is the principal component of connective tissues, playing  
27 important role in tissue structural support and in cell matrix communication. Their  
28 main limitation is their lack of rigidity, which limits their space making capabilities and  
29 requires their combination with a scaffold, Yet, collagen membranes can be used  
30 alone for alveolar bone defects which do not require extra fixation and stability such  
31 as bone dehiscence and fenestration defects. Moreover, as their degradation is fast  
32 (approximately four weeks) they may not meet the duration of time required for  
33 optimal tissue regeneration  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43

#### 44 *Chemically modified collagen*

45

46 In order to slow down the bio-absorption process of collagen membranes, a number  
47 of different methods of physical/chemical cross-linking have been developed, which  
48 may also enhance the membrane mechanical properties. Although chemical cross-  
49 linking has resulted in improvement of collagen stability, release of chemicals  
50 residues (e.g. amides or aldehydes) has been associated with severe inflammation  
51 at the implantation site. Generally, the predictability of the collagen membrane not  
52 only depends on the origin of the collagen material but also its preparation and  
53 manufacturing process (de-cellularization, sterilization, and method of cross-linking).  
54  
55  
56  
57  
58  
59  
60

### *Chitosan, Alginate*

Their material properties include biocompatibility, biodegradability, low immunogenicity, and a bacteriostatic effect. Experimental results indicate similar regenerative outcomes when compared with collagen membranes. Despite the experimental studies, their clinical use for GBR has not been documented.

### *Metals*

Titanium is a commonly used material in dentistry. Among its properties are biocompatibility, high strength and rigidity for space maintenance, low density and weight, the ability to withstand high temperatures, and resistance to corrosion. The use of titanium for GBR was inspired from a successful outcome of using a titanium mesh for reconstruction of maxillofacial defects. Titanium mesh alone or with bone substitutes is a procedure for localized alveolar ridge augmentation prior to, or simultaneously with, implant placement. Occlusive titanium and micro-perforated titanium membranes have also been introduced and used for treatment of peri-implant bone defects and ridge augmentation. Limited experimental data exists on CoCr membrane for bone augmentation. Their limitations include difficulties in their removal due to connective tissue integration, mainly associated with the titanium mesh. Conversely, lack of tissue integration has been reported with the use of solid titanium materials.

### ***Which is the role of the exogenous administration of biological cues to the membrane?***

The use of growth factors and/or cell therapies have provided promising experimental results when used in combination with GBR, mainly in combination with resorbable membranes. The evidence of efficacy in clinical trials is, however, lacking and these strategies may be hampered by financial and regulatory constraints as well as for the potential adverse risks associated with these therapies.

### ***Recommendations for Future Research***

The bone promoting environments in the membrane and defect compartment during GBR can likely be optimized by several strategies targeting both material aspects and host-tissue responses.

1  
2  
3  
4 The membrane, the main component of GBR, can be improved depending on the  
5 functional requirements and the involved biological mechanism. These modifications  
6 may include: (i) optimizing the physicochemical and mechanical properties, e.g., the  
7 porosity, structure, thickness, rigidity and plasticity; (ii) incorporating biological  
8 factors and synthetic bioactive materials (iii) incorporating antibacterial agents and  
9 antibiotics.

10  
11  
12  
13  
14  
15 From scientific, developmental and clinical perspectives, new developments in tissue  
16 engineering and drug delivery may enhance the barrier concept associated with  
17 GBR and expand the clinical opportunities for bone regeneration in the future.  
18  
19  
20  
21

## 22 **Bioactive factors used in regenerative interventions in the** 23 **carionaxillo-facial region to enhance bone regeneration** 24 25

26  
27 Bioactive agents or factors are defined as natural mediators of tissue repair capable  
28 of eliciting a response from a living tissue, organism or cell. The majority of pre-  
29 clinical and clinical studies on bone regeneration have focused on Bone  
30 Morphogenetic Proteins (mainly BMP-2) and Autologous Platelet Concentrates  
31 (APCs). Less evidence is available for other growth factors (mainly Platelet Derived  
32 Growth Factor PDGF-BB, Fibroblast Growth Factor FGF-2 and Vascular Endothelial  
33 Growth Factor VEGF) and amelogenins. The combination of different bioactive  
34 factors has also been proposed, with the aim to reduce the dosages of each factor  
35 (and associated side effects) and, at the same time, promote synergistic effects.  
36 While significant literature has documented the use of bioactive factors (mainly  
37 amelogenins, FGF-2, PDGF-BB and APCs) for the regeneration of periodontal  
38 intrabony defects, it is outside the remit of this consensus to comment on periodontal  
39 regeneration.  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49

### 50 ***What are advantages of the use of bioactive factors in bone regeneration?*** 51

52 The consensus was based on a critical review assessing the outcomes of the use of  
53 bioactive substances in preclinical models and clinical applications. The preclinical  
54 bone regeneration models included ridge/socket preservation, alveolar ridge  
55 augmentation (horizontal and vertical), regeneration of bone defects at the moment  
56 of implant placement, sinus augmentation and regeneration of critical/sub-critical  
57  
58  
59  
60



1  
2  
3  
4 bone defects. The clinical evidence is based only on RCTs, CCT and Case Series (>  
5 5 cases) that included histological and/or radiographical assessment of bone  
6 regeneration for ridge preservation, ridge augmentation, regeneration of bone  
7 defects at the moment of implant placement and sinus augmentation.  
8  
9

### 10 11 *Bone morphogenetic protein (rh-BMP-2)* 12

13  
14 Overall, pre-clinical studies suggest that rhBMP-2 (at different dosages) significantly  
15 promotes, either directly or indirectly, bone regeneration in critical and sub-critical  
16 size bone defects and *de novo* bone formation regardless of the carrier adopted.  
17 rhBMP-2 enhances ridge augmentation in chronic and combined defects and  
18 promotes ridge preservation. Conflicting results have been reported regarding the  
19 benefits in peri-implant circumferential defects and sinus augmentation. In  
20 challenging ridge augmentation or peri-implant defects the combination with a space  
21 providing material is recommended.  
22  
23  
24  
25  
26

27  
28 As a carrier, the Absorbable Collagen Sponge (ACS) can be successfully used, with  
29 or without space-providing materials. For clinical application, an absorbable collagen  
30 sponge carrier (ACS) impregnated with rhBMP-2 was approved by the Food and  
31 Drug Administration for ridge preservation and sinus augmentation. Therefore, most  
32 of the clinical studies have employed BMP-2/ACS, although a combination of  
33 rhBMP-2 with different grafts has also been suggested. Clinical studies suggest 1.50  
34 mg/ml as the optimal dosage for ridge preservation and a range between 1.05 and  
35 4.2 mg/ml for ridge augmentation purposes, while in some studies on sinus  
36 augmentation high supra-physiological doses of up to 48mg of BMP-2 per subject  
37 have been reported.  
38  
39  
40  
41  
42  
43  
44  
45

46 Based on 3 RCTs, rhBMP-2/ACS combined with osteoconductive grafts and/or a  
47 titanium mesh for ridge augmentation is comparable to autologous bone and titanium  
48 mesh or Deproteinized Bovine Bone Mineral based on radiographic/histological  
49 outcomes. In a recent RCT of 4 months duration, the use of autologous block grafts  
50 was superior in terms of amounts of mineralized tissue when compared to DBBM  
51 block grafts loaded with BMP-2. 3 RCTs have used BMP-2 combined with ACS or  
52 other carriers for ridge preservation. The evidence of using rhBMP-2 in regeneration  
53 of bone defects following implant placement is scarce. The results of the studies  
54 using rhBMP-2 as a graft material in sinus floor augmentation are conflicting.  
55  
56  
57  
58  
59  
60

1  
2  
3  
4 Amongst the available bioactive factors BMP-2 is supported with the highest  
5 evidence, albeit heterogeneous. The existing RCTs suggest that there is a similar  
6 beneficial effect of rhBMP-2/ACS compared to commercially available bone grafting  
7 materials for socket preservation and ridge augmentation. Currently, this material  
8 has not been approved in Europe for clinical use in oro-maxillofacial applications.  
9  
10

#### 11 *Other growth factors (PDGF-BB, FGF-2, VEGF)*

12  
13 Direct administration of PDGF-BB, FGF-2 and VEGF with different carriers and  
14 indirect administration has been studied in pre-clinical studies with demonstration of  
15 enhanced bone regeneration when used with GBR. The available evidence for their  
16 use for ridge preservation is not robust enough to draw conclusions and make  
17 recommendations  
18  
19

20  
21 Studies in chronic alveolar defects have shown that rhPDGF-BB combined with  
22 block or particulate grafts can significantly promote ridge augmentation. It is unclear  
23 whether the addition of a barrier membrane has an impact on the final regeneration  
24 outcome. The pre-clinical evidence that rhFGF-2 associated with different synthetic  
25 biomaterials can promote ridge augmentation is still not robust. There is limited  
26 evidence from experimental studies suggesting that rhPDGF-BB alone or combined  
27 with IGF-1 or bone grafts might enhance the regeneration of peri-implant defects.  
28 There is insufficient evidence on the use of growth factors other than BMP-2 for  
29 sinus augmentation.  
30  
31

32  
33 rhPDGF-BB and rhGDF-5 are the growth factors, apart from rhBMPs, which have  
34 been evaluated in clinical studies for bone regeneration when combined with  
35 different bone replacement grafts. The available studies have used 0.5 mL PDGF-BB  
36 at concentration of 0.3 mg/ml, and between 500 µg to 500 mg rhGDF-5 per gram of  
37 β-TCP. There is limited evidence on the efficacy of rhPDGF-BB for ridge  
38 preservation and ridge augmentation. RCTs are needed to clarify whether rhPDGF-B  
39 combined with different grafting materials can promote post-extraction ridge  
40 preservation and the horizontal and vertical regeneration of alveolar defects. No  
41 controlled studies have investigated the use of rhPDGF-BB for the treatment of peri-  
42 implant defects and there are only limited studies reporting on the use of rhPDGF-BB  
43 or rhGDF-5 for sinus augmentation. Therefore, no robust conclusions could be  
44 drawn.  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

### *Enamel matrix derivatives (EMD)*

Although various fractions of EMD have shown osteoinductive properties, only few pre-clinical studies have investigated the use of amelogenins (EMD) for bone regeneration. EMD has limited effect in enhancing bone formation and does not offer significant advantages over the use of a membrane or a bone graft or the combination of both.

The available clinical evidence does not support the use of EMD in sinus augmentation, ridge augmentation, ridge preservation or bone defects following implant placement.

### *Autologous platelet concentrates (APCs)*

APCs are intended to enhance bone regeneration by triggering the natural healing process with a supplement of highly concentrated bioactive factors. One of the main challenges in reviewing the scientific evidence is the heterogeneity of the APC preparation protocols and accuracy in the use of the terminology.

Some pre-clinical studies have shown that Platelet Rich Plasma (PRP) or Platelet Rich Fibrin (PRF) compared to spontaneous healing, fibrin glue derived from Platelet Poor Plasma, collagen sponge or hydrogel improve the amount and quality of regeneration of experimentally induced bone defects. However, there is no robust evidence supporting the addition of these APCs to bone grafts or for *de novo* bone formation.

The evidence on the potential of Plasma Rich in Growth Factors (PRGF) to promote bone regeneration in experimentally induced bone defects is limited. Regarding the use of all APCs, there is insufficient evidence for ridge/socket preservation, regeneration of bone defects after implant placement and sinus augmentation. The limited pre-clinical studies on the use of PRP for ridge augmentation suggest that, while autologous bone remains the gold standard, the combination of PRP, cells and different bone substitutes could be a promising alternative.

Clinical research has shown that the combined therapy of APCs with bone grafts and/or cells offer promising results for ridge augmentation procedures. More RCTs are needed to clarify which of the APCs is superior. For ridge preservation

1  
2  
3  
4 procedures, APCs may accelerate clinical healing, soft tissue epithelialization and  
5 reduce postoperative pain, but there is insufficient and contrasting evidence of a  
6 significant effect on hard tissue regeneration. The clinical effect of APCs on defects  
7 after implant placement has been studied in a limited number of investigations but  
8 the available evidence does not allow for robust conclusions. Conflicting outcomes in  
9 terms of bone formation and implant stability emerged from the available studies on  
10 the use of APCs for sinus augmentation, with no clear benefits of one APC over the  
11 other.  
12  
13  
14  
15  
16  
17

### 18 **Which are the limitations of the use of growth factors for bone regeneration?**

19  
20  
21 When evaluating the application of growth factors for bone regeneration, there is lack  
22 of a clear understanding of their mechanism of action, the ideal dosage, frequency  
23 and mode of administration and the delivery system. Furthermore, there is a clear  
24 need for developing standardized protocols for controlling their release and  
25 clearance at the application site. In general, despite some encouraging results, the  
26 available evidence does not support the use of bioactive factors as a routine  
27 alternative to the currently used bone regenerative interventions in the caniomaxillo-  
28 facial area.  
29  
30  
31  
32  
33  
34

35 High dosages of rhBMP-2 have been associated with side effects, including long  
36 lasting oedema formation, as well as mucosal erythema, osteoclast-mediated bone  
37 resorption and inappropriate adipogenesis. Combinations of rhBMP-2 with other  
38 growth factors (preclinical studies) have been suggested to reduce these dosage-  
39 related complications of rhBMP-2.  
40  
41  
42  
43

44 Moreover, a concern that could be raised on the use of rhBMP-2 is the development  
45 of immunological factors, such as anti-rhBMP-2 and anti-bovine collagen type I (6%  
46 and 20% incidence, respectively, as per FDA report on rhBMP-2/ACS).  
47  
48  
49  
50

### 51 ***Recommendations for Future Research***

52  
53 Well-designed and adequately powered RCTs showing clinical and histological  
54 outcomes of bioactive agents are required to clarify their potential and actual need in  
55 regenerative dentistry.  
56  
57  
58  
59  
60

1  
2  
3  
4 In the future, studies need to be designed to overcome the heterogeneity currently  
5 present in the literature concerning biological agent's dosage, formulation,  
6 concomitant biomaterials used, types of defects, methods of investigation and follow-  
7 up periods.  
8  
9

10  
11 Research efforts should be also directed towards the development of delivery  
12 systems enabling controlled spatio-temporal delivery of single or combination of  
13 bioactive factors. The ultimate aim should be mimicking the synergistic wound  
14 healing activity of the combinational release profiles of growth factors and  
15 extracellular matrix components that occurs in physiological wound healing.  
16  
17  
18  
19  
20

## 21 22 23 **Cell therapies used in regenerative interventions in the** 24 **carionaxillo-facial region to enhance bone regeneration** 25

26  
27 This consensus report is based on the review of the evidence from pre-clinical and  
28 clinical studies on the use of cell therapies for cranio-maxillofacial bone regeneration.  
29 Three main approaches using cell therapies have been evaluated: a) "*minimally*  
30 *manipulated*" whole tissue fractions; b) *ex-vivo* expanded "*uncommitted*"  
31 stem/progenitor cells, and c) *ex-vivo* expanded "*committed*" bone-/periosteum-derived  
32 cells. Minimally manipulated whole tissue fractions, preserve the physiological  
33 microenvironment or 'niche' of multiple cell types in their natural ratios; these mainly  
34 include bone marrow aspirates – either whole (BMA) or concentrated (BMAC), adipose  
35 stromal vascular fractions (A-SVF), and tissue "micrografts". The major limitation of  
36 this approach is that mesenchymal stem (and progenitor) cells (MSCs) represent a  
37 very limited fraction of the implanted cells.  
38  
39  
40  
41  
42  
43  
44  
45

46  
47 *Ex vivo* expansion strategies exponentially increase the number of cells of a specific  
48 phenotype, i.e., *uncommitted* or *committed*, available for transplantation. The most  
49 commonly used source of *uncommitted* MSCs is the bone marrow (BMSCs), but more  
50 recently less invasive sources such as adipose tissue (ASCs) and dental tissues, have  
51 been tested. Sources of *committed* cells are the periosteum and cancellous  
52 bone/marrow of the alveolar bone itself. The major limitation of *ex vivo* expansion  
53 strategies is the need for highly sophisticated laboratories according to Good  
54  
55  
56  
57  
58  
59  
60

1  
2  
3  
4 Manufacturing Practices (GMP), thereby significantly increasing the cost of this  
5 therapy.  
6  
7

8 **What is the effect of cell therapies (either whole tissues or ex vivo expanded**  
9 **cells) added to biomaterials or autologous bone, or combinations thereof, when**  
10 **compared with biomaterials or autologous bone alone, or combinations thereof,**  
11 **for the different clinical indications in the craniomaxillo-facial area?**  
12  
13  
14

15 Clinical studies have evaluated cell therapies, mainly BMA and BMAC, in combination  
16 with biomaterials or autologous bone, or combinations thereof vs. relevant controls in  
17 various indications (e.g. sinus augmentation, horizontal ridge augmentation, alveolar  
18 cleft defects) and at various observation times. The clinical evidence is mostly based  
19 on randomized (sinus and ridge augmentation) and non-randomized controlled trials  
20 (alveolar cleft repair).  
21  
22  
23  
24  
25

26 Specifically:  
27  
28

- 29 a) in sinus augmentation, significantly more bone regeneration was observed after cell  
30 therapy in 1 meta-analysis of histomorphometric results (6 studies, vs. scaffolds, 6  
31 months) and in 1 meta-analysis of micro-computed tomography ( $\mu$ -CT) results (3  
32 studies, vs. scaffolds, 4-7 months), while in 1 meta-analysis of histomorphometric  
33 results no benefit was observed (12 studies, vs. scaffolds, 3-4 months). Based on  
34 a meta-regression analysis of histomorphometric data from 15 studies, there were  
35 no differences between the various cell therapy strategies, i.e. whole tissues vs.  
36 expanded *uncommitted* cells vs. expanded *committed* cells, in terms of the amount  
37 of bone regeneration.  
38  
39  
40  
41  
42  
43  
44  
45 b) in horizontal ridge augmentation, significantly more bone regeneration was  
46 observed after cell therapy in 1 meta-analysis of histomorphometric results (3  
47 studies, vs. scaffolds; 1 study, vs. scaffolds + autogenous bone, 4-6 months).  
48  
49  
50  
51 c) in alveolar cleft defects, 1 meta-analysis failed to show a benefit of cell therapy over  
52 autogenous bone, as evaluated with CT (3 studies, 6 months).  
53  
54  
55  
56 d) limited clinical evidence suggests that the 'conditioned medium' or 'secretome' from  
57 MSCs may promote bone regeneration in sinus augmentation [1 study (4 patients),  
58 vs. scaffold, 6 months].  
59  
60

1  
2  
3  
4 The above observations are in general supported by the results of meta-analyses of  
5 pre-clinical *in vivo* studies in large animals, which have mainly evaluated *ex vivo*  
6 expanded “uncommitted” cells, mainly BMSCs, in combination with biomaterials vs.  
7 relevant controls in various models (e.g. sinus augmentation, critical size defects,  
8 alveolar cleft defects) and platforms (e.g. dogs, pigs, sheep/goats) at various  
9 observation times.  
10  
11  
12  
13  
14

15 Based on the data from the clinical studies, the following conclusions may be derived:  
16

- 17 1. Transplantation of cells, most commonly whole BMA or BMAC, in combination with  
18 biomaterial scaffolds results in superior bone regeneration compared to  
19 implantation of scaffolds alone in sinus augmentation and horizontal ridge  
20 augmentation, and comparable bone regeneration to autogenous bone in alveolar  
21 cleft repair.  
22  
23  
24  
25
- 26 2. Based on studies of sinus augmentation, no superiority of *ex vivo* expanded cells,  
27 either *uncommitted* or *committed*, over whole tissue fractions (BMA/C or A-SVF)  
28 was observed. However, the appropriateness of sinus augmentation as a model to  
29 test cell therapies and detect clinically relevant benefits may be questioned, owing  
30 to the “self-healing” capacity in this site (Duan, 2017).  
31  
32  
33  
34  
35
- 36 3. The analyzed studies, both clinical and pre-clinical, showed a wide range of effect  
37 sizes and prediction intervals, suggesting a high degree of heterogeneity, and  
38 emphasizing the need for well-designed future studies to ascertain the true effect  
39 of cell therapies.  
40  
41  
42

### 43 **Is osteogenic pre-differentiation in *ex vivo* expansion strategies beneficial?**

44  
45 Based on limited evidence from pre-clinical *in vivo* and uncontrolled clinical studies,  
46 use of pre-differentiated BMSCs has not demonstrated a significant added effect in  
47 terms of enhancing bone regeneration compared with using undifferentiated BMSCs,  
48 while osteogenic pre-differentiation of ASCs or the addition of osteoinductive factors,  
49 e.g., BMP-2, seemed to enhance bone regeneration. However, no included studies in  
50 the review (pre-clinical or clinical) directly compared pre-differentiated cells vs.  
51 undifferentiated cells, either BMSCs or ASCs.  
52  
53  
54  
55  
56

57  
58 Based on this limited evidence:  
59  
60

1  
2  
3  
4 a) In BMSCs, osteogenic pre-differentiation may not show any additional beneficial  
5 effect.  
6

7 b) In ASCs, additional osteogenic stimulation, via pre-differentiation or addition of  
8 osteoinductive factors, e.g., BMP-2, may be beneficial.  
9  
10

### 11 12 **Recommendations for Future Research** 13

14  
15 The relatively large effect sizes in favor of cell therapy observed in pre-clinical *in vivo*  
16 studies are diminished in clinical trials, suggesting a gap in translation and the need  
17 for better pre-clinical models.  
18  
19

20  
21 Vascularization remains a key challenge in cell therapy, especially in large defects,  
22 since an inadequate vascularization in the internal parts of the cell-scaffold construct  
23 can impair cell survival and thereby compromise clinical outcomes. Promising pre-  
24 clinical research has been conducted in the area of 'pre-vascularized' bone constructs  
25 but remains to be translated clinically.  
26  
27  
28

29  
30 The potential of using allogeneic cells as an "off-the-shelf" therapy has been tested  
31 with favorable results in a limited number of pre-clinical studies. However, the  
32 possibilities of immune reactions associated with using allogeneic human cells are still  
33 unclear and require further investigation (Kiernan, 2018).  
34  
35  
36

37  
38 The potential use of 'cell-free' strategies, which exploit the paracrine or trophic effects  
39 of MSC-secretomes to promote regeneration, should be explored. Similarly,  
40 alternative mechanisms of MSC activity, for example, via "empowerment" of host cells  
41 and modulation of immune cells in the context of bone regeneration should be  
42 investigated.  
43  
44  
45

46  
47 There is a need to evaluate the cost-effectiveness of cell therapy in comparison to  
48 current standards of care. Moreover, there is a need of well-designed studies to  
49 evaluate the efficacy/cost-effectiveness of different cell therapy strategies, i.e., whole  
50 tissues vs. expanded *uncommitted* cells vs. expanded *committed* cells.  
51  
52  
53

### 54 55 **Acknowledgements:** 56 57 58 59 60



1  
2  
3  
4 The authors acknowledge the contribution of the representative from the Osteology  
5 Foundation, Benjamin Muller and the sponsor (Geistlich Pharma), Niklaus Steifel for  
6 their contribution during the workshop.  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

**References:**

Albrektsson, T. & Johansson, C. (2001) Osteoinduction, osteoconduction and osseointegration. *European Spine Journal* 10 Suppl 2, S96-101.  
doi:10.1007/s005860100282.

Donos, N., Dereka, X., and Calciolari, E. (2019). The use of bioactive factors to enhance bone regeneration. A narrative review *Journal of Clinical Periodontology, Spec Issue*

Duan, D.H., Fu, J.H., Qi, W., Du, Y., Pan, J., Wang, H.L. (2017) Graft-Free Maxillary Sinus Floor Elevation: A Systematic Review and Meta-Analysis. *Journal of Periodontology*, 88, 550-564.

Haugen, H.J., Lyngstadaas, S.P., Rossi, F., and Perale, G, (2019). Bone grafts: which is the ideal biomaterial? *Journal of Clinical Periodontology, Spec Issue*

Hutmacher, D. W. (2000) Scaffolds in tissue engineering bone and cartilage. *Biomaterials*, 21, 2529-2543. doi:10.1016/S0142-9612(00)00121-6.

Kiernan, C.H., Wolvius, E.B., Brama, P.A.J., Farrell, E. (2018). The Immune Response to Allogeneic Differentiated Mesenchymal Stem Cells in the Context of Bone Tissue Engineering. *Part B Reviews*, 24, 75-83.

Omar O., Elgali, I, Dahlin, C., and Thomsen, P. (2019). Barrier membranes: more than the barrier effect? *Journal of Clinical Periodontology, Spec Issue*

Shanbhag, S., Suliman, S., Pandis, N., Stavropoulos, A., Sanz, M., and Mustafa K (2019) Cell therapy for orofacial bone regeneration: a systematic review and meta-analysis. *Journal of Clinical Periodontology, Spec Issue*

Wang, Y, Chen, X, Cao, W, Shi, Y. (2014) Plasticity of mesenchymal stem cells in immunomodulation: pathological and therapeutic implications. *Nature Immunology* 15, 1009-16.

Williams, D. F. (2017) Biocompatibility Pathways: Biomaterials-Induced Sterile Inflammation, Mechano-transduction, and Principles of Biocompatibility Control. *ACS Biomaterials Science & Engineering* 3, 2-35.