

Title: Interactions between plant genome size, nutrients and herbivory by rabbits, molluscs and insects on a temperate grassland

Authors: Maïté S. Guignard^{1,2}, Michael J. Crawley³, Dasha Kovalenko¹, Richard A. Nichols¹, Mark Trimmer¹, Andrew R. Leitch¹, and Ilia J. Leitch²

Author affiliations:

¹ School of Biological and Chemical Sciences, Queen Mary University of London, Mile End Road, London, E1 4NS, UK

² Royal Botanic Gardens, Kew, Richmond, Surrey TW9 3DS, UK

³ Faculty of Natural Sciences, Department of Life Sciences, Silwood Park, Imperial College London, UK

Corresponding authors:

Andrew R Leitch: a.r.leitch@gmul.ac.uk. School of Biological and Chemical Sciences, Queen Mary University of London, Mile End Road, London, E1 4NS, UK. tel: 0207 882 5294

Ilia J Leitch: i.leitch@kew.org. Royal Botanic Gardens, Kew, Richmond, Surrey TW9 3DS, UK. tel: 0208 332 5329

ABSTRACT

Angiosperm genome sizes (GS) vary c. 2,400-fold. Recent research has shown that GS influences plant abundance, and plant competition. There are also tantalising reports that herbivores may select plants as food dependent on their GS. To test the hypothesis that GS plays a role in shaping plant communities under herbivore pressure, we exploit a grassland experiment that has experimentally excluded herbivores and applied nutrient over 8 years. Using phylogenetically-informed statistical models and path analyses, we show that under rabbit-grazing, plant species with small GS generated the most biomass. In contrast, on mollusc and insect-grazed plots, it was the plant species with larger GS that increased in biomass. GS was also shown to influence plant community properties (e.g. competitive strategy, total biomass) although the impact varied between different herbivore guilds (i.e. rabbits versus invertebrates) and nutrient inputs. Overall, we demonstrate that GS plays a role in influencing plant-herbivore interactions, and suggest potential reasons for this response, which include the impact of GS on a plant's response to different herbivore guilds, and on a plant's nutrient quality. The inclusion of GS in ecological models has the potential to expand our understanding of plant productivity and community ecology under nutrient and herbivore stress.

Key words: Genome size, herbivory, plant community ecology, competition, nitrogen, grassland experiment.

2 1. INTRODUCTION

3 Terrestrial ecological communities are shaped by interactions between plants and herbivores, in
4 which the availability of resources play a central role [e.g. 1]. These interactions and their
5 components are driven by a number of biotic and abiotic, top-down and bottom-up factors [2], one
6 of which is nutrient availability, where a near-universal pattern following nutrient enrichment is an
7 increase in plant biomass and a decrease in plant species diversity [3, 4]. Nutrient availability also
8 mediates the impact of herbivores on plant biomass and on community structure. For example,
9 plants with access to nutrient resources are better able to tolerate herbivory [5]. In addition, grazing
10 pressure on fertilized plant communities is associated with increased plant functional diversity [6].
11 Herbivores can counteract a decrease in plant species loss following nutrient enrichment by keeping
12 fast-growing plants in check and promoting the growth of less competitive, but better defended taxa
13 [7, 8]. Conversely, higher nutrient concentrations promote investment in plant growth rather than
14 plant defences, increasing leaf palatability [9]. The production of enzymes and other proteins in
15 growing plant tissues can also increase nitrogen (N) and phosphorus (P) concentrations, and such
16 nutritious tissues are often favoured by herbivores, especially during the growing season [e.g. 10].
17 Numerous studies have shown that certain plant functional traits can influence and mediate plant
18 responses to stress caused by herbivory [e.g. 11, 12], and since plants form the basis of terrestrial
19 food chains, any factor that can influence plant abundance and productivity has implications for all
20 trophic levels. One factor that has been little explored in considering plant herbivore interactions is
21 plant genome size (GS; i.e. the amount of DNA in the unreplicated haploid nucleus – 1C-value),
22 which ranges c. 2,400-between plant species [13]. Genome size can impact a wide diversity of plant
23 traits, influencing how and where plants grow and interact across different ecosystems [14]. It may
24 also impact how plant communities respond to grazing pressure, and plant-herbivore interactions.
25 Certainly, there are tantalising reports which suggest that GS can play a role in herbivore choice,
26 although the responses are variable, e.g. some herbivorous insects favour polyploid over diploid
27 cytotypes of the same species [15-18] or vice versa [19]; and cows may graze preferentially on a
28 tetraploid versus diploid grass cytotype [20]. There are several reasons as to why plant GS might play

29 such a role in plant herbivore interactions. For example, plant species with very large genomes (e.g.
30 1C-values ≥ 25 pg (1 pg=978 Mbp) are slow growing, obligate perennials, perhaps due to longer DNA
31 replication times [21]. Such plants are likely to recover poorly following herbivory compared with
32 faster growing, more competitive species with smaller GSs. In addition, GS can influence leaf
33 stoichiometry, as GS is positively correlated with leaf N concentrations [22], which could influence
34 herbivore preference.

35 This paper explores the hypothesis that there are interactions between plant GS and herbivory that
36 influence plant community composition and dynamics. We predict that plant communities under
37 grazing pressure are primarily composed of species with smaller GS in contrast to communities
38 where herbivores have been experimentally excluded, because tissue recovery from herbivore
39 damage may be slower and costlier to produce in taxa with larger genomes and/or plants with larger
40 genomes are nutritionally favoured by herbivores. We take advantage of an ongoing long-term
41 ecological experiment which includes experimental manipulations of herbivory and nutrients (N and
42 P). The experiment was established in 1992 at Nash's Field, Silwood Park, UK, on an acid
43 mesotrophic grassland with intense herbivory by rabbits (*Oryctolagus cuniculus*) which are a
44 keystone species. Our results reveal that GS clearly plays a key role in influencing interactions
45 between herbivores, nutrients, and plant biomass production. However, we also show that the
46 direction of these interactions is dependent on the type of herbivore guild, with analyses at the
47 plant community level revealing contrasting interactions and dynamics imposed by rabbit versus
48 invertebrate (mollusc and insect) herbivory.

49 **2. METHODS**50 **(a) Study site**

51 The experimental study was started in 1992 on Nash's Field in Silwood Park, UK (National Grid
52 reference 41/944691). Rabbits have been present at this site since their recovery from myxomatosis
53 in the 1950s, and their grazing has prevented the establishment of woody species (e.g. *Quercus*
54 species) and the succession from grassland to woodland. The experiment is set up in a split-plot,
55 factorial design, comprising a total of eight herbivore exclusion treatments (\pm insect \times \pm mollusc \times \pm
56 rabbits) (Fig. SI.1). Each herbivore exclusion block (22m²) is further divided in half with pH-
57 controlled (limed at pH = 7, and unlimed at pH = 4.1) plots. In the first three years of the experiment,
58 the limed plots also received one of three herbicide treatments: –grasses, –forbs, and control. At
59 the smallest plot level (4m²) are the nutrient treatments, which comprise 12 combinations of:
60 \pm nitrogen (N) as ammonium nitrate (100 kg ha⁻¹), \pm phosphate (P) (35 kg ha⁻¹), \pm potassium (K), and
61 \pm magnesium (Mg), all of which are added once a year. Insects are controlled by a permethrin
62 synthetic pyrethroid and dimethoate-40; molluscs (snails and slugs) by metaldehyde pellets; and
63 rabbits by wire mesh fencing [23]. Insecticide and molluscicide are applied three times a year. Small
64 mammals such as field voles (*Microtus agrestis*) and large mammals such as roe deer (*Capreolus*
65 *capreolus*) are not excluded by rabbit fencing. In the first three years, herbicide was also applied
66 within each herbivore plot for plant type control (\pm grasses \pm forbs). The natural plant community is
67 an intensely rabbit-grazed sward dominated by perennial C3 grasses, primarily *Festuca rubra*, but
68 other common species include *Agrostis capillaris*, *Holcus lanatus*, *Arrhenatherum elatius* (Poaceae),
69 and *Jacobaea vulgaris* (Asteraceae). We based our investigations on data from 190 limed, herbicide-
70 free plots in the species-level analyses, and 556 limed plots in the plant community analyses.

71 We focus on limed plots rather than the acidic (pH 4.1) unlimed plots to avoid the confounding
72 effects that would result from interactions between fertilization, in particular N, soil acidity, and
73 aluminium toxicity [24]. In addition, the analysis of limed plots enables comparisons with the only

74 other analyses published on interactions between fertilizers and genome size in the generation of
75 biomass [40, 47], which are both also from temperate grassland field sites.

76 **(b) Data collection**

77 For analyses, we used biomass data collected in 1997 and 2000 from the 556 limed 4m² subplots; we
78 excluded 20 plots containing *Pteridium aquilinum* (an invasive fern) to focus on angiosperm plant
79 communities. Where possible, species were sampled in 2015 to estimate their GS (1C-value) using
80 standard flow cytometry methods [25, see Table SI.1 for details]. GS were estimated from one to
81 eight individuals of 36 species collected from Nash's Field, with species with known cytotypes
82 (different ploidy levels) being more extensively sampled. We used our own GS estimates when the
83 coefficients of variations were <5% in the flow cytometry analysis, and obtained the remaining 1C-
84 values from the prime values given in the Plant DNA C-values database [26] (Table SI.2). No infra-
85 specific differences in ploidy level were found.

86

87 **(c) Phylogenetic data**

88 We pruned the DaPhnE phylogenetic tree [27], to include the 56 species present in the data with the
89 R package *ape* [28]. We tested whether assumptions of a Brownian motion model were met (e.g.
90 branch length is proportional to amount of variation) with the *caper* package [29]. The most
91 appropriate tree was one in which the branch lengths were transformed using the cladogram option
92 in FigTree (<http://tree.bio.ed.ac.uk/software/figtree/>, version 1.4.3). We used this tree to account
93 for non-independent evolutionary relationships among species in the statistical analyses (Fig. SI.2).
94 Phylogenetic signal in log-transformed 1C-values was significant (Pagel's lambda=0.761, p <0.0001,
95 Blomberg's K=0.502, p= 0.001; estimated with *phytools* [30]).

96

97 **(d) Data analysis**

98 We first tested the effect of GS and experimental treatment on plant above-ground biomass at the
99 species level. We then tested the effects of herbivory and macronutrient input on mean GS at the

100 community (plot) level. Finally, we carried out path analyses to explore dynamics between plant
 101 community properties (*sensu* [31]): community-weighted mean (CWM) GS, CWM C-strategy (see
 102 below), total biomass, and phylogenetic diversity, under the experimental treatments. Statistical
 103 analyses were carried out in R version 3.3.3 [32].

104 We refer to the C-strategy of a species as the competitor strategy in Grime's C - S (stress tolerant) - R
 105 (ruderal) plant strategies [33]. Each species is attributed to one, or more often, to a combination of
 106 these strategies (following [34]), based on how a species persists in its natural habitat, and where
 107 the sum of these values is equal to one (Table SI.2).

108

109 **(i) Species level analyses**

110 We analysed the effect of GS, herbivore exclusion, and nutrient treatment on plant biomass at the
 111 species level using phylogenetic generalised linear mixed models (PGLMM) with Bayesian
 112 estimation, by fitting Markov chain Monte Carlo generalised linear mixed models from the R package
 113 *MCMCglmm* [35]. Ideally, we would have analysed all the species in the model here, but this was not
 114 possible because many species were very rare, and in an experiment containing 556 plots, this gave
 115 a dataset with many absences (zeros). Statistically, comparisons of treatment effects on species
 116 biomass cannot be made on species with such few occurrences, preventing us from testing whether
 117 the treatment is having an effect versus whether species occurrence is random. Consequently, and
 118 after excluding plots where herbicides had been applied, (i) we limited the analysis to those species
 119 that occur on two or more herbivore treatments. This criterion still left the data too zero inflated for
 120 analysis because so many species were so uncommon that rounding to the nearest tenth of a
 121 percent equaled 0 (see Table SI.12). Therefore: (ii) we limited the analysis to the most common
 122 species that had at least 1% of biomass on two or more plots. Together these two criteria did enable
 123 the model to converge, leaving for analysis a total of $n = 12$ species analysed over 190 plots. The
 124 results, whilst being restricted to the common species, generated data that support, and are fully

125 congruent with, the community level analyses in the forthcoming sections, where all species, their
126 genome sizes and biomass data are considered together in the same models.

127 We analysed species biomass in two-parts, similar to a hurdle model. Species presence/absence
128 were first fitted in a logistic model, testing the probability of a species occurring on a plot, given the
129 experimental treatments. The second part uses biomass data and fits only the non-zero biomass of
130 the 12 species. The biomass data were log-transformed and analysed using MCMCglmm, assuming a
131 Gaussian distribution. The exclusion of insects, molluscs, rabbits, and the application of N, P, and K
132 fertilizer were scored as binary factors, with untreated plots coded as the reference levels. Genome
133 size was centred on the median GS of the 12 species. Evolutionary non-independence was controlled
134 for by specifying a correlation matrix estimated from the phylogeny. Random effects were specified
135 as: block + block x fencing + species + phylogeny.

136 We first tested four-way interactions between GS, N, P, and herbivore exclusion. Phosphorus
137 showed no significant interactions with herbivore exclusion, consequently, models were re-fitted
138 without P in these interactions. Similarly, Mg had no effect on species occurrence and biomass and
139 was removed to simplify the model. Fixed effects were thus specified as: (GS x insecticide x N) + (GS
140 x molluscicide x N) + (GS x fencing x N) + (GS x N x P) + (molluscicide x insecticide x fencing) + K. We
141 used priors where nu=0.002 and variance=1 [35] and ran the model with 2.5 million generations
142 including a burn-in of 30,000 and a thinning interval of 500. We ran three chains for each model and
143 assessed multiple chain convergence and trace and autocorrelation plots [36].

144

145 **(ii) Community level analyses**

146 We investigated the effects of GS, herbivore exclusion, and nutrient treatment at the community
147 level. Each plot is representative of a plant community growing under various combinations of
148 nutrient availability and herbivore guilds. For each plot (n = 556), CWM GS was estimated using the
149 phylogenetic generalized least squares method (PGLS). We fitted regressions with a Brownian
150 motion correlation structure derived from the phylogeny [37], and maintained the same

151 phylogenetic correlation structure across all plots. Species GS was log-10 transformed, and species
152 percent biomass was used for weighting: $gls(\log_{10} GS \sim 1, cor = corBrownian(phylogeny), weights =$
153 $varFixed(\sim 1/\text{species biomass}))$. We then back-transformed this mean, for use in the subsequent
154 analyses and figures, to facilitate interpretation. To assess whether CWMGS is a function of
155 herbivory and nutrient application, we fitted linear mixed effect (LME) models with the *lme4*
156 package [38] where each herbivore type (insect, mollusc, rabbit) and each nutrient (N, P, K, Mg),
157 were scored as binary factors, and with random effects reflecting the split-plot design
158 (plot/fencing/herbicide). Herbicide treatment was also included as a fixed effect, to account for its
159 application in the early years of the experiment. Interactions between herbivore and nutrient
160 treatments and the significance of each factor were tested using maximum likelihood (ML) stepwise
161 model reduction methods, and the final most reduced model refitted with restricted ML (REML).
162 Estimations of parameter significance (p-values) were obtained with the *lmerTest* package [39].

163

164 **(iii) Confirmatory Path Analysis**

165 To determine how herbivore guilds, nutrient availability, and GS impact plant communities, we
166 examined the effects of N, P, K and: 1) rabbit exclusion, and 2) mollusc and insect exclusion on plant
167 community structure using confirmatory path analysis. Specifically, we tested (i) whether GS
168 influenced the abundance of plants with a C-strategy (*sensu* Grime [33]) at the community level [e.g.
169 40]; (ii) how the abundance of plants with a C-strategy impacts total biomass and phylogenetic
170 diversity; and (iii) the role of herbivory and nutrients on these community properties. Data were
171 partitioned into two datasets: 1) unfenced plots and fenced plots (rabbit exclosures) to test the
172 effect of rabbit exclusion (n=144 plots without insecticide or molluscicide treatment), and 2) \pm insect
173 and \pm mollusc treatment (within rabbit exclosures only) (n=281).

174 We examined four plant community properties: (i) CWMGS; (ii) CWMC-strategy; (iii) total above-
175 ground community biomass (estimated from total dry sample weight); and (iv) phylogenetic
176 diversity. CWMC-strategy of each plot was calculated from each taxon's C-strategy (estimated with

177 PGLS, as described above). Phylogenetic diversity (PD) corresponds to Faith's PD [41] and is the sum
178 of the phylogenetic branch lengths of each community, estimated with the *picante* package [42].
179 We used directional separation (*d-sep*) path analysis methods [43] to assess fourteen hypotheses
180 about the effects of experimental treatment (herbivore exclusion, N and P plus K fertilizer) on the
181 plant community properties (Fig. SI.3, see also Table SI.3 for a more complete description of *d-sep*
182 methods). The conditional independencies were fitted with LME models [48,49]. Random effects
183 were specified as plot/fencing/herbicide in the rabbit exclusion analyses, and as plot/herbicide in
184 the insect-mollusc exclusion analyses. Herbicide treatment was also accounted for as a fixed effect.
185 The experiment was fitted as herbivore treatment x N x P + K + herbicide; where herbivore
186 treatment = \pm rabbits, or \pm insects x \pm molluscs. Continuous independent variables were
187 standardized by two standard deviations [44] with the 'rescale' function from the *arm* package [45].
188 Because p-values are the main determinant in assessing acyclic diagrams, each equation was also
189 systematically fitted with ten unequal variance structures using the 'varIdent' function from the
190 *nlme* package [37] to account for heteroscedasticity in the residuals (Table SI. 3). We retained the p-
191 value from the regression resulting in the lowest second-order AIC (AICc), which corrects for small
192 sample sizes, implemented in *MuMIn* [46], but only if an analysis of variance showed that a
193 regression fitted with the variance structure was significantly better ($p < 0.05$) than without.
194 Interactions among community properties, and between the experiment and community properties,
195 were not allowed. We present one directed acyclic diagram for each dataset, based on the CIC-
196 statistic and a preference for a more parsimonious diagram (Table SI.4), but this does not exclude
197 alternative hypotheses that passed the goodness of fit tests. The conditional R^2 values for LME
198 equations were estimated with *MuMIn*, and are measures of how much variance is explained by
199 both fixed and random effects [46].
200

201 **3. RESULTS**

202 Of the 56 species collected during the field sampling, 12 species generated at least 1% mean
 203 biomass in at least two herbivore treatments. Seven were grasses (Poaceae): *Agrostis capillaris*,
 204 *Arrhenatherum elatius*, *Dactylis glomerata*, *Festuca rubra*, *Holcus lanatus*, *Holcus mollis*, and *Phleum*
 205 *pratense* subsp. *bertolonii*. The remaining five were *Cirsium arvense*, *Jacobaea vulgaris* (Asteraceae),
 206 *Plantago lanceolata*, *Veronica chamaedrys*, (Plantaginaceae), and *Rumex acetosa* (Polygonaceae).
 207 Overall, for the 56 species, GS ranged from 0.28 pg/1C in *Juncus effusus* to 11.06 pg/1C in
 208 *Ranunculus repens* (Table SI.2).

209 **(a) Species occurrence and species biomass are a function of interactions between GS, herbivory,**
 210 **and nutrients**

211 We investigated the impact of herbivores, nutrient treatment, and species GS on the occurrence and
 212 biomass of the 12 species listed above. Herbivore exclusion and GS significantly impacted species
 213 occurrence. With insect exclusion, the probability of a species occurrence (inv.logit(B)) was 0.813
 214 (95% credible intervals (CI) = 0.397, 0.974, pMCMC=0.0425) increased by 0.472, or 47.2% in
 215 comparison to control plots with all herbivores present (Table SI.5A). Two significant interactions
 216 between GS and herbivores were apparent: (i) GS and rabbit exclusion - species with larger GS were
 217 more likely to be present on plots without rabbits. Each pg increase in GS was associated with an
 218 8.0% increased likelihood of a species occurrence (inv.logit(B)=0.573, CI=0.530, 0.613,
 219 pMCMC<0.0002), in comparison to control plots (inv.logit(B)=0.49, Table SI.5A); (ii) GS and mollusc
 220 exclusion - removing molluscs increased the probability of a species with a smaller GS being present.
 221 Each pg decrease in GS was associated with a 6.6% increase in species occurrence
 222 (inv.logit(B)=0.427, CI=0.387, 0.465, pMCMC=0.0020, Table SI. 5A).

223 Species above-ground biomass was influenced by interactions between GS, herbivory and N (Table
 224 SI.5B). In control plots, the estimated effect of GS was a c. 62% increase in species biomass per pg
 225 increase in GS (exp(B)=1.618, CI=1.19, 2.36, pMCMC=0.0057). The above-ground biomass of the 12
 226 species was further impacted by three interactions between GS and experimental treatments. On

227 plots with all herbivores present, the addition of N increased species biomass by more than two-fold
 228 ($\exp(B)=2.33$, $CI=1.186, 2.358$, $pMCMC=0.0105$). However, this was dependent on GS, with most of
 229 the biomass increase coming from species with small GS - on +N plots, biomass decreased by c. 23%
 230 per pg increase in GS ($\exp(B)=0.765$, $CI=0.608, 0.974$, $pMCMC=0.0243$, Table SI.5B). The second
 231 interaction was an increase in biomass of species with larger GS on plots without rabbits - species
 232 biomass increased by 21% per pg increase in GS in comparison to control plots ($\exp(B)=1.2099$,
 233 $CI=1.047, 1.396$, $pMCMC=0.0117$). This contrasted with the third interaction observed as the
 234 amount of biomass produced by species with larger GS on plots decreased with mollusc exclusion - a
 235 biomass decrease of c. 14% per pg increase in GS was observed when molluscs were excluded
 236 compared to control plots ($\exp(B)=0.8604$, $CI=0.736, 0.987$, $pMCMC=0.0466$). Insect removal did
 237 not significantly affect growth of the 12 species.

238

239 **(b) Herbivory and nitrogen influence community-weighted mean (CWM) GS**

240 For community level analyses, CWMGS were estimated for all 556 plots using biomass values for the
 241 56 species; these ranged from 1.36 pg/1C (on a plot with +N and -rabbits and -molluscs (i.e. insects
 242 only) to 8.61 pg/1C (on a plot with +N and -rabbits). For control plots (all herbivores present, no
 243 nutrient input) the CWMGS was estimated at 5.76 pg/1C (95% confidence interval (CI) 5.02, 6.50)
 244 (Table SI.7).

245 The most parsimonious linear mixed effects (LME) model (obtained with stepwise reduction, Table
 246 SI.6) revealed that: (i) on unfertilized plots, the biomass from plants with larger GS increased without
 247 rabbits - CWMGS of plots increased by 1.10 pg/1C when rabbits were excluded compared to control
 248 plots (Fig. 1, $CI=0.26, 1.95$, $p=0.0538$, Table SI.7). This effect became significant with the addition of
 249 N, with CWMGS of plots increasing by 0.33 pg/1C with rabbit exclusion, compared to control plots
 250 ($CI=0.46, 1.73$, $p=0.0008$). (ii) In contrast, the biomass from plants with smaller GS increased on
 251 plots with both mollusc and insect exclusion and N input. The CWMGS of plots decreased by
 252 2.73 pg/1C on +N plots with rabbit grazing only ($B=1.433$, $CI=0.52, 2.35$, $p=0.0024$). (iii) Nitrogen

253 fertilizer decreased CWMGS of plots with all herbivores present by 1.87 pg/1C (CI=-2.42, -1.32,
 254 $p < 0.0001$), and (iv) P fertilizer decreased CWMGS by 0.31 pg/1C (CI=-0.54, -0.08, $p = 0.0078$). (v)
 255 CWM GS of plots with only insect grazing (rabbit and mollusc exclusion) and N fertilization decreased
 256 by 1.54 pg/1C compared to control plots (Fig. 1, CI=-2.10, -0.28, $p = 0.0114$, Tables SI.7).

257 We also estimated CWMGS (i) without including a phylogenetic correlation structure; (ii) with GS
 258 untransformed; (iii) with the lambda parameter optimised; and showed that these were similar to
 259 CWM GS used above and that the key results presented here remain unchanged (Fig. SI.4 and Table
 260 SI.9).

261 **(c) Contrasting and similar effects of rabbit and invertebrate herbivory on plant community**
 262 **properties**

263 Using confirmatory path analysis, we investigated how the following four plant community
 264 properties were influenced by each other and by herbivory and nutrient treatments: (i) CWMGS, (ii)
 265 CWM C-strategy (i.e. species competitiveness), (iii) total above-ground community biomass, and (iv)
 266 phylogenetic diversity (Fig. 2a and b, Fig. SI.5 and Tables SI.10 (rabbit exclusion) and SI.11 (insect and
 267 mollusc exclusion) for complete regression tables of Fig. 2a and b respectively).

268 In testing the impact of rabbits, the four community properties were influenced by various factors.
 269 The interaction between N input and rabbit exclusion increased CWMGS by 0.99 pg (CI=0.44, 2.24,
 270 $p = 0.0047$) (Fig. 2a, Table SI.10). CWM C-strategy increased with N input ($B = 0.089$, CI=0.04, 0.15,
 271 $p = 0.0016$), and with CWM GS ($B = 0.025$, CI=0.02, 0.04, $p < 0.0001$), particularly on plots without
 272 rabbits (Fig. SI.6a, b). Total biomass increased with rabbit exclusion and N input ($B = 19.97$, CI=10.85,
 273 29.08, $p = 0.0001$). The single largest standardized effect is the reduction in phylogenetic diversity
 274 with rabbit exclusion ($B = -15.48$, CI=-22.16, -9.54, $p = 0.0003$).

275 In contrast, removal of molluscs and insects did not reduce phylogenetic diversity. Instead, N input,
 276 and to a lesser extent, K input, were the main drivers of this ($B = -12.11$, CI=-18.54, -5.68, $p = 0.0004$;
 277 and $B = -3.58$, CI=-6.02, -1.15, $p = 0.0052$ respectively) (Fig. 2b, Table SI.11).

278 The main driver influencing occurrence of species with a competitive life strategy (i.e. CWM C-
 279 strategy) was a four-way interaction between insect and mollusc exclusion and +N and +P input,
 280 increasing CWM C-strategy by 0.23 units (CI=-0.46, -0.11, $p=0.002$). Similar to \pm rabbit plots, CWM C-
 281 strategy also increased with CWM GS (B=0.025, CI=0.02, 0.03, $p<0.0002$, Fig. SI.6c-f). Total biomass
 282 increased significantly with both +N and mollusc (but not insect) removal by a mean of 28.05 g (in
 283 dry weight) ($p=0.0073$). In addition, increased biomass was associated with increased CWM C-
 284 strategy (B=65.65, CI=43.63, 85.97, $p<0.0001$). Finally, in contrast to rabbit exclusion, a four-way
 285 interaction between +N, +P, insect and mollusc exclusion led to a decrease of 2.39 pg in CWM GS
 286 ($p=0.0464$). Total biomass and phylogenetic diversity were not significantly associated with each
 287 other in both path analyses.

288 **(d) Changes in species composition with herbivore exclusion and N fertilizer**

289 Species that increased with rabbit exclusion, include the larger genomed *Arrhenatherum. elatius*
 290 (8.61 pg/1C), and to a lesser extent *H. mollis* (4.1 pg/1C), and *D. glomerata* (4.4 pg/1C). The response
 291 of *F. rubra* (7.31 pg/1C) was dependent on N, decreasing on -N plots, and increasing on +N plots.
 292 The removal of molluscs led to an increase in the smaller genomed *H. lanatus* (1.7 pg/1C), followed
 293 by *P. pratense* subsp. *bertolonii* (1.99 pg/1C) and *H. mollis* (4.1 pg/1C), at the expense of the larger
 294 genomed *F. rubra* and *A. elatius*. In contrast, species that increased most with just insect removal
 295 had a range of GS and included *A. elatius* (on -N plots), *H. mollis* (on +N plots), *J. vulgaris*
 296 (2.25 pg/1C), *C. arvense* (1.42 pg/1C), and *H. lanatus*; whilst *A. capillaris* (3.6 pg/1C) and *F. rubra*
 297 decreased.

298 Within fenced plots, species that increased with removal of molluscs and insects include *H. mollis*,
 299 *H. lanatus*, *D. glomerata*, *Achillea millefolium* (7.98 pg/1C), *P. pratense* subsp. *bertolonii*, and
 300 *V. chamaedrys* (2.16 pg/1C). *Holcus mollis* increased consistently on +N plots, whereas *F. rubra*,
 301 *A. capillaris*, *C. arvense* (1.42 pg/1C), decreased on -N plots (see also Fig. SI.7 for biplots representing
 302 species abundances for the experimental treatments, and Table SI.12 for percentage change in
 303 species biomass).

304

305 **4. DISCUSSION**

306 The experiment at Nash's Field involves the input of nutrients and the exclusion of grazing by
307 molluscs, insects and rabbits, allowing us to detect significant interactions between GS, herbivory
308 and N fertilization. We show that these interactions impact plant community structure, plant
309 biomass production and species diversity. Previous work showed that plant community structure
310 was influenced by interactions between plant GS and nutrients directly, and that species with larger
311 GS only contributed significant biomass to plant communities when nutrients were not limiting [40,
312 47]. We did not observe this effect here, perhaps because: (i) The experiment is still young (data
313 were collected only 6 to 8 years after the start of the experiment), compared with the Park Grass
314 (>150 years) and Rengen Grassland (>70 years) Experiments, and thus plant communities may still
315 be adapting to the experimental treatments and are in a transient state [48]; (ii) Since the 1950s
316 intensive rabbit grazing is known to have been prevalent in the area used to establish Nash's Field
317 Experiment in 1992 and this would have influenced the species that colonised the plots towards
318 those that are grazing-tolerant and/or grazing-resistant [49, 50].

319 **(a) Impact of rabbit herbivory**

320 We observed that the plant species which generated most biomass on rabbit-grazed plots had
321 smaller GS than those on ungrazed plots, especially when N fertilizer was added. There could be
322 because:

323 (i) Rabbits preferentially eat plant taxa with large GS. Plant species which increase in biomass when
324 herbivores are excluded are generally species that are preferentially grazed by herbivores [51].
325 Rabbits are known to favour high-nutrient plants [52]. N input qualitatively alters plant nutrient
326 content [53] and plants growing under higher N may be more attractive to consumers and
327 increase herbivore numbers [10, 54]. Potentially rabbits may prefer species with larger GS
328 because at the cellular level, they have higher N and P content (both DNA and RNA are rich in
329 these macronutrients) compared to species with smaller GS [53, 55]. In addition, plants with

330 larger cells may be more palatable as cell size tends to increase with GS, and larger and fewer
331 cells per leaf would decrease the amount of cell wall, potentially rendering the plant more
332 succulent.

333 (ii) Plant taxa with smaller GS are better able to recover from the rabbit grazing pressure. Rapid
334 growth might be best achieved in plants with small GS which have faster cell cycle times, shorter
335 durations of DNA synthesis, and hence shorter minimum generation times [reviewed in 14], and
336 may be less constrained by nutrient availability for growth and repair [56]. Indeed, tolerance to
337 grazing may be a key survival trait on fertilized plots, where rabbit abundance can be four times
338 greater than on unfertilized plots [57]. Rabbits generate sustained stresses that impact grassland
339 composition and dynamics, such as the selection of plants with rapid growth rates [58]. Rapid
340 growth rate is fundamental to tolerance of herbivory, allowing for regrowth of damaged tissue.
341 Such regrowth is achieved via rapid resource allocation, increased photosynthetic rates, and
342 increased nutrient uptake [59]. Trade-offs may therefore exist between the biochemical costs
343 associated with building and maintaining a large genome and tolerance to herbivore damage.

344 (iii) Plant taxa with smaller GS may be less constrained to allocate resources to the production of
345 secondary metabolites for defence. Investment in defence has been shown to lead to decreased
346 rates of photosynthesis, and the diversion of elemental resources such as C and N towards the
347 production of defence compounds at the expense of growth [60]. Defence is also costly in both
348 water and nutrient resources [61], leads to the remobilization of elements to roots [62], and
349 would compete with the N and P costs of building and maintaining a large genome, especially in
350 nutrient deficient environments.

351 In addition to the direct effects of preferential grazing, rabbits increase habitat and resource
352 heterogeneity by trampling, burrowing, decreasing ground cover, and deposition of droppings.
353 These effects may enable functionally more diverse species to colonize the community [7]. We
354 observed that the presence of rabbits leads to a significant increase in phylogenetic diversity, which
355 is also associated with a decrease in plant species with a competitive life strategy and a decrease in

356 total biomass (Fig. 2a). Rabbit grazing on the fast-growing, dominant plants keeps these in check,
 357 allowing the establishment of subordinate species. This effect has also been reported across various
 358 types of grasslands with mammalian herbivores, but depends on plant productivity, generally being
 359 positive in grasslands with high productivity, but decreasing plant diversity when productivity is low
 360 [63, 64]. Experiments are now needed determine whether plants with larger GSs, are indeed of
 361 higher nutrient quality, altered N and P contents and different recovery periods following damage by
 362 herbivory.

363 **(b) Impact of invertebrate herbivory**

364 In contrast to rabbit grazing, mollusc and insect grazing leads to communities composed of plants
 365 with larger GS. In the absence of fertilizer, mollusc and insect herbivory appear to mitigate the
 366 effects of rabbit grazing, as plant communities on plots with no herbivores have similar CWMGS as
 367 the control plots where all herbivores are present (Fig. 1). Numerous studies have shown that
 368 molluscs have species-specific food preferences, which relate to different food qualities and a plant's
 369 palatability, the latter being influenced by, for example, the presence of plant anti-herbivory
 370 chemicals, silica content, and a plant's pubescence which can deter feeding [65]. Molluscs are also
 371 reported to have a preference for seedlings, however in a temperate grassland seedling recruitment
 372 is low, where plant regeneration is mostly via vegetative regrowth [23]. Potentially the invertebrates
 373 at Nash's Field are specialists on plants or plant organs that are not favoured by rabbits. Nutrient
 374 acquisition strategies vary between and within these herbivore guilds, and the scale and impact of
 375 different insect herbivores (predominantly grasshoppers) on grassland plant community biomass has
 376 been shown to be linked closely with, for example, mandibular trait diversity [66]. It is possible that
 377 insects as a guild feed on a larger range of species and tissues than molluscs as a guild, making the
 378 effects of insects on community plant GS more difficult to detect.

379 Whilst insect and mollusc herbivory decreased total community biomass, the effects of insects and
 380 molluscs on community composition were negligible when fertilizers were added, perhaps because
 381 fertilizers can obscure the effects of nutrient recycling by grazers, although interactions are complex

382 [1]. The application of N and K led to a reduction in phylogenetic diversity, for which the presence of
383 insects and molluscs did not compensate. This is opposite to the significant effect of rabbits; their
384 larger size and activities (as noted above) may result in more pronounced effects on plant diversity
385 [63]. Previous findings on the unlimed plots at Nash's Field, included a decrease in species diversity
386 with insect removal, and, to some extent, mollusc removal [23]. The effects of insects and molluscs
387 may take longer to occur on limed plots, as these do not have the additional stress of low pH
388 conditions.

389 **(c) Genome size and ecological models**

390 Investigations of plant traits have uncovered global trends that help to predict plant responses to
391 abiotic and biotic factors [67-69]. Here we propose that GS should also be considered as an
392 important plant functional trait which influences plant community structure through bottom-up
393 (nutrients) [70] and top-down (herbivory) interactions with GS. We show here that larger CWM GS is
394 linked with higher CWMC-strategy, which is, in turn, linked with increased community biomass.
395 Higher species competitiveness is also associated with decreased species diversity in the absence of
396 rabbit grazing. Through plant consumption, herbivores also influence plant community composition,
397 including the abundance of plants with a competitive life strategy, which can impact nutrient cycling
398 by altering litter quality [1]. The effect of GS on plant tolerance to herbivory, and possible influence
399 on herbivore preference may also have wider ecological impact, influencing processes such as N
400 cycling, and hence add a new dimension to improve the performance of ecological models.

401

402 **Data accessibility**

403 All data will be made available in Dryad if the manuscript is accepted.

404 **Competing interests**

405 We have no competing interests.

406 **Authors' contributions**

407 MSG and DK carried out field-work and GS estimation. MSG carried out the data analyses. MJC
408 established the experimental plots, assisted with species identification and provided biomass data.
409 RAN provided statistical advice. MSG, IJL, and ARL devised the fieldwork, analyses, and wrote the
410 manuscript. All authors read, revised, and approved the final manuscript.

411 **Acknowledgements.** We would like to thank the anonymous referees for their useful and helpful
412 comments. A list of the C-S-R strategies for over 1000 European species was made available by JG
413 Hodgson (UCPE Sheffield) from [http://people.exeter.ac.uk/rh203/plant-scientist-recent-science-](http://people.exeter.ac.uk/rh203/plant-scientist-recent-science-functional-types-allocating-csr.html)
414 [functional-types-allocating-csr.html](http://people.exeter.ac.uk/rh203/plant-scientist-recent-science-functional-types-allocating-csr.html)

415 **Funding**

416 This study was supported by funding from the Research Council of Norway (196468/v40) and the
417 Natural Environment Research Council (NERC) (NE/J012106/1).

418

419 **References**

- 420 1. Sitters J, Bakker ES, Veldhuis MP, Veen GF, Olde Venterink H, et al. 2017 The stoichiometry of
 421 nutrient release by terrestrial herbivores and its ecosystem consequences. *Frontiers in Earth*
 422 *Science* **5**.
- 423 2. Hunter MD, Price PW. 1992 Playing chutes and ladders: Heterogeneity and the relative roles
 424 of bottom-up and top-down forces in natural communities. *Ecology* **73**, 724-732.
- 425 3. Harpole WS, Ngai JT, Cleland EE, Seabloom EW, Borer ET, et al. 2011 Nutrient co-limitation of
 426 primary producer communities. *Ecol Lett* **14**, 852-862.
- 427 4. Humbert J-Y, Dwyer JM, Andrey A, Arlettaz R. 2016 Impacts of nitrogen addition on plant
 428 biodiversity in mountain grasslands depend on dose, application duration and climate: a
 429 systematic review. *Glob Change Biol* **22**, 110-120.
- 430 5. Cronin JP, Tonsor SJ, Carson WP. 2010 A simultaneous test of trophic interaction models:
 431 which vegetation characteristic explains herbivore control over plant community mass? *Ecol*
 432 *Lett* **13**, 202-212.
- 433 6. Laliberté E, Tylianakis JM. 2012 Cascading effects of long-term land-use changes on plant traits
 434 and ecosystem functioning. *Ecology* **93**, 145-155.
- 435 7. Olff H, Ritchie ME. 1998 Effects of herbivores on grassland plant diversity. *Trends Ecol Evol* **13**,
 436 261-265.
- 437 8. Borer ET, Seabloom EW, Gruner DS, Harpole WS, Hillebrand H, et al. 2014 Herbivores and
 438 nutrients control grassland plant diversity via light limitation. *Nature* **508**, 517-520.
- 439 9. Throop HL, Lerdau MT. 2004 Effects of nitrogen deposition on insect herbivory: Implications
 440 for community and ecosystem processes. *Ecosystems* **7**, 109-133.
- 441 10. Augustine DJ, McNaughton SJ, Frank DA. 2003 Feedbacks between soil nutrients and large
 442 herbivores in a managed savanna ecosystem. *Ecological Applications* **13**, 1325-1337.
- 443 11. Eskelinen A, Harrison S, Tuomi M. 2012 Plant traits mediate consumer and nutrient control on
 444 plant community productivity and diversity. *Ecology* **93**, 2705-2718.
- 445 12. Diaz S, Lavorel S, McIntyre S, Falczuk V, Casanoves F, et al. 2007 Plant trait responses to
 446 grazing – a global synthesis. *Glob Change Biol* **13**, 313-341.
- 447 13. Pellicer J, Hidalgo O, Dodsworth S, Leitch IJ. 2018 Genome size diversity and its impact on the
 448 evolution of land plants. *Genes* **9**, 88.
- 449 14. Greilhuber J, Leitch IJ. 2013 Genome size and the phenotype. In *Plant genome diversity, vol 2,*
 450 *Physical structure, behaviour and evolution of plant genomes* (eds. Leitch IJ, Greilhuber J,
 451 Doležel J, Wendel JF), pp. 323-344. Wien, Springer-Verlag.
- 452 15. Thompson JN, Nuismer SL, Merg K. 2004 Plant polyploidy and the evolutionary ecology of
 453 plant/animal interactions. *Biol J Linn Soc* **82**, 511-519.
- 454 16. Munzbergova Z. 2006 Ploidy level interacts with population size and habitat conditions to
 455 determine the degree of herbivory damage in plant populations. *Oikos* **115**, 443-452.
- 456 17. Münzbergová Z, Skuhrovec J, Maršík P. 2015 Large differences in the composition of herbivore
 457 communities and seed damage in diploid and autotetraploid plant species. *Biol J Linn Soc* **115**,
 458 270-287.
- 459 18. Richardson ML, Hanks LM. 2011 Differences in spatial distribution, morphology, and
 460 communities of herbivorous insects among three cytotypes of *Solidago altissima* (Asteraceae).
 461 *Am J Bot* **98**, 1595-1601.
- 462 19. Meyerson LA, Cronin JT, Bhattarai GP, Brix H, Lambertini C, et al. 2016 Do ploidy level and
 463 nuclear genome size and latitude of origin modify the expression of *Phragmites australis* traits
 464 and interactions with herbivores? *Biological Invasions* **18**, 2531-2549.
- 465 20. Balocchi OA, López IF. 2009 Herbage production, nutritive value and grazing preference of
 466 diploid and tetraploid perennial ryegrass cultivars (*Lolium perenne* L.). *Chilean Journal of*
 467 *Agricultural Research* **69**, 331-339.
- 468 21. Bennett MD. 1972 Nuclear DNA content and minimum generation time in herbaceous plants.
 469 *Proc R Soc Lond Ser B* **181**, 109-135.

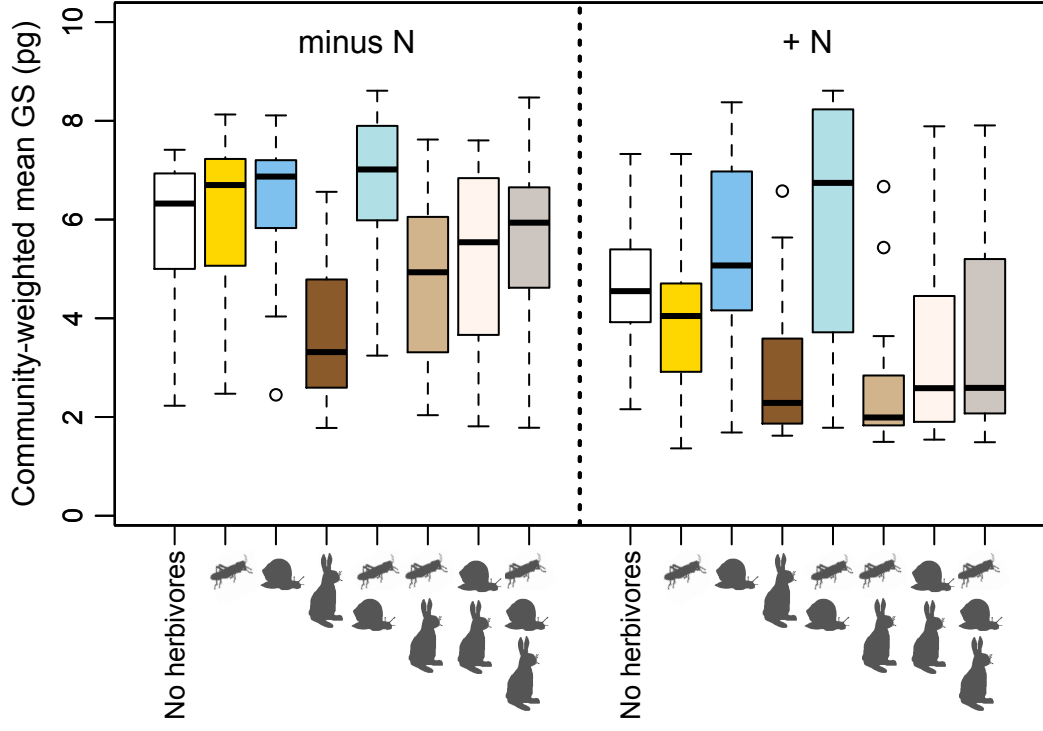
- 470 22. Kang M, Wang J, Huang H. 2015 Nitrogen limitation as a driver of genome size evolution in a
471 group of karst plants. *Scientific Reports* **5**, 11636.
- 472 23. Allan E, Crawley MJ. 2011 Contrasting effects of insect and molluscan herbivores on plant
473 diversity in a long-term field experiment. *Ecol Lett* **14**, 1246-1253.
- 474 24. Zhao XQ, Shen RF. 2018 Aluminum–nitrogen interactions in the soil–plant system. *Front Plant*
475 *Sci* **9**, 807.
- 476 25. Pellicer J, Leitch IJ. 2014 The application of flow cytometry for estimating genome size and
477 ploidy level in plants. In *Molecular Plant Taxonomy* (ed. Besse P), pp. 279-307, Humana Press.
- 478 26. Bennett MD, Leitch IJ. 2012 Plant DNA C-values database (release 6.0, Dec. 2012).
479 <http://data.kew.org/cvalues/>.
- 480 27. Durka W, Michalski SG. 2012 Daphne: a dated phylogeny of a large European flora for
481 phylogenetically informed ecological analyses. *Ecology* **93**, 2297-2297.
- 482 28. Paradis E, Claude J, Strimmer K. 2004 APE: Analyses of Phylogenetics and Evolution in R
483 language. *Bioinformatics* **20**, 289-290.
- 484 29. Orme D, Freckleton RP, Thomas G, Petzoldt T, Fritz S, et al. 2018 caper: Comparative Analyses
485 of Phylogenetics and Evolution in R. R package version 1.0.1. [https://CRAN.R-](https://CRAN.R-project.org/package=caper)
486 [project.org/package=caper](https://CRAN.R-project.org/package=caper).
- 487 30. Revell LJ. 2012 phytools: an R package for phylogenetic comparative biology (and other
488 things). *Methods Ecol Evo* **3**, 217-223.
- 489 31. Violle C, Navas M-L, Vile D, Kazakou E, Fortunel C, et al. 2007 Let the concept of trait be
490 functional! *Oikos* **116**, 882-892.
- 491 32. R Development Core Team. 2017 R: A language and environment for statistical computing.
492 (doi: ISBN 3-900051-07-0).
- 493 33. Grime JP. 1977 Evidence for the existence of three primary strategies in plants and its
494 relevance to ecological and evolutionary theory. *Am Nat* **111**, 1169-1194.
- 495 34. Hodgson JG, Wilson PJ, Hunt R, Grime JP, Thompson K. 1999 Allocating CSR plant functional
496 types: a soft approach to a hard problem. *Oikos* **85**, 282–294.
- 497 35. Hadfield JD. 2010 MCMC methods for multi-response generalized linear mixed models: the
498 MCMCglmm R package. *J Stat Softw* **33**, 1-22.
- 499 36. Plummer M, Best N, Cowles K, Vines K. 2006 CODA: convergence diagnosis and output analysis
500 for MCMC. *R News* **6**, 7–11.
- 501 37. Pinheiro J, Bates D, DebRoy S, Sarkar D, R Core Team. 2012 nlme: Linear and Nonlinear Mixed
502 Effects Models. R package version 3.1-103. <http://cran.r-project.org/package=nlme>.
- 503 38. Bates D, Mächler M, Bolker B, Walker S. 2015 Fitting Linear Mixed-Effects models using lme4. *J*
504 *Stat Softw* **67**, 48.
- 505 39. Kuznetsova A, Brockhoff PB, Christensen RHB. 2017 lmerTest Package: Tests in Linear Mixed
506 Effects Models. *J Stat Softw* **82**, 26.
- 507 40. Guignard MS, Nichols RA, Knell RJ, Macdonald A, Romila C-A, et al. 2016 Genome size and
508 ploidy influence angiosperm species' biomass under nitrogen and phosphorus limitation. *New*
509 *Phytol* **210**, 1195-1206.
- 510 41. Faith DP. 1992 Conservation evaluation and phylogenetic diversity. *Biol Conserv* **61**, 1-10.
- 511 42. Kembel SW, Cowan PD, Helmus MR, Cornwell WK, Morlon H, et al. 2010 Picante: R tools for
512 integrating phylogenies and ecology. *Bioinformatics* **26**, 1463-1464.
- 513 43. Shipley B. 2009 Confirmatory path analysis in a generalized multilevel context. *Ecology* **90**,
514 363-368.
- 515 44. Gelman A. 2008 Scaling regression inputs by dividing by two standard deviations. *Statistics in*
516 *Medicine* **27**, 2865-2873.
- 517 45. Gelman A, Su Y-S. 2018 arm: Data Analysis using Regression and Multilevel/Hierarchical
518 models. R package version 1.10-1. <https://CRAN.R-project.org/package=arm>.
- 519 46. Bartoń K. 2018 MuMIn: Multi-Model Inference. R Package version 1.42.1. [https://CRAN.R-](https://CRAN.R-project.org/package=MuMIn)
520 [project.org/package=MuMIn](https://CRAN.R-project.org/package=MuMIn)
- 521 47. Šmarda P, Hejčman M, Březinová A, Horová L, Steigerová H, et al. 2013 Effect of phosphorus
522 availability on the selection of species with different ploidy levels and genome sizes in a long-
523 term grassland fertilization experiment. *New Phytol* **200**, 911-921.

- 524 48. Lehman CL, Tilman D. 2000 Biodiversity, stability, and productivity in competitive
525 communities. *Am Nat* **156**, 534-552.
- 526 49. Rasmann S, De Vos M, Casteel CL, Tian D, Halitschke R, et al. 2012 Herbivory in the previous
527 generation primes plants for enhanced insect resistance. *Plant Physiol* **158**, 854-863.
- 528 50. Howe HF, Brown JS. 2001 The ghost of granivory past. *Ecol Lett* **4**, 371-378.
- 529 51. Kempel A, Razanajatovo M, Stein C, Unsicker SB, Auge H, et al. 2015 Herbivore preference
530 drives plant community composition. *Ecology* **96**, 2923-2934.
- 531 52. Miller GR. 1968 Evidence for selective feeding on fertilized plots by red grouse, hares, and
532 rabbits. *The Journal of Wildlife Management* **32**, 849-853.
- 533 53. Güsewell S. 2004 N : P ratios in terrestrial plants: variation and functional significance. *New*
534 *Phytol* **164**, 243-266.
- 535 54. Ball JP, Danell K, Sunesson P. 2000 Response of a herbivore community to increased food
536 quality and quantity: an experiment with nitrogen fertilizer in a boreal forest. *Journal of*
537 *Applied Ecology* **37**, 247-255.
- 538 55. Hessen DO, Jeyasingh PD, Neiman M, Weider LJ. 2010 Genome streamlining and the
539 elemental costs of growth. *Trends Ecol Evol* **25**, 75-80.
- 540 56. Elser JJ, Dobberfuhl DR, MacKay NA, Schampel JH. 1996 Organism size, life history, and N:P
541 stoichiometry toward a unified view of cellular and ecosystem processes. *Bioscience* **46**, 674-
542 684.
- 543 57. Bakker ES, Reiffers RC, Olff H, Gleichman JM. 2005 Experimental manipulation of predation
544 risk and food quality: effect on grazing behaviour in a central-place foraging herbivore.
545 *Oecologia* **146**, 157-167.
- 546 58. Turley NE, Odell WC, Schaefer H, Everwand G, Crawley MJ, et al. 2013 Contemporary
547 evolution of plant growth rate following experimental removal of herbivores. *Am Nat* **181**,
548 S21-S34.
- 549 59. Strauss SY, Agrawal AA. 1999 The ecology and evolution of plant tolerance to herbivory.
550 *Trends Ecol Evol* **14**, 179-185.
- 551 60. Züst T, Agrawal AA. 2017 Trade-offs between plant growth and defense against insect
552 herbivory: an emerging mechanistic synthesis. *Annual Review of Plant Biology* **68**, 513-534.
- 553 61. Kirk H, Vrieling K, Van Der Meijden E, Klinkhamer PG. 2010 Species by environment
554 interactions affect pyrrolizidine alkaloid expression in *Senecio jacobaea*, *Senecio aquaticus*,
555 and their hybrids. *Journal of Chemical Ecology* **36**, 378-387.
- 556 62. Liu M, Gong J, Li Y, Li X, Yang B, et al. 2018 Growth–defense trade-off regulated by hormones
557 in grass plants growing under different grazing intensities. *Physiologia Plantarum* **0**,
558 doi:10.1111/ppl.12802.
- 559 63. Bakker ES, Ritchie ME, Olff H, Milchunas DG, Knops JMH. 2006 Herbivore impact on grassland
560 plant diversity depends on habitat productivity and herbivore size. *Ecol Lett* **9**, 780-788.
- 561 64. Mortensen B, Danielson B, Harpole WS, Alberti J, Arnillas CA, et al. 2018 Herbivores safeguard
562 plant diversity by reducing variability in dominance. *Journal of Ecology* **106**, 101-112.
- 563 65. Motheral SM, Orrock JL. 2009 Gastropod herbivore preference for seedlings of two native and
564 two exotic grass species. *The American Midland Naturalist* **163**, 106-114.
- 565 66. Deraison H, Badenhausser I, Loeuille N, Scherber C, Gross N. 2015 Functional trait diversity
566 across trophic levels determines herbivore impact on plant community biomass. *Ecol Lett* **18**,
567 1346-1355.
- 568 67. Reich PB, Oleksyn J. 2004 Global patterns of plant leaf N and P in relation to temperature and
569 latitude. *Proc Natl Acad Sci USA* **101**, 11001-11006.
- 570 68. Gillison AN. 2013 Plant functional types and traits at the community, ecosystem and world
571 level. In *Vegetation Ecology* (eds. van der Maarel E, Franklin J), pp. 347-386. Chichester, UK,
572 John Wiley & Sons Ltd.
- 573 69. Díaz S, Kattge J, Cornelissen JHC, Wright IJ, Lavorel S, et al. 2015 The global spectrum of plant
574 form and function. *Nature* **529**, 167–171
- 575 70. Guignard MS, Leitch AR, Acquisti C, Eizaguirre C, Elser JJ, et al. 2017 Impacts of nitrogen and
576 phosphorus: from genomes to natural ecosystems and agriculture. *Frontiers in Ecology and*
577 *Evolution* **5**, 70.

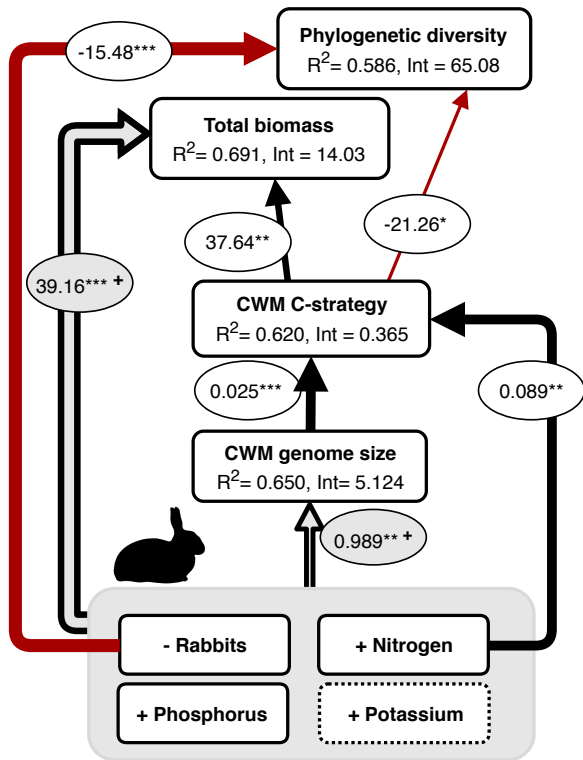
579 **FIGURE LEGENDS**

580 **Fig. 1:** Community-weighted mean genome size (GS; 1C-value, pg) of each plot (n = 556) under eight
 581 herbivore exclusion treatments and \pm nitrogen (N) input. Herbivore treatments, in order as shown
 582 below are: 1) control (all herbivores present); 2) – insects; 3) – molluscs; 4) – rabbits; 5) rabbits only
 583 (– insects, – molluscs); 6) molluscs only (– insects, – rabbits); 7) insects only (– molluscs, – rabbits); 8)
 584 no herbivores. Boxes show median of community-weighted mean GS, first and third quartiles, and
 585 minimum and maximum values (summary statistics in Table SI.8).

586 **Fig. 2:** Path analysis examining effects of (A) rabbit exclusion and (B) mollusc and insect exclusion
 587 together with nutrient input (N, P, K) on four plant community properties: (i) community-weighted
 588 mean (CWM) genome size (GS), (ii) CWM competitive (C)-strategy, (iii) total biomass, and (iv)
 589 phylogenetic diversity. Values in ovals show the effect size of one variable, or interaction, on the
 590 other. The conditional R^2 and the intercept are given for each community property. Arrow widths
 591 are proportional to standardised coefficients. P-values: ***<0.0001, **<0.001, *<0.05 (see Tables
 592 SI.10 and SI.11 for details in A and B respectively). (A) For rabbits, a three-way interaction (\pm rabbits
 593 x N x P) did not contribute significantly towards any community property, thus only two-way
 594 interactions were included between \pm rabbits, N and P. (B) For molluscs and insects, CWM GS could
 595 not be unlinked from total biomass in a conditional independency test, however its partial
 596 regression coefficient is below the alpha level (0.050 at $p=0.0709$).



A



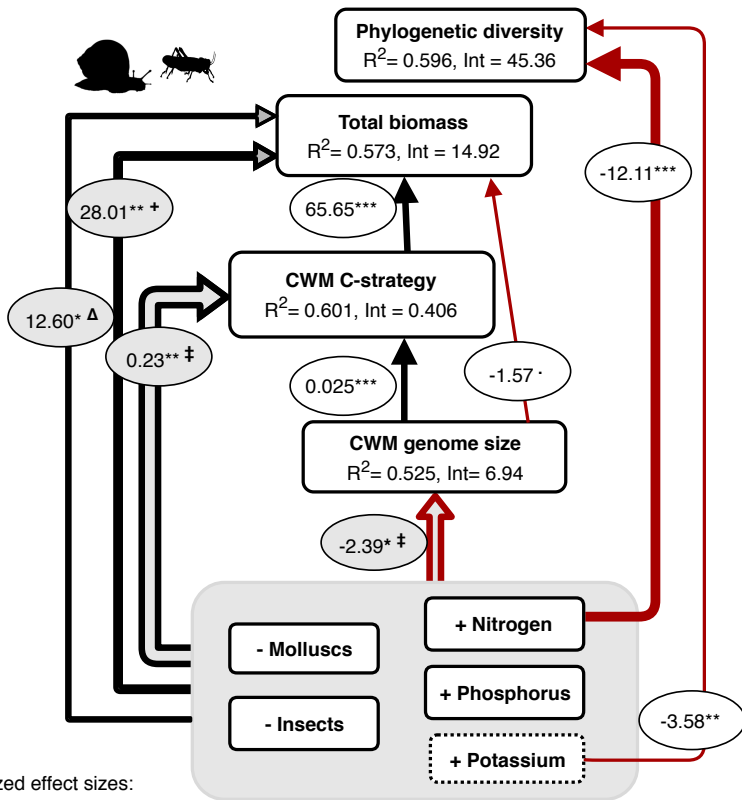
+ Two-way interaction: - rabbits * N

Black: Increasing effect

Red: Decreasing effect

Grey: Interaction

B

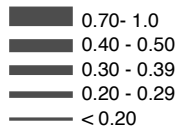


+ Two-way interaction: - molluscs * N

Δ Two-way interaction: - insects - molluscs

‡ Four-way interaction: - insects - molluscs * N * P

Standardized effect sizes:



SUPPLEMENTARY INFORMATION

Supplementary Text:

Phylogenetic tree data

Supporting figures SI.1 to SI.7

Fig. SI.1: Plot layout of experimental site

Fig. SI.2: Phylogenetic relationships of the 56 species in the data

Fig. SI.3: Directed acyclic graphs representing path model hypotheses

Fig. SI.4: Boxplots of three alternative estimations of community-weighted mean GS, including non-phylogenetic community-weighted mean GS.

Fig. SI.5: Boxplots showing total plot biomass, phylogenetic diversity, and community-weighted C-strategy.

Fig. SI.6: Scatterplots showing associations between community-weighted mean C-strategy and community-weighted mean C-value

Fig. SI.7: Non-metric multidimensional scaling (NMDS) biplots showing species distributions

Supporting tables SI.1 to SI.12

Table SI.1: Flow cytometry output:

(A) Summary

(B) Data for individual samples

Table SI.2: Species list: family, C-value (genome size), and C-strategy

Table SI.3: Path analysis conditional independence claims:

(A) \pm Rabbits

(B) \pm Insects \pm Molluscs

Table SI.4: CIC, C-statistics, p-values of path models

Table SI.5: MCMCglmm output:

(A) logistic

(B) linear

Table SI.6: Model reduction assessing community-weighted mean genome size

Table SI.7: LME output: community-weighted mean genome size

Table SI.8: Summary stats: community-weighted mean genome size, total biomass, species number, number of plots

Table SI.9: Model reduction assessing alternative estimations of community-weighted mean GS.

Table SI.10: Regression output for path models: \pm Rabbits

Table SI.11: Regression output for path models: \pm Insects \pm Molluscs

Table SI.12: Changes in species biomass with herbivore exclusion

SUPPLEMENTARY TEXT

Phylogenetic data (.tre format)

```
((((Luzula_campestris:0.4248200191,Juncus_effusus:0.4248200191):0.1805852116,(Carex_muricata:0.2934622637,Carex_hirta:0.2934622637):0.3119429669):0.04632828067,((Agrostis_capillaris:0.09725913344,(Anthoxanthum_odoratum:0.09104158341,(Trisetum_flavescens:0.08467450896,Arrhenatherum_elatius:0.08467450896):0.006367074449):0.006217550033):0.06543000109,(((Poa_annua:0.09472861627,(Poa_trivialis:0.07606022523,Poa_pratensis:0.07606022523):0.01866839104):0.01826946938,Phleum_pratense_subsp.bertolonii:0.1129980856):0.03557347513,(((Holcus_mollis:0.06599311723,Holcus_lanatus:0.06599311723):0.06455192943,Festuca_rubra:0.1305450467):0.002778278898,Dactylis_glomerata:0.1333233256):0.01524823522):0.01411757375):0.4890443768):0.3482664887,((Ranunculus_acris:0.09474228479,(Ranunculus_repens:0.01939019019,Ranunculus_bulbosus:0.01939019019):0.0753520946):0.8752854804,(((Viola_riviniiana:0.8572563263,(Lotus_corniculatus:0.4610654557,(Medicago_lupulina:0.2980689585,(Trifolium_dubium:0.2789913037,(Trifolium_pratense:0.295768755,Trifolium_repens:0.2295768755):0.04941442819):0.009715317616,Vicia_sativa_subsp.nigra:0.2887066213):0.009362337217):0.1629964972):0.3649087155,((Rubus_fruticosus:0.3578745186,(Potentilla_erecta:0.3055679279,Aphanes_microcarpa:0.3055679279):0.05230659071):0.4492908674,(Quercus_cerris:0.1035439427,Quercus_robur:0.1035439427):0.7036214434):0.01880878511):0.03128215515):0.03058731348,(Epilobium_ciliatum:0.8819912736,Malva_moschata:0.8819912736):0.00585236612):0.03863424171,(((Rumex_acetosa:0.1411790086,Rumex_acetosella:0.1411790086):0.5728411468,((Cerastium_fontanum:0.09480689681,(Stellaria_graminea:0.05426395017,Stellaria_media:0.05426395017):0.04054294664):0.4275584997,Chenopodium_album:0.5223653965):0.1916547589):0.1854017041,(((Galium_aparine:0.08744358065,(Galium_saxatile:0.07552341084,Galium_verum:0.07552341084):0.01192016982):0.6722013385,(Plantago_lanceolata:0.3177616705,(Veronica_arvensis:0.1058103944,Veronica_chamaedrys:0.1058103944):0.2119512761):0.4418832487):0.09261361901,(((Cirsium_arvense:0.1442739057,Centaurea_nigra:0.1442739057):0.1713946448,((Hieracium_pilosella:0.197383422,(Scorzoneroidees_autumnalis:0.0837967898,Hypochaeris_radicata:0.0837967898):0.09417060483,(Taraxacum_officinale:0.1229382555,Crepis_capillaris:0.1229382555):0.05502913918):0.01941602742):0.06938138647,(Jacobaea_vulgaris:0.2430958731,Achillea_millefolium:0.2430958731):0.0236689354):0.04890374194):0.4835595089,(Heracleum_sphondylium:0.7857858522,Sambucus_nigra:0.7857858522):0.01344220714):0.05303047879):0.04716332134):0.02705602196):0.04354988372):0.02997223481);
```

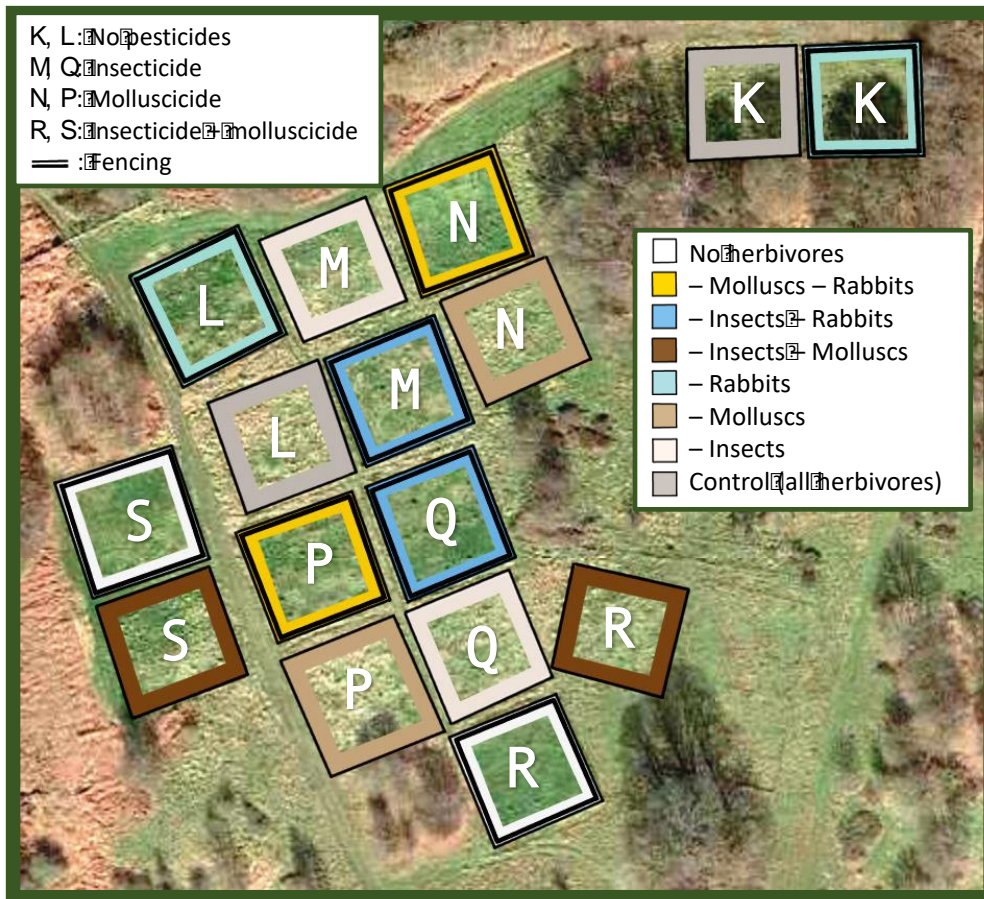


Fig. SI.1 Layout of experimental plots in Nash's Field, established in 1991 at Silwood Park (Imperial College, London). Further details are available at <http://www.imperial.ac.uk/silwood-park/research/silwood-lte/nashk-s/>.

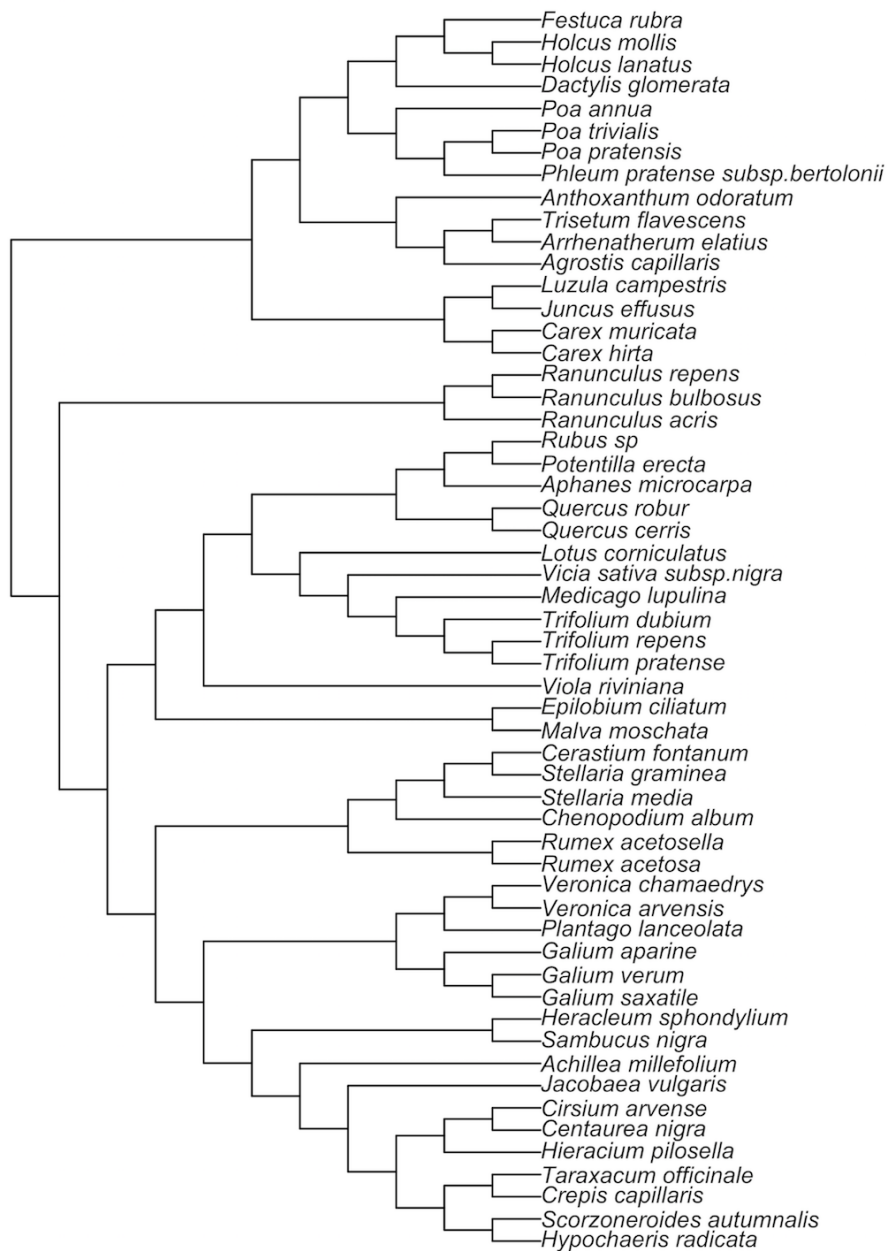


Fig. SI.2 Phylogenetic tree of the 56 species occurring on the 556 limed plots of Nash's Field, Silwood Park, UK.

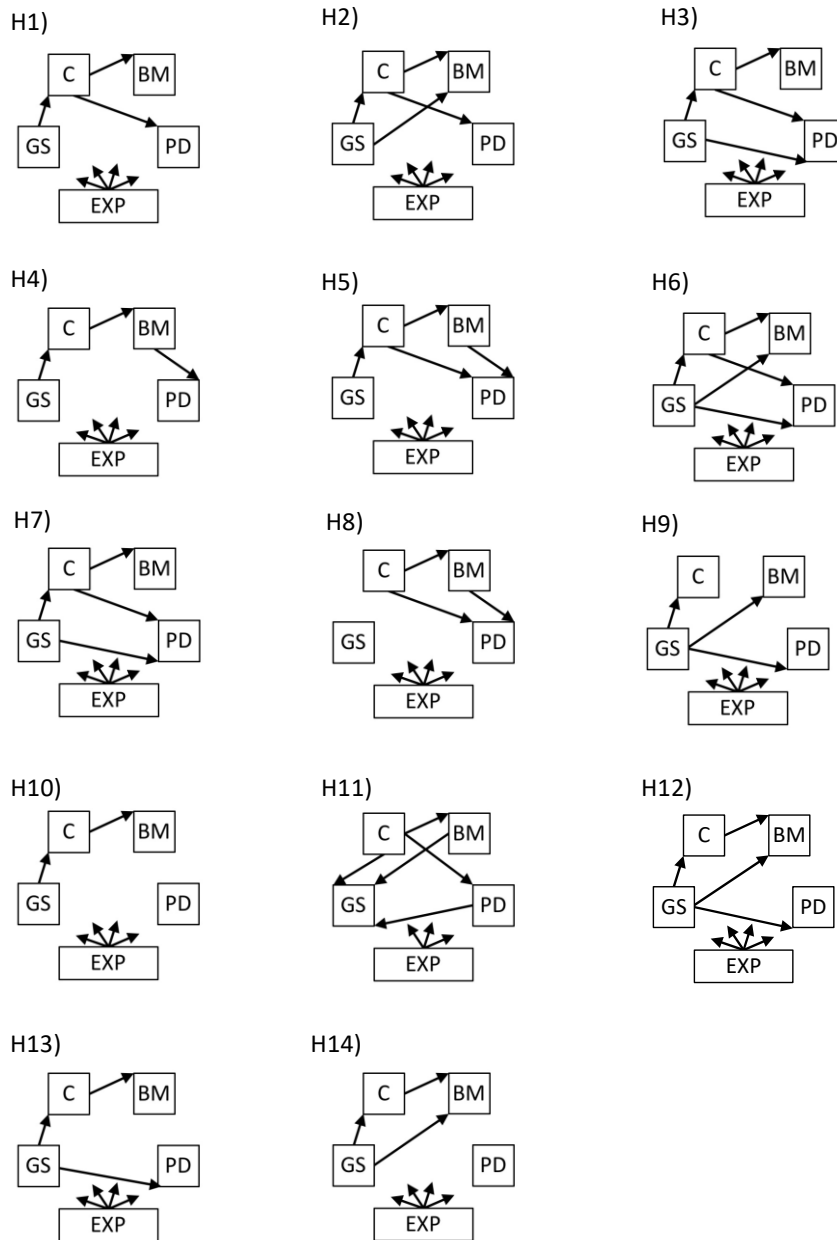


Fig. SI.3 Directed acyclic graphs representing path models investigating the effects exerted by the experiment on the four plant community parameters comprising (i) community-weighted mean genome size (GS), (ii) community-weighted mean competitive (C)-strategy (C), (iii) total community biomass (BM), and (iv) phylogenetic diversity (PD), and how these four properties are associated with each other. These hypotheses are built upon previous studies investigating the influence of GS on plant communities and from examining correlations present in the data. The small arrows pointing towards each variable represent the effect of experimental treatment (EXP: interactions between herbivore exclusion, N and P, plus K). An arrow originates from the experiment to each of the four community properties in all model hypotheses (diagrams H1-H14), except H11, where conditional independence was tested between the experiment and genome size (see also Table SI.3 for a more complete description of *d-sep* methods).

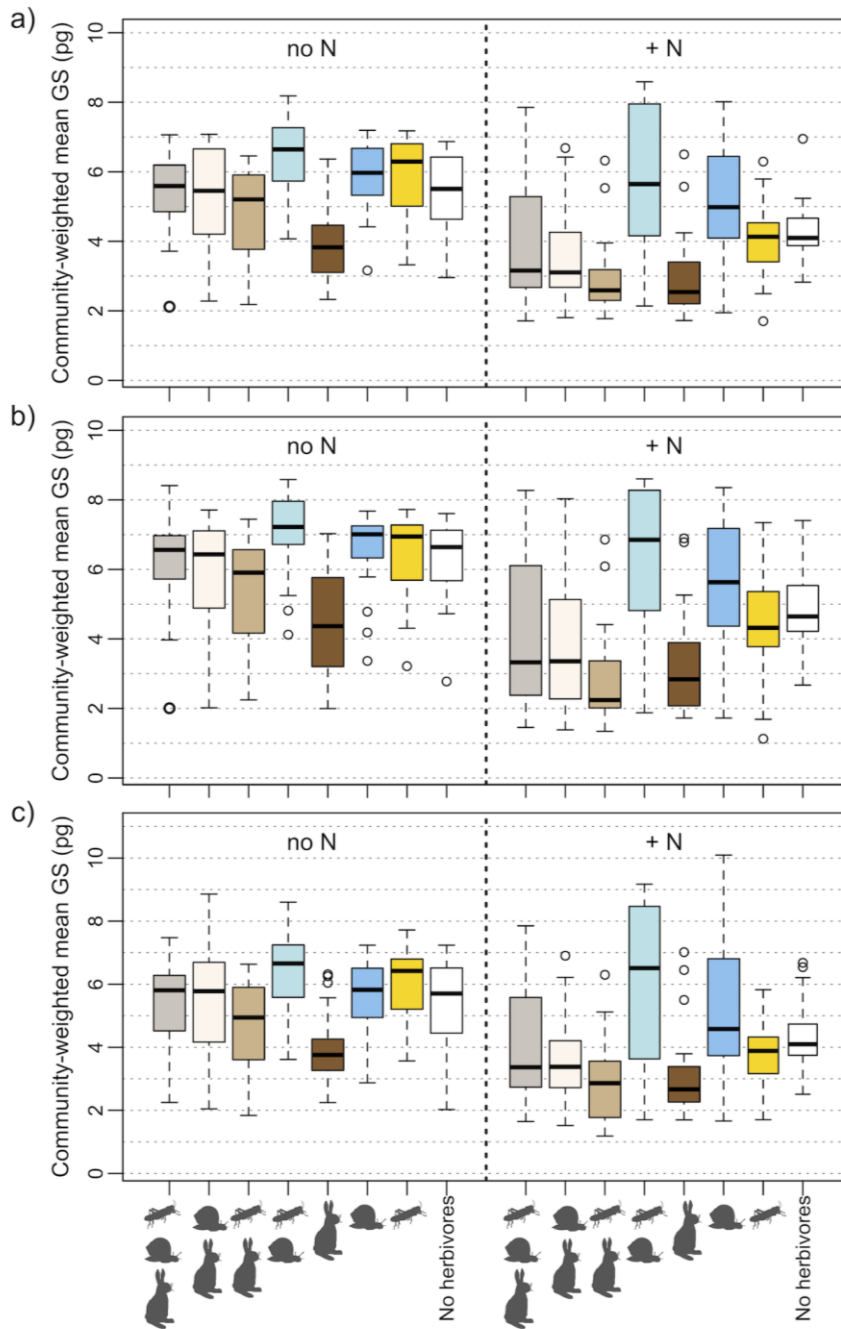


Fig. SI.4 Three alternative estimations of community-weighted mean GS (1C-value in picograms): **a)** simple weighted mean GS, without accounting for phylogenetic non-independence. As in the main text, species GS were log₁₀-transformed, and the weighted mean back-transformed; **b)** estimated with PGLS assuming a Brownian motion of evolution structure (as described in the main text); however GS was *not* transformed; **c)** estimated with PGLS with lambda optimisation, thus relaxing the assumptions of Brownian motion of evolution. As in the main text, species GS were log₁₀-transformed, and the community-weighted mean back-transformed. See also Table SI.9 for significance of experimental treatments (N fertilizer and herbivore exclusions) on community-weighted mean GS.

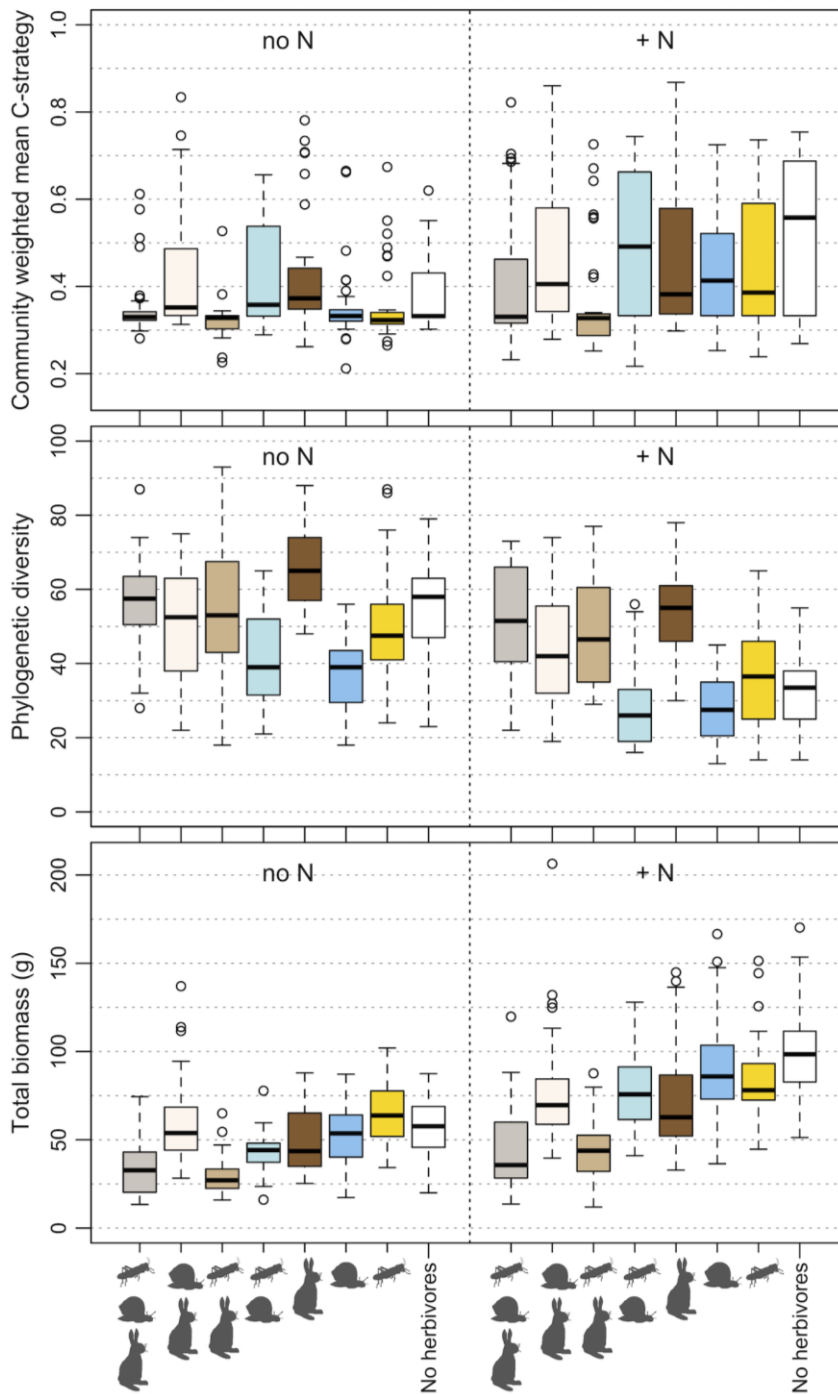


Fig. SI.5 Community-weighted mean competitive (C)-strategy; phylogenetic diversity (Faith's PD [1]), and total community biomass (dry weight g/m²) (n = 556), shown according to herbivore treatment. Herbivore treatments, in order as shown above are: 1) control (all herbivores present); 2) – insects; 3) – molluscs; 4) – rabbits; 5) rabbits only (– insects, – molluscs); 6) molluscs only (– insects, – rabbits); 7) insects only (– molluscs, – rabbits); 8) no herbivores. See also Table SI.8.

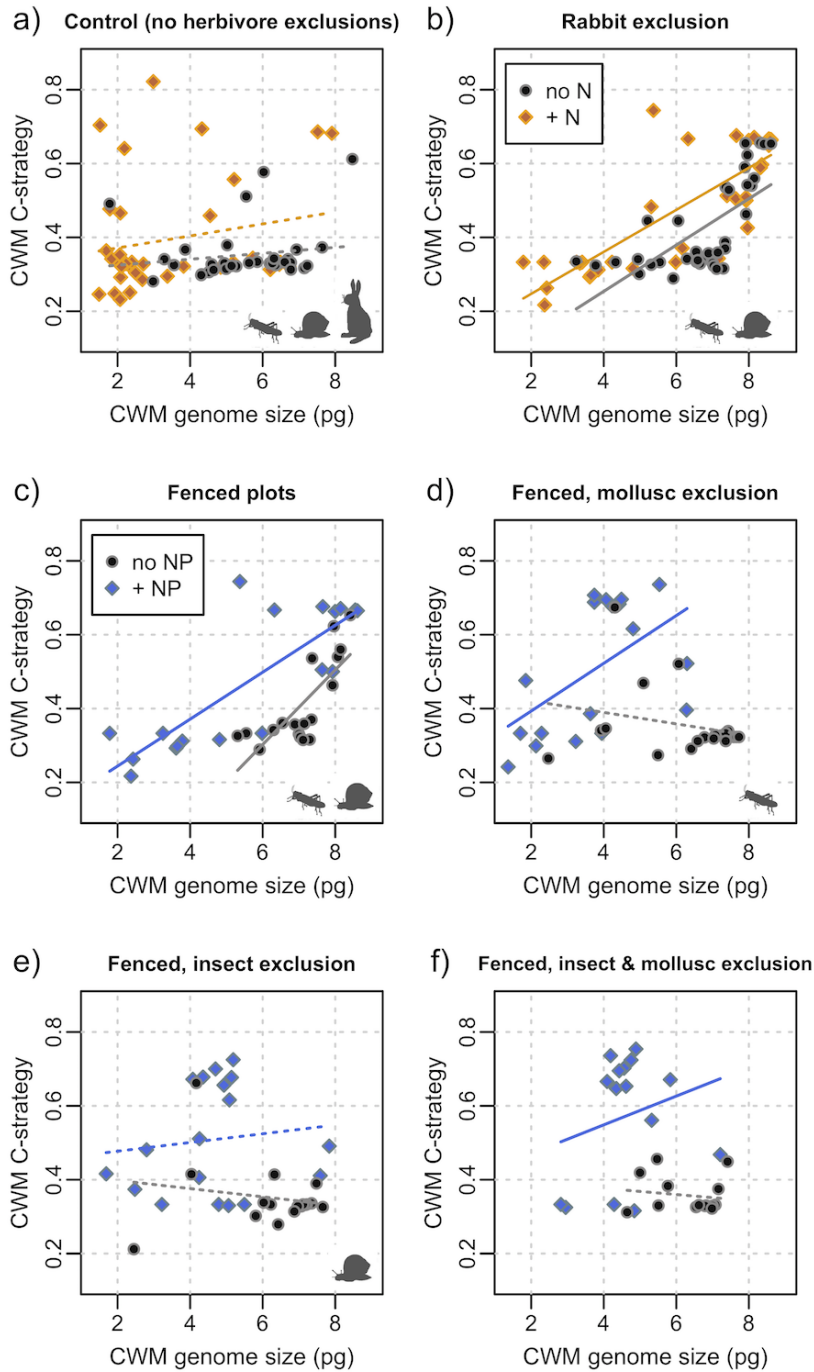


Fig. SI.6 Associations between community-weighted mean (CWM) C-strategy (CWM C-strategy) and CWM GS under different herbivore exclusion treatments and \pm N input. Herbivore exclusion treatments correspond to those of the path analyses: effects of rabbit herbivory (**a, b**); and effects of molluscs and insects on fenced plots (**c-f**). The rabbit herbivory path analysis showed that N input (but not P) had a significant influence on CWM GS and CWM C-strategy. In the path analysis on mollusc and insect herbivory, the 4-way interaction [–insects –molluscs N P] was significant, thus shown in **c-f** are plots without N and P input, and with both N and P input. Both community properties were estimated with PGLS, taking phylogeny into account. Trend lines were estimated with a simple bivariate linear model. A solid line shows a significant association ($p < 0.05$), a dashed line shows non-significance.

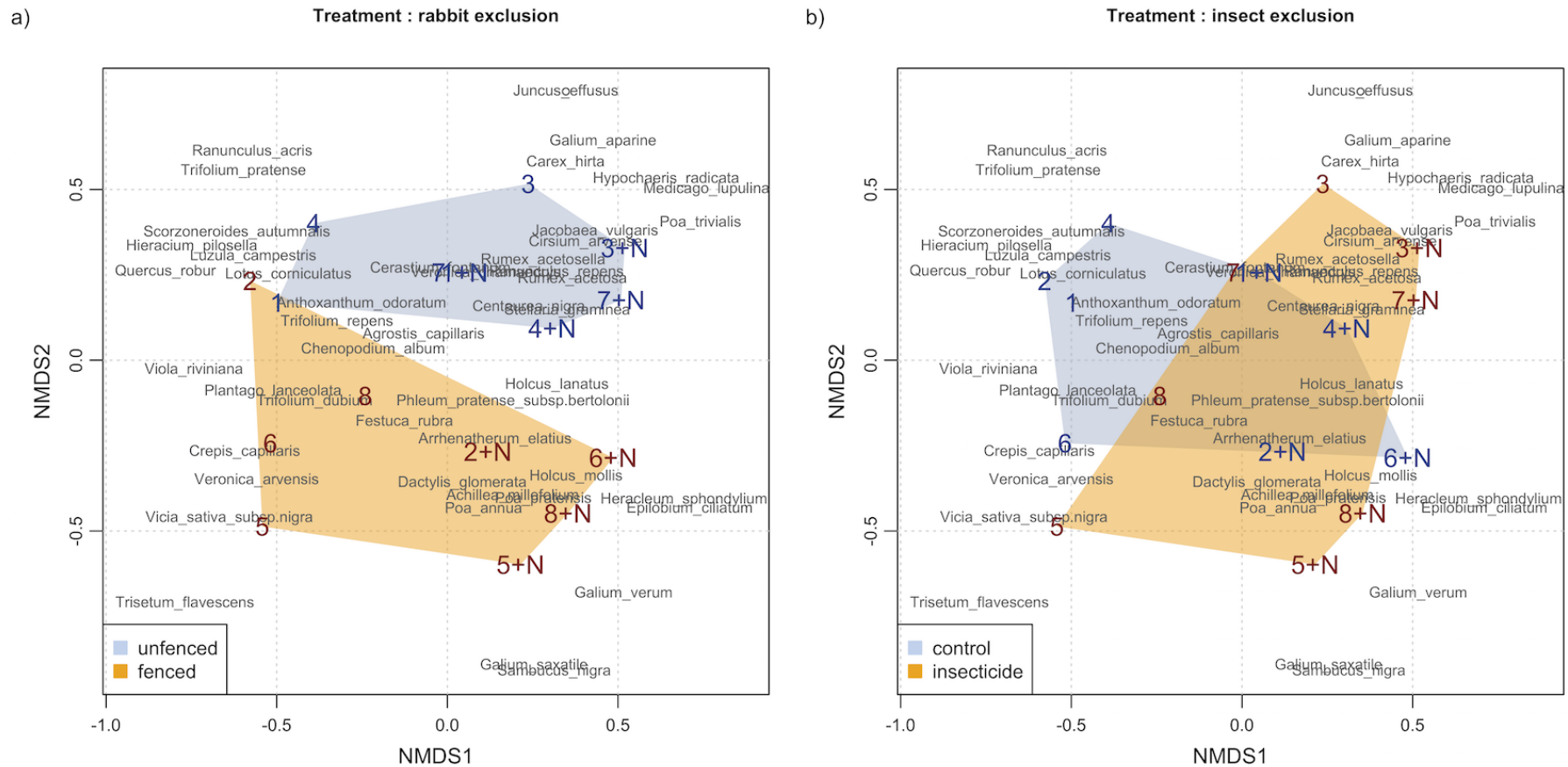


Fig. SI.7 Non-metric multidimensional scaling (NMDS) biplots showing species abundances in two-dimensional space. Stress = 0.173. Experimental treatments are coded as follows: 1 = control (no treatments); 2 = fencing; 3 = insecticide; 4 = molluscicide; 5 = insecticide + fencing; 6 = molluscicide + fencing; 7 = insecticide + molluscicide; 8 = no herbivores (insecticide + molluscicide + fencing). For each treatment, there is also an equivalent with N fertilizer (+ N). Convex hull polygons delineate experimental treatments and the plant species found within them: **a)** \pm fencing (rabbit exclusion); **b)** \pm insect exclusion; **c)** \pm mollusc exclusion; and **d)** \pm N fertilizer. See Table SI.12 for species lists with mean percent change in biomass.

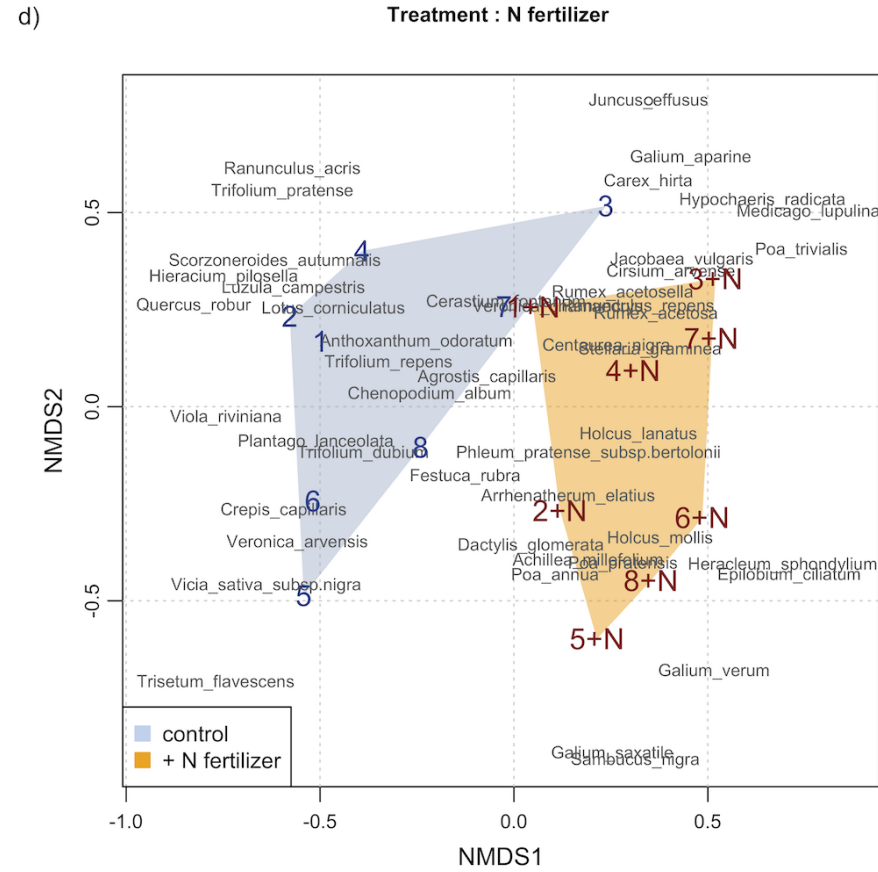
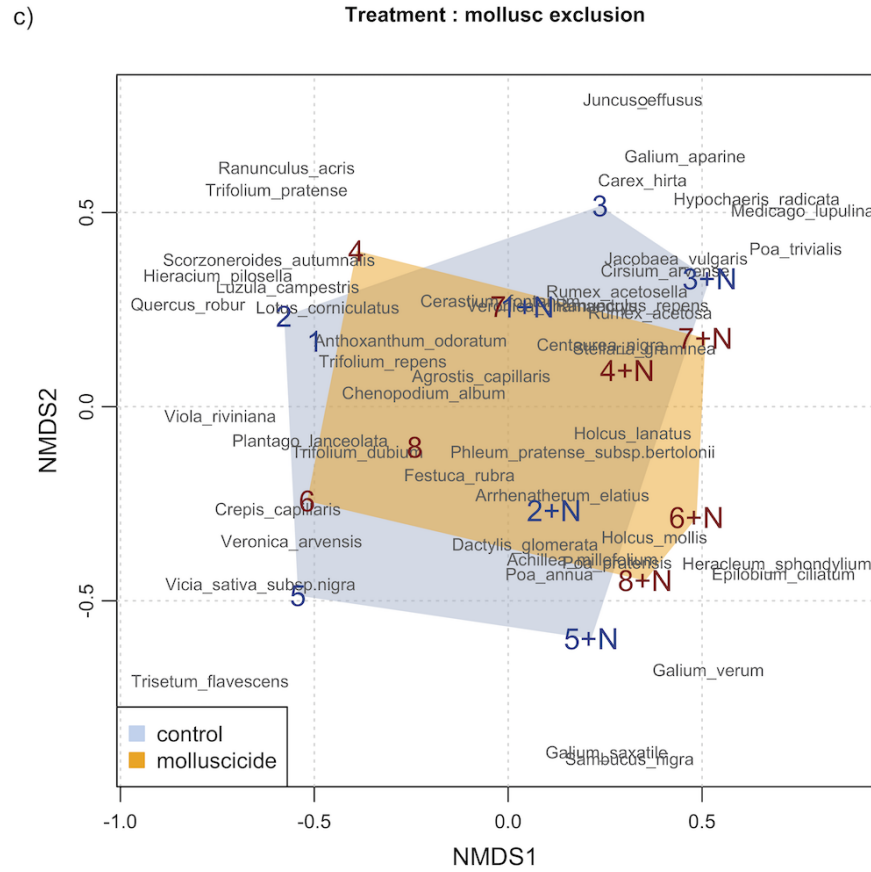


Table SI.1 (A) Flow cytometry results for 36 species collected at Nash’s field, Silwood Park, showing the mean estimated 1C-value in picograms (pg) obtained, number of plants measured (n), standard deviation in 1C-value (sdev), and mean target and standard coefficients of variation (CoV). 1C-values were estimated using a Partec CyFlow Space flow cytometer fitted with a Cobalt Samba green (532 nm, 100 mW) laser. Internal standards were either parsley (*Petroselinum crispum* “Champion Moss Curled”; 1C=2.22 pg), pea (*Pisum sativum* “Minerva Maple”; 1C=4.86 pg), tomato (*Solanum lycopersicum*, 1C=0.98 pg), or rice (*Oryza sativa*; 1C = 0.5 pg) (the 1C-values for the calibration standards were taken from Pellicer and Leitch [2]), and samples were prepared with Galbraith’s or LB01 buffers. For some species several flow cytometry runs were made using different calibration standards. These are indicated in the final column below, while the results of all individual flow cytometry runs used to calculate the Mean 1C-values together with the calibration standard used are given in Table SI.1 (B). Note that for species with a high CoV (e.g. > 5 - 8 %), we used the Prime C-value from the Plant DNA C-values database [3]. See Table SI.1 (B) for flow cytometry output of each sample run. The species’ C-values used in the statistical analyses are shown in Table SI.2.

Taxon	Mean 1C-value (pg)	n	Sdev	Mean target CoV	Mean standard CoV	Calibration standard(s)
<i>Achillea millefolium</i>	7.98	3	0.03	3.16	3.40	Pea
<i>Agrostis capillaris</i>	3.60	8	0.25	4.43	4.37	Parsley, pea
<i>Anthoxanthum odoratum</i>	7.28	3	0.36	4.05	4.27	Parsley, pea
<i>Arrhenatherum elatius</i>	8.58	3	0.47	3.73	4.71	Parsley, pea
<i>Carex muricata</i>	0.38	7	0.01	6.52	4.61	Parsley, rice, tomato
<i>Centaurea nigra</i>	2.13	2	0.29	7.17	6.23	Pea
<i>Cerastium fontanum</i>	3.23	2	0.20	4.29	3.68	Parsley, pea
<i>Chenopodium album</i>	1.95	1	NA	6.76	6.84	Rice
<i>Cirsium arvense</i>	1.48	1	NA	4.64	3.74	Pea
<i>Crepis capillaris</i>	2.45	2	0.13	8.50	5.81	Pea
<i>Dactylis glomerata</i>	4.44	2	0.21	4.60	5.20	Parsley
<i>Festuca rubra</i>	7.31	2	0.02	5.25	5.46	Pea
<i>Galium aparine</i>	1.11	1	NA	6.88	5.14	Parsley
<i>Galium saxatile</i>	1.72	1	NA	4.21	2.44	Pea
<i>Galium verum</i>	2.25	1	NA	6.99	8.67	Pea
<i>Heracleum sphondylium</i>	2.46	2	0.15	5.09	5.26	Rice, pea
<i>Hieracium pilosella</i>	3.52	2	0.05	6.46	6.95	Parsley
<i>Holcus lanatus</i>	1.70	1	NA	4.14	2.94	Parsley
<i>Holcus mollis</i>	4.03	3	0.04	6.30	7.31	Parsley
<i>Jacobaea vulgaris</i>	2.30	3	0.11	4.21	4.46	Rice, pea
<i>Juncus effusus</i>	0.28	2	0.02	5.37	4.61	Parsley, rice
<i>Lotus corniculatus</i>	1.30	4	0.05	4.75	4.41	Parsley, rice
<i>Luzula campestris</i>	0.40	2	0.01	5.54	4.32	Parsley, tomato
<i>Medicago lupulina</i>	0.55	1	NA	15.07	4.82	Parsley
<i>Phleum pratense</i> subsp. <i>bertolonii</i>	1.88	3	0.08	3.13	3.54	Parsley, pea, rice
<i>Plantago lanceolata</i>	1.43	2	0.03	4.99	4.79	Parsley, rice
<i>Poa trivialis</i>	2.01	4	0.02	4.41	3.68	Pea
<i>Ranunculus acris</i>	4.98	2	0.13	3.79	4.24	Parsley
<i>Ranunculus repens</i>	11.06	2	0.14	5.68	5.69	Pea

Table SI.1 (A) continued

Taxa	Mean 1C-value (pg)	n	Sdev	Mean target CoV	Mean standard CoV	Calibration standard(s)
<i>Rubus</i> sp.	0.87	1	NA	19.0	6.72	Parsley
<i>Rumex acetosella</i>	1.07	2	0.01	7.50	6.00	Parsley
<i>Stellaria graminea</i>	1.01	4	0.05	4.22	4.59	Rice, pea
<i>Trifolium repens</i>	1.12	2	0.02	6.41	6.11	Parsley
<i>Veronica chamaedrys</i>	2.16	2	0.02	5.02	3.80	Pea
<i>Vicia sativa</i> subsp. <i>nigra</i>	2.21	2	0.02	6.71	5.80	Pea
<i>Viola riviniana</i>	1.48	2	0.15	6.93	4.08	Parsley, pea

Table SI.1 (B) Flow cytometry output of individual samples run to estimate the mean 1C-values (pg) of the 36 species given in Table SI.1 (A). It gives the peak values of the target and standard obtained from the flow histograms. The table shows the standard used in each sample and the 1C-value (pg) of the calibration standard used. The target 1C-value is estimated as: (target peak/standard peak) x standard 1C-value. Also shown are the coefficients of variation (CoV) of the target and standard peaks, the estimated genome size (1C-value in pg) of the target.

Taxon	Standard	Standard 1C-value	Target peak	Standard peak	Target 1C-value	Target CoV	Standard Cov
<i>Achillea millefolium</i>	pea	4.86	385.12	235.34	7.95	3.58	3.48
<i>Achillea millefolium</i>	pea	4.86	392.75	239.76	7.96	3.64	3.14
<i>Achillea millefolium</i>	pea	4.86	415.56	251.92	8.02	2.27	3.57
<i>Agrostis capillaris</i>	parsley	2.22	163.54	110.84	3.28	4.58	4.72
<i>Agrostis capillaris</i>	parsley	2.22	287.30	188.10	3.39	3.83	3.60
<i>Agrostis capillaris</i>	parsley	2.22	262.84	166.78	3.50	4.89	5.15
<i>Agrostis capillaris</i>	parsley	2.22	314.98	199.81	3.50	4.51	4.35
<i>Agrostis capillaris</i>	parsley	2.22	198.60	124.31	3.55	4.51	4.51
<i>Agrostis capillaris</i>	parsley	2.22	182.58	111.21	3.64	4.17	4.95
<i>Agrostis capillaris</i>	pea	4.86	201.59	248.93	3.94	4.20	4.23
<i>Agrostis capillaris</i>	pea	4.86	135.91	166.02	3.98	4.77	3.44
<i>Anthoxan. odoratum</i>	parsley	2.22	345.25	110.73	6.92	3.52	4.42
<i>Anthoxan. odoratum</i>	parsley	2.22	346.00	105.60	7.27	4.86	3.56
<i>Anthoxan. odoratum</i>	pea	4.86	289.27	184.06	7.64	3.77	4.82
<i>Arrhenatherum elatius</i>	parsley	2.22	373.76	99.72	8.32	3.50	7.50
<i>Arrhenatherum elatius</i>	parsley	2.22	214.89	55.02	8.67	2.66	3.01
<i>Arrhenatherum elatius</i>	pea	4.86	253.04	140.42	8.76	5.03	3.63
<i>Carex muricata</i>	tomato	0.98	28.60	77.04	0.36	6.37	5.05
<i>Carex muricata</i>	rice	0.50	106.00	140.35	0.38	6.70	4.74
<i>Carex muricata</i>	rice	0.50	108.35	142.57	0.38	6.55	4.44
<i>Carex muricata</i>	parsley	2.22	18.01	104.53	0.38	6.35	4.97
<i>Carex muricata</i>	parsley	2.22	18.21	105.08	0.38	6.79	4.80
<i>Carex muricata</i>	parsley	2.22	18.14	104.03	0.39	6.52	4.91
<i>Carex muricata</i>	rice	0.50	44.81	57.70	0.39	6.36	3.36
<i>Centaurea nigra</i>	pea	4.86	54.17	137.08	1.92	6.41	5.44
<i>Centaurea nigra</i>	pea	4.86	82.71	172.37	2.33	7.93	7.01
<i>Cerastium fontanum</i>	parsley	2.22	96.75	69.51	3.09	5.00	4.52
<i>Cerastium fontanum</i>	pea	4.86	100.63	144.80	3.38	3.57	2.83
<i>Chenopodium album</i>	rice	0.50	143.70	36.78	1.95	6.76	6.84
<i>Cirsium arvense</i>	pea	4.86	58.16	191.51	1.48	4.64	3.74
<i>Crepis capillaris</i>	pea	4.86	100.14	206.88	2.35	10.37	6.06
<i>Crepis capillaris</i>	pea	4.86	78.48	150.03	2.54	6.63	5.55
<i>Dactylis glomerata</i>	parsley	2.22	232.63	120.30	4.29	5.05	5.55
<i>Dactylis glomerata</i>	parsley	2.22	192.52	93.11	4.59	4.15	4.84
<i>Festuca rubra</i>	pea	4.86	267.44	178.24	7.29	5.27	5.34
<i>Festuca rubra</i>	pea	4.86	247.31	164.12	7.32	5.22	5.94

Table SI.1 (B) continued

Taxon	Standard	Standard 1C-value	Target peak	Standard peak	Target 1C-value	Target CoV	Standard Cov
<i>Galium aparine</i>	parsley	2.22	68.14	136.37	1.11	6.88	5.14
<i>Galium saxatile</i>	pea	4.86	172.61	488.13	1.72	4.21	2.44
<i>Galium verum</i>	pea	4.86	67.42	145.49	2.25	6.99	8.67
<i>Heracleum sphondylium</i>	rice	0.50	232.10	49.37	2.35	4.72	5.25
<i>Heracleum sphondylium</i>	pea	4.86	78.63	148.95	2.57	5.46	5.26
<i>Hieracium pilosella</i>	parsley	2.22	221.55	141.21	3.48	6.98	8.36
<i>Hieracium pilosella</i>	parsley	2.22	247.57	154.92	3.55	5.94	5.54
<i>Holcus lanatus</i>	parsley	2.22	147.43	192.18	1.70	4.14	2.94
<i>Holcus mollis</i>	parsley	2.22	245.02	136.48	3.99	6.48	6.43
<i>Holcus mollis</i>	parsley	2.22	121.87	66.82	4.05	5.91	7.51
<i>Holcus mollis</i>	parsley	2.22	244.91	133.96	4.06	6.52	7.99
<i>Jacobaea vulgaris</i>	pea	4.86	72.75	161.09	2.19	4.55	2.60
<i>Jacobaea vulgaris</i>	rice	0.50	113.51	24.88	2.28	4.03	4.93
<i>Jacobaea vulgaris</i>	pea	4.86	97.01	194.88	2.42	4.05	5.86
<i>Juncus effusus</i>	rice	0.50	48.33	88.89	0.27	5.66	5.58
<i>Juncus effusus</i>	parsley	2.22	13.78	103.51	0.30	5.08	3.63
<i>Lotus corniculatus</i>	parsley	2.22	119.58	212.15	1.25	5.34	4.94
<i>Lotus corniculatus</i>	parsley	2.22	140.90	243.72	1.28	4.27	3.17
<i>Lotus corniculatus</i>	rice	0.50	142.91	54.75	1.31	6.17	5.13
<i>Lotus corniculatus</i>	parsley	2.22	153.43	248.56	1.37	3.20	4.40
<i>Luzula campestris</i>	tomato	0.98	24.97	63.08	0.39	5.51	5.16
<i>Luzula campestris</i>	parsley	2.22	21.58	119.88	0.40	5.56	3.48
<i>Medicago lupulina</i>	parsley	2.22	37.89	153.10	0.55	15.07	4.82
<i>Phleum pratense</i> subsp. <i>bertolonii</i>	parsley	2.22	111.21	135.99	1.82	4.31	3.54
<i>Phleum pratense</i> subsp. <i>bertolonii</i>	rice	0.50	223.27	59.97	1.86	2.32	3.89
<i>Phleum pratense</i> subsp. <i>bertolonii</i>	pea	4.86	57.84	142.53	1.97	2.76	3.18
<i>Plantago lanceolata</i>	rice	0.50	112.71	39.86	1.41	4.81	5.84
<i>Plantago lanceolata</i>	parsley	2.22	73.44	112.44	1.45	5.17	3.73
<i>Poa trivialis</i>	pea	4.86	82.92	204.13	1.97	4.37	3.57
<i>Poa trivialis</i>	pea	4.86	82.49	198.88	2.02	4.84	3.74
<i>Poa trivialis</i>	pea	4.86	87.99	212.11	2.02	4.97	4.60
<i>Poa trivialis</i>	pea	4.86	118.46	283.72	2.03	3.47	2.81
<i>Ranunculus acris</i>	parsley	2.22	213.90	97.20	4.89	4.01	3.84
<i>Ranunculus acris</i>	parsley	2.22	186.99	81.93	5.07	3.57	4.63
<i>Ranunculus repens</i>	pea	4.86	251.80	111.63	10.96	6.81	7.18
<i>Ranunculus repens</i>	pea	4.86	545.29	237.58	11.15	4.54	4.19
<i>Rubus</i> sp.	parsley	2.22	14.93	38.12	0.87	19.00	6.72
<i>Rumex acetosella</i>	parsley	2.22	67.70	141.90	1.06	7.03	4.50
<i>Rumex acetosella</i>	parsley	2.22	90.73	186.81	1.08	7.97	7.49
<i>Stellaria grmainea</i>	pea	4.86	53.99	244.42	1.07	3.87	3.44
<i>Stellaria grmainea</i>	rice	0.50	125.98	65.74	0.96	4.91	4.75
<i>Stellaria grmainea</i>	rice	0.50	117.22	58.72	1.00	4.30	5.58
<i>Stellaria grmainea</i>	rice	0.50	105.17	52.09	1.01	3.80	4.57

Table SI.1 (B) continued

Taxon	Standard	Standard 1C-value	Target peak	Standard peak	Target 1C-value	Target CoV	Standard Cov
<i>Trifolium repens</i>	parsley	2.22	71.51	143.44	1.11	6.51	6.53
<i>Trifolium repens</i>	parsley	2.22	75.88	148.80	1.13	6.30	5.68
<i>Veronica chamaedrys</i>	pea	4.86	88.21	199.93	2.14	4.65	3.30
<i>Veronica chamaedrys</i>	pea	4.86	112.32	250.77	2.18	5.39	4.29
<i>Vicia sativa</i> subsp. <i>nigra</i>	pea	4.86	86.23	191.09	2.19	7.20	6.15
<i>Vicia sativa</i> subsp. <i>nigra</i>	pea	4.86	90.78	198.60	2.22	6.21	5.44
<i>Viola riviniana</i>	parsley	2.22	92.46	150.05	1.37	5.96	3.31
<i>Viola riviniana</i>	pea	4.86	60.17	185.61	1.58	7.90	4.84

Table SI.2 Angiosperm Phylogeny Group IV family [4], genome size (1C-value), C-S-R type and competitive (C)-strategy for each species in the dataset (n=56). In bold are the 12 species with $\geq 1\%$ mean biomass on at least two herbivore exclusion treatment plots, and which comprise the data analysed using PGLMMs. Where applicable, the column “Dif.” shows the difference between the C-value that we used to analyse our data, and the C-value that we obtained with flow cytometry. When empty, this indicates we estimated the taxon’s C-value by flow cytometry following field sampling at Nash’s field, Silwood Park. The remaining 1C-values were obtained from the Plant DNA C-values database [3] (The original reference for the GS data if taken from the C-values database are given in the column labelled ‘Ref.’). They were taken from the database when our coefficient of variations were $>5-8\%$, or when we were unable to estimate a C-value with flow cytometry (difference = NA).

Taxon	Family (n=25)	C-S-R type	C-strategy	1C-value (pg)	Dif. (pg)	Ref.*
<i>Achillea millefolium</i>	Aster.	CSR	0.333	7.98	-	-
<i>Agrostis capillaris</i>	Poa.	CSR	0.333	3.60	-	-
<i>Anthoxanthum odoratum</i>	Poa.	SR/CSR	0.117	7.28	-	-
<i>Aphanes microcarpa</i>	Rosa.	SR	0	0.58	NA	1
<i>Arrhenatherum elatius</i>	Poa.	C/CSR	0.667	8.58	-	-
<i>Carex hirta</i>	Cyper.	C/CSR	0.667	0.53	NA	2
<i>Carex muricata</i>	Cyper.	S/CSR	0.167	0.38	-	-
<i>Centaurea nigra</i>	Aster.	CSR	0.333	1.80	0.33	3
<i>Cerastium fontanum</i>	Caryophy.	R/CSR	0.167	3.24	-	-
<i>Chenopodium album</i>	Amaranth.	CR	0.5	1.63	0.32	4
<i>Cirsium arvense</i>	Aster.	C	1	1.42	0.06	5
<i>Crepis capillaris</i>	Aster.	R/SR	0	2.10	0.35	6
<i>Dactylis glomerata</i>	Poa.	C/CSR	0.667	4.40	0.04	7, 8
<i>Epilobium ciliatum</i>	Onagr.	R/CSR	0.167	0.53	NA	9
<i>Festuca rubra</i>	Poa.	CSR	0.333	7.31	-	-
<i>Galium aparine</i>	Rubia.	CR	0.5	1.03	0.08	10
<i>Galium saxatile</i>	Rubia.	S/CSR	0.167	1.45	0.27	10
<i>Galium verum</i>	Rubia.	SC/CSR	0.417	1.89	0.36	11
<i>Heracleum sphondylium</i>	Api.	C/CSR	0.667	2.19	0.27	12
<i>Hieracium pilosella</i>	Aster.	S/CSR	0.167	3.45	0.07	13
<i>Holcus lanatus</i>	Poa.	CSR	0.333	1.70	-	-
<i>Holcus mollis</i>	Poa.	C/CSR	0.667	4.03	-	-
<i>Hypochaeris radicata</i>	Aster.	CSR	0.333	1.34	NA	14
<i>Jacobaea vulgaris</i>	Aster.	R/CR	0.417	2.25	0.05	10
<i>Juncus effusus</i>	Junca.	C/SC	0.75	0.3	-0.02	15
<i>Lotus corniculatus</i>	Faba.	S/CSR	0.167	1.3	-	-
<i>Luzula campestris</i>	Cyper.	S/CSR	0.167	0.49	-0.1	16
<i>Malva moschata</i>	Malva.	C/CSR	0.667	1.10	NA	28, 29
<i>Medicago lupulina</i>	Faba.	R/CSR	0.167	0.65	-0.1	11
<i>Phleum pratense</i> subsp. <i>bertolonii</i>	Poa.	CSR	0.167	1.88	-	-
<i>Plantago lanceolata</i>	Planta.	CSR	0.333	1.43	-	-
<i>Poa annua</i>	Poa.	R	0	2.88	NA	-

Table SI.2 continued

Taxon	Family (n=25)	C-S-R type	C-strategy	1C-value (pg)	Dif. (pg)	Ref.
<i>Poa pratensis</i>	Poa.	CSR	0.333	4.24	NA	17
<i>Poa trivialis</i>	Poa.	R/CSR	0.167	2.01	-	-
<i>Potentilla erecta</i>	Rosa.	S/CSR	0.167	0.45	NA	18
<i>Quercus cerris</i>	Faga.	SC	0.5	0.95	NA	19
<i>Quercus robur</i>	Faga.	SC	0.5	0.93	NA	20
<i>Ranunculus acris</i>	Ranun.	CSR	0.333	4.98	-	-
<i>Ranunculus bulbosus</i>	Ranun.	SR	0	5.63	NA	21
<i>Ranunculus repens</i>	Ranun.	CR	0.5	11.2	-0.14	21
<i>Rubus</i> sp.	Rosa.	SC	0.5	0.7	0.17	12, 30
<i>Rumex acetosa</i>	Polygon.	CSR	0.333	1.65	NA	22
<i>Rumex acetosella</i>	Polygon.	SR/CSR	0.117	1.68	-0.61	23
<i>Sambucus nigra</i>	Adoxa.	C	1	15.25	NA	24
<i>Scorzonerooides autumnalis</i>	Aster.	R/CSR	0.167	1.16	NA	12
<i>Stellaria graminea</i>	Caryophy.	CSR	0.333	1.01	-	-
<i>Stellaria media</i>	Caryophy.	R	0	1.05	NA	1
<i>Taraxacum officinale</i>	Aster.	R/CSR	0.167	1.28	NA	23
<i>Trifolium dubium</i>	Faba.	R/SR	0	0.73	NA	25
<i>Trifolium pratense</i>	Faba.	CSR	0.333	0.43	NA	25
<i>Trifolium repens</i>	Faba.	CR/CSR	0.417	1.12	-	25
<i>Trisetum flavescens</i>	Poa.	CSR	0.333	2.55	NA	10
<i>Veronica arvensis</i>	Planta.	SR	0	0.33	NA	26
<i>Veronica chamaedrys</i>	Planta.	CSR	0.333	2.16	-	-
<i>Vicia sativa</i> subsp. <i>nigra</i>	Faba.	R/CSR	0.25	2.25	-0.04	27
<i>Viola riviniana</i>	Viola.	S/CSR	0.167	1.35	0.12	3

Notes:

Dactylis glomerata: two references listed in the Plant DNA C-values database, both with the same C-value of 1C = 4.4 pg.

Malva moschata: mean of *M. parviflora* (28) and *M. sylvestris* (29).

Rubus sp.: mean of *R. chamaemorus* (30), *R. idaeus* (12), and of own measurement.

***Original sources for each C-value:**

1. Bennett MD. 1972. Nuclear DNA content and minimum generation time in herbaceous plants. Proceedings of the Royal Society of London Series B-Biological Sciences 181: 109-135.
2. Lipnerová I, Bureš P, Horová L, Šmarda P. 2013. Evolution of genome size in *Carex* (Cyperaceae) in relation to chromosome number and genomic base composition. Annals of Botany 111: 79-94.
3. Grime JP, Shacklock JML, Band SR. 1985. Nuclear DNA contents, shoot phenology and species co-existence in a limestone grassland community. New Phytologist 100: 435-445.
4. Ohri D. (pers. comm. 2002; Prime value, ref. 455)
5. Bureš P, Wang Y-F, Horova L, Suda J. 2004. Genome size variation in Central European species of *Cirsium* (Compositae) and their natural hybrids. Annals of Botany 94: 353-363.
6. Evans GM, Rees H, Snell CL, Sun S. 1972. The relationship between nuclear DNA amount and the duration of the mitotic cycle. Chromosomes Today 3: 24-31.
7. Creber HMC, Davies MS, Francis D, Walker HD. 1994. Variation in DNA C value in natural populations of *Dactylis glomerata* L. New Phytologist 128: 555-561.

8. Horjales M, Redondo N, P´erez B, Brown S. 1995. Presencia en Galicia de *Dactylis glomerata* L. Hexaploide. Boletim da Sociedade Broteriana (Ser. 2) 67: 223-230.
9. Bennett MD, Leitch IJ, Hanson L. 1998. DNA amounts in two samples of angiosperm weeds. Annals of Botany 82.
10. Band SR. (pers. comm. 1984; Prime value, ref 154)
11. Tensch EM, Tensch W, Ehrendorfer-Schratt L, Greilhuber J. 2010. Heavy metal pollution, selection, and genome size: The species of the Žerjav study revisited with flow cytometry. Journal of Botany 2010.
12. Siljak-Yakovlev, S., Pustahija, F., Šolić, E.M., Bogunić, F., Muratović, E., Bašić, N., Catrice, O. and Brown, S.C., 2010. Towards a genome size and chromosome number database of Balkan flora: C-values in 343 taxa with novel values for 242. Advanced Science Letters, 3(2), pp.190-213.
- 13: Suda J, Krahulcova A, Travnicek P, Rosenbaumova R, Peckert T, Krahulec F. 2007. Genome size variation and species relationships in *Hieracium* sub-genus *Pilosella* (Asteraceae) as inferred by flow cytometry. Annals of Botany 100: 1323-1335.
14. Cerbah, M., Coulaud, J., Brown, S.C. and Siljak-Yakovlev, S., 1999. Evolutionary DNA variation in the genus *Hypochaeris*. Heredity, 82(3), p.261.
15. Greilhuber J. 1988. "Self-tanning" - a new and important source of stoichiometric error in cytophotometric determination of nuclear DNA content in plants. Plant Systematics and Evolution 158: 87-96.
16. Bacic T, Jogan N, Dolenc Koce J. 2007. *Luzula* sect. *Luzula* in the south-eastern Alps - karyology and genome size. Taxon 56: 129-136.
17. Arumuganathan K, Tallury SP, Fraser ML, Bruneau AH, Qu R. 1999. Nuclear DNA content of thirteen turfgrass species by flow cytometry. Crop Science 39: 1518-1521.
18. Vidic T, Greilhuber J, Vilhar B, Dermastia M. 2009. Selective significance of genome size in a plant community with heavy metal pollution. Ecological Applications 19: 1515-1521.
19. Zoldos V, Papes D, Brown SC, Panaud O, Siljak-Yakovlev S. 1998. Genome size and base composition of seven *Quercus* species: inter- and intra-population variation. Genome 41: 162-168.
20. Favre JM, Brown S. 1996. A flow cytometric evaluation of the nuclear DNA content and GC percent in genomes of European oak species. Annales des Sciences Forestieres 53: 915-917.
21. Smith JB, Bennett MD. 1975. DNA variation in the genus *Ranunculus*. Heredity 35: 231-239.
22. Mowforth MAG. 1986. Variation in nuclear DNA amounts in flowering plants: an ecological analysis. Ph. D. University of Sheffield.
23. Bennett MD, Smith JB, Lewis Smith RI. 1982. DNA amounts of angiosperms from the Antarctic and South Georgia. Environmental and Experimental Botany 22: 307-318.
24. Nagl W, Jeanjour M, Kling H, Kuhner S, Michels I, Muller T, Stein B. 1983. Genome and chromatin organization in higher plants. Biologisches Zentralblatt 102: 129-148.
25. Vižintin, L., Javornik, B. and Bohanec, B., 2006. Genetic characterization of selected *Trifolium* species as revealed by nuclear DNA content and ITS rDNA region analysis. Plant Science, 170(4), pp.859-866.
26. Albach DC, Greilhuber J. 2004. Genome size variation and evolution in *Veronica*. Annals of Botany 94: 897-911.
27. Rees H, Cameron FM, Hazarika MH, Jones GH. 1966. Nuclear variation between diploid angiosperms. Nature 211: 828- 830.
28. Bidak L, Brandham PE. 1995. Intraspecific uniformity of chromosome number and nuclear DNA quantity in two Egyptian weedy species, *Malva parviflora* (Malvaceae) and *Trigonella stellata* (Leguminosae). Kew Bulletin 50: 595-599.
29. Ceccarelli M, Morosi L, Cionini PG. 1998. Chromocenter association in plant cell nuclei: determinants, functional significance, and evolutionary implications. Genome 41: 96-103.
30. Thiem B, Sliwiska E. 2003. Flow cytometric analysis of nuclear DNA content in cloudberry (*Rubus chamaemorus* L.) in vitro cultures. Plant Science 164: 129-134.

Table SI.3 Conditional independence claims tested in the context of path analyses. Each number refers to a hypothetical directed acyclic path diagram (Fig. SI.3).

The *d-sep* path analysis method tests conditional independence between parameters in a path diagram. For example, the conditional independence between community-weighted mean genome size and community-weighted mean C-strategy, given the experimental treatment (i.e. herbivory and nutrient input), can be drawn as genome size \leftarrow treatment \rightarrow competition, and written as: (genome size, competition) | {treatment}[3]. Independence between these two community properties is tested while holding experimental treatment constant i.e.: community-weighted mean C-strategy is a function of herbivory, nutrient treatment, and community-weighted mean genome size (competition \sim treatment + genome size). If the p-value for the coefficient of community-weighted mean genome size is below the alpha level ($p \leq 0.05$), this indicates that genome size and C-strategy are not independent of each other for a given experimental treatment.

Each conditional independence claim (set of parameters *not* connected by a path) in a path model is tested in this way to calculate its p-value. A variable may function as a dependent variable (e.g. C-strategy as a function of genome size and the experimental treatments), or as predictor (e.g. C-strategy as a predictor of total biomass). Fisher's C-statistic [5] is calculated from the p-values of the conditional independencies and the Chi-square distributed parameter k , which is equal to the number of conditional independencies in the model. The hypothetical path model is rejected when the C-statistic is below the alpha p-value, meaning that useful information is contained in one or more of the missing paths. Another goodness-of-fit statistic is the C-statistic Information Criterion (CICc) [6, 7] which takes into account sample size and the number of parameters in the model.

The experimental treatments were fitted as N x P x herbivore exclusion + K + herbicide (where each is a binary factor, except for herbicide, which is a three-level factor). In terms of the coefficients and p-values returned for each community property, this is equivalent to scoring the experiment as a single factor variable with eight levels in the rabbit exclusion experiment and 16 levels in the \pm insects \pm molluscs experiment, with additional information on their interactions, while controlling for K and herbicide treatments. We thus have a total of five parameters: the experiment, community-weighted mean genome size, community-weighted mean C-strategy, total biomass, and phylogenetic diversity.

Shown below are the p-values obtained for each conditional independence claim, which were tested with generalised least squares (GLS) with ten different variance structures (varID): 1) plot; 2) N; 3) P; 4) herbivore treatment (HT); 5) N + herbivore treatment; 6) N + P; 7) plot * N; 8) herbivore treatment * N; 9) plot * herbivore treatment * N; 10) herbivore treatment * P * N; and, 11) no variance structure. If the varID column = na, no variance structures were applied. This was assessed by whether it contributed to 1) a lower AICc, and 2) whether the difference in AICc was significant ($p \leq 0.05$) with an ANOVA test. GS= community-weighted mean

genome size of each plot, PD= phylogenetic diversity, BM= total plot biomass, C = mean weighted competitive (C)- strategy of each plot also estimated by PGLS; exp = experimental treatment, i.e.: herbivore * N * P + K. HT = herbivore treatment.

Table SI.3 (A) ± Rabbits:

			± Rabbits			
No.	Conditional independence claim	Claim test	p-value: LME	p-value: LME + varID	varID	ANOVA
1	(GS, PD) exp, C	PD ~ exp + C + GS	0.3594	na	na	na
	(GS, BM) exp, C	BM ~ exp + C + GS	0.5934	0.7363	N+P	<0.0001
	(BM, PD) exp, C	PD ~ exp + C + BM	0.5556	na	na	na
2	(GS, PD) exp, C	PD ~ exp + C + GS	0.3594	na	na	na
	(BM, PD) exp, GS, C	PD ~ exp + GS + C + BM	0.5664	na	na	na
3	(GS, BM) exp, C	BM ~ exp + C + GS	0.5934	0.7363	N+P	<0.0001
	(BM, PD) exp, GS, C	PD ~ exp + GS + C + BM	0.5664	na	na	na
4	(GS, PD) exp, BM	PD ~ exp + BM + GS	0.1101	na	na	na
	(GS, BM) exp, C	BM ~ exp + C + GS	0.5934	0.7363	N+P	<0.0001
	(C, PD) exp, GS, BM	PD ~ exp + GS + BM + C	0.3084	na	na	na
5	(GS, PD) exp, C, BM	PD ~ exp + C + BM + GS	0.3799	na	na	na
	(GS, BM) exp, C	BM ~ exp + C + GS	0.5934	0.7363	N+P	<0.0001
6	(BM, PD) exp, GS, C	PD ~ exp + GS + C + BM	0.5664	na	na	na
7	(GS, BM) exp, C	BM ~ exp + C + GS	0.5934	0.7363	N+P	<0.0001

Table SI.3 (A) continued ± Rabbits

8	(GS, PD) exp, C	PD ~ exp + C + GS	0.3594	na	na	na
	(GS, C) exp	C ~ exp + GS	<0.0001	0.0043	plot*HT*N	<0.0001
	(GS, BM) exp, C	BM ~ exp + C + GS	0.5934	0.7363	N+P	<0.0001
9	(BM, PD) exp, C	PD ~ exp + C + BM	0.5556	na	na	na
	(C, PD) exp, GS	PD ~ exp + GS + C	0.1547	na	na	na
	(C, BM) exp, GS	BM ~ exp + GS + C	0.0055	0.0088	N+P	<0.0001
10	(GS, PD) exp	PD ~ exp + GS	0.0485	na	na	na
	(BM, PD) exp, C	PD ~ exp + C + BM	0.5556	na	na	na
	(GS, BM) exp, C	BM ~ exp + C + GS	0.5934	0.7363	N+P	<0.0001
	(C, PD) exp, GS	PD ~ exp + GS + C	0.1547	na	na	na
11	(BM, PD) exp, C	PD ~ exp + C + BM	0.5556	na	na	na
	(exp, GS) PD, C, BM	GS ~ PD + C + BM + exp	<0.0001	<0.0001	plot*HT*N	<0.0001
12	(C, PD) exp, GS	PD ~ exp + GS + C	0.1547	na	na	na
	(BM, PD) exp, GS, C	PD ~ exp + GS + C + BM	0.5664	na	na	na
13	(BM, PD) exp, GS, C	PD ~ exp + GS + C + BM	0.5664	na	na	na
	(GS, BM) exp, C	BM ~ exp + C + GS	0.5934	0.7363	N+P	<0.0001
	(C, PD) exp, GS	PD ~ exp + GS + C	0.1547	na	na	na
14	(GS, PD) exp	PD ~ exp + GS	0.0485	na	na	na
	(BM, PD) exp, GS, C	PD ~ exp + GS + C + BM	0.5664	na	na	na
	(C, PD) exp, GS	PD ~ exp + GS + C	0.1547	na	na	na

Table SI.3 (B) Conditional independence claim tests on plots with \pm insects \pm molluscs. See Fig. SI.3 for path diagrams being tested below.

\pm insects \pm molluscs						
No.	Conditional independence claim	Claim test	p-value: LME	p-value: LME + varID	varID	ANOVA
1	(GS, PD) exp, C	PD \sim exp + C + GS	0.1395	0.0648	HT	0.0163
	(GS, BM) exp, C	BM \sim exp + C + GS	0.0625	0.5167	N*P	<0.0001
	(BM, PD) exp, C	PD \sim exp + C + BM	0.2439	0.2556	HT	0.0322
2	(GS, PD) exp, C	PD \sim exp + C + GS	0.1395	0.0648	HT	0.0163
	(BM, PD) exp, GS, C	PD \sim exp + GS + C + BM	0.1625	0.1778	HT	0.0163
3	(GS, BM) exp, C	BM \sim exp + C + GS	0.0625	0.5167	N*P	<0.0001
	(BM, PD) exp, GS, C	PD \sim exp + GS + C + BM	0.1625	0.1778	HT	0.0163
4	(GS, PD) exp, BM	PD \sim exp + BM + GS	0.0587	0.1018	HT	0.0161
	(GS, BM) exp, C	BM \sim exp + C + GS	0.0625	0.5167	N*P	<0.0001
	(C, PD) exp, GS, BM	PD \sim exp + GS + BM + C	0.7011	0.7335	HT	0.0163
5	(GS, PD) exp, C, BM	PD \sim exp + C + BM + GS	0.1025	0.0473	HT	0.0163
	(GS, BM) exp, C	BM \sim exp + C + GS	0.0625	0.5167	N*P	<0.0001
6	(BM, PD) exp, GS, C	PD \sim exp + GS + C + BM	0.1625	0.1778	HT	0.0163
7	(GS, BM) exp, C	BM \sim exp + C + GS	0.0625	0.5167	N*P	<0.0001

Table SI.3 (B) continued \pm insects \pm molluscs

8	(GS, PD) exp, C	PD ~ exp + C + GS	0.1395	0.0648	HT	0.0163
	(GS, C) exp	C ~ exp + GS	<0.0001	0	plot*N*P	<0.0001
	(GS, BM) exp, C	BM ~ exp + C + GS	0.0625	0.5167	N*P	<0.0001
9	(BM, PD) exp, C	PD ~ exp + C + BM	0.2439	0.2556	HT	0.0322
	(C, PD) exp, GS	PD ~ exp + GS + C	0.3676	0.8739	HT	0.0163
	(C, BM) exp, GS	BM ~ exp + GS + C	<0.0001	0	N*P	<0.0001
10	(GS, PD) exp	PD ~ exp + GS	0.0593	0.0449	HT	0.0116
	(BM, PD) exp, C	PD ~ exp + C + BM	0.2439	0.2556	HT	0.0322
	(GS, BM) exp, C	BM ~ exp + C + GS	0.0625	0.5167	N*P	<0.0001
	(C, PD) exp, GS	PD ~ exp + GS + C	0.3676	0.8739	HT	0.0163
11	(BM, PD) exp, C	PD ~ exp + C + BM	0.2439	0.2556	HT	0.0322
	(exp, GS) PD, C, BM	GS ~ PD + C + BM + exp	<0.0001	<0.0001	plot+N	0.0006
12	(C, PD) exp, GS	PD ~ exp + GS + C	0.3676	0.8739	HT	0.0163
	(BM, PD) exp, GS, C	PD ~ exp + GS + C + BM	0.1625	0.1778	HT	0.0163
13	(BM, PD) exp, GS, C	PD ~ exp + GS + C + BM	0.1625	0.1778	HT	0.0163
	(GS, BM) exp, C	BM ~ exp + C + GS	0.0625	0.5167	N*P	<0.0001
	(C, PD) exp, GS	PD ~ exp + GS + C	0.3676	0.8739	HT	0.0163
14	(GS, PD) exp	PD ~ exp + GS	0.0593	0.0449	HT	0.0116
	(BM, PD) exp, GS, C	PD ~ exp + GS + C + BM	0.1625	0.1778	HT	0.0163
	(C, PD) exp, GS	PD ~ exp + GS + C	0.3676	0.8739	HT	0.0163

Table SI.4 Summary path model goodness-of-fit statistics: Fisher’s C-statistic and CICc (C-statistic information criterion) and p-values. P-values above the alpha value (0.05) indicate the conditional independencies are satisfied and the model is a plausible model. The column “V, A” shows the number of vertices and number of arrows for each acyclic path diagram.

± Rabbits					± insect ± molluscs				
No.	V, A	C	p-value	CIC	No.	V, A	C	p-value	CIC
1	5, 7	4.266	0.641	30.648	1	5, 7	12.306	0.055	37.47
2	5, 8	3.184	0.528	31.984	2	5, 8	7.573	0.109	34.937
3	5, 8	2.181	0.703	30.981	3	5, 8	9.178	0.057	36.542
4	5, 7	7.808	0.931	34.19	4	5, 7	11.926	0.064	37.09
5	5, 8	2.979	0.561	31.779	5	5, 8	10.101	0.039	37.464
6	5, 9	1.137	0.566	32.393	6	5, 9	3.634	0.163	33.213
7	5, 9	1.044	0.593	32.3	7	5, 9	5.545	0.163	35.123
8	5, 7	21.511	0.001	47.893	8	5, 7	32.51	<0.0001	57.674
9	5, 7	15.328	0.018	41.71	9	5, 7	27.85	<0.0001	53.014
10	5, 6	12.005	0.151	36.005	10	5, 6	16.019	0.042	39
11	5, 8	19.596	0.001	48.396	11	5, 8	21.243	<0.0001	48.606
12	5, 8	4.869	0.301	32.232	12	5, 8	5.636	0.228	32.999
13	5, 7	5.913	0.433	32.295	13	5, 7	11.18	0.083	36.344
14	5, 7	10.923	0.091	36.087	14	5, 7	11.286	0.08	36.45

Table SI.5 (A) Logistic PGLMM: MCMCglmm output where absence/presence (0/1) of the abundant species (n=12) were fitted as a function of genome size (GS), herbivore and nutrient treatment. Insect, rabbit and mollusc exclusion, N, P, and K are scored as \pm binary factors. Baseline levels are the untreated plots (i.e. without fencing, without insecticide, without molluscicide and without fertilizer). Random effects (G-structure) account for experimental design, repeated measurements, and phylogenetic correlation. The posterior mean shows the log odds; and means (categorical variables) and slopes (continuous variables) were estimated as the sum of the posterior means for all parameters involved: e.g.: the slope of $-rabbits : GS = [GS + (-rabbits:GS)]$. These values were transformed with inverse logit [4] to obtain the probability. “:” in the fixed effects denotes an interaction.

G-structure:	Posterior mean	95% Credible intervals	Eff. sample size			
plot	0.614	3.11E-04, 2.10	4940			
plot : fencing	0.327	2.77E-04, 1.06	4940			
species	5.291	3.57E-04, 13.38	4940			
phylogeny	5.781	2.02E-04, 25.07	4940			

Fixed effects:	Posterior mean	95% Credible intervals	Eff. sample size	pMCMC	Mean or slope	Probability
Intercept	-0.659	-4, 2.99	4940	0.6332	-0.659	0.341
GS	-0.028	-0.84, 0.72	4940	0.9490	-0.028	0.493
- insects	2.127	0.24, 4.27	4940	0.0425	1.469	0.813
N	-0.401	-1.04, 0.25	4572	0.2304	-1.060	0.257
- molluscs	0.782	-1.2, 2.7	4550	0.3818	0.123	0.531
- rabbits	-1.207	-2.61, 0.06	4940	0.0664	-1.866	0.134
P	0.061	-0.34, 0.48	4940	0.7696	-0.598	0.355
K	-0.241	-0.52, 0.04	4940	0.0850	-0.900	0.289
- insects : GS	-0.032	-0.2, 0.14	5153	0.6988	-0.060	0.485
GS : N	0.140	-0.12, 0.41	5288	0.2988	0.113	0.528
- insects : N	-0.431	-1, 0.19	4940	0.1672	0.636	0.654
- molluscs : GS	-0.266	-0.44, -0.11	4940	0.0020	-0.294	0.427
- molluscs : N	0.357	-0.2, 0.96	4913	0.2308	0.079	0.520
- rabbits : GS	0.322	0.15, 0.49	4940	< 0.0002	0.294	0.573
- rabbits : N	-0.269	-0.8, 0.36	4940	0.3563	-2.537	0.073
GS : P	0.096	-0.08, 0.26	4940	0.2502	0.068	0.517
N : P	-0.247	-0.84, 0.29	4940	0.3968	-1.246	0.223
- insects - molluscs	-1.149	-3.96, 1.52	4687	0.3458	1.102	0.751
- molluscs - rabbits	0.920	-1.21, 2.61	4940	0.2413	-0.164	0.459
- insects - rabbits	-0.784	-2.88, 0.97	4940	0.3057	-0.523	0.372
- insects : N : GS	0.124	-0.11, 0.36	4940	0.3008	0.205	0.551
- molluscs : N : GS	-0.018	-0.26, 0.2	4940	0.8656	-0.172	0.457
- rabbits : GS : N	0.054	-0.18, 0.28	4940	0.6526	0.488	0.620
GS : N : P	-0.121	-0.35, 0.12	4940	0.3219	0.088	0.522
-insects -molluscs -rabbits	0.375	-2.15, 3.17	4940	0.7194	0.405	0.600

Table SI.5 (B) Linear PGLMM: MCMCglmm output in which species biomass > 0 was fitted with a Gaussian distribution. Biomass was log-transformed. G-structure shows the variance of the random effects. R-structure = residual variance. GS = genome size. The effects of each category were estimated as the combined effect of all posterior means (coefficients) involved; e.g. change in slope for GS:N:P = GS:N + GS:P + GS:N:P. These effect values were then exponentiated to estimate percent change, where change in percent = 100(exp(effect) – 1).

G-structure:	Posterior mean	95% Credible intervals	Eff. sample size			
plot	0.041	1.49E-04, 0.17	4940			
plot : fencing	0.145	1.85E-04, 0.42	4940			
species	0.391	1.79E-04, 1.75	4940			
phylogeny	1.826	3.73E-04, 4.56	4598			

R-structure:	Posterior mean	95% Credible intervals	Eff. sample size			
units	3.797	3.46, 4.14	5284			

Fixed effects:	Posterior mean	95% Credible intervals	Eff. sample size	pMCMC	Effect	exp(Effect)
Intercept	-0.955	-2.8, 0.71	4940	0.2397	-0.955	0.3850
GS	0.482	0.17, 0.86	4940	0.0057	0.482	1.6187
– insects	0.240	-0.74, 1.3	4940	0.6061	0.240	1.2716
N	0.845	0.21, 1.53	4940	0.0105	0.845	2.3291
– molluscs	0.667	-0.31, 1.73	4423	0.1575	0.667	1.9479
– rabbits	-0.740	-1.7, 0.31	5049	0.1458	-0.740	0.4772
P	0.217	-0.19, 0.6	4940	0.2858	0.217	1.2428
K	-0.172	-0.42, 0.07	4940	0.1891	-0.172	0.8423
– insects : GS	-0.108	-0.26, 0.04	4940	0.1640	-0.108	0.8976
GS : N	-0.268	-0.5, -0.03	4940	0.0243	-0.268	0.7653
– insects : N	0.085	-0.53, 0.62	4940	0.7806	1.171	3.2245
– molluscs : GS	-0.150	-0.31, -0.01	4940	0.0466	-0.150	0.8604
– molluscs : N	-0.459	-1.04, 0.11	4940	0.1178	1.053	2.8660
– rabbits : GS	0.191	0.05, 0.33	4940	0.0117	0.191	1.2099
– rabbits : N	-0.108	-0.68, 0.46	4940	0.7130	-0.002	0.9977
GS : P	-0.053	-0.2, 0.09	4940	0.4830	-0.053	0.9484
N : P	0.238	-0.35, 0.81	5100	0.4198	1.301	3.6735
– insects – molluscs	-0.190	-1.54, 1.11	4940	0.7696	0.717	2.0489
– molluscs – rabbits	0.206	-1.13, 1.53	4940	0.7235	0.133	1.1417
– insects – rabbits	-0.035	-1.43, 1.25	4940	0.9696	-0.535	0.5859
– insects : N : GS	0.012	-0.18, 0.23	4940	0.9154	-0.364	0.6952
– molluscs : N : GS	-0.124	-0.32, 0.09	4940	0.2296	-0.542	0.5815
– rabbits : GS : N	0.156	-0.05, 0.36	4940	0.1304	0.079	1.0827
GS : N : P	-0.183	-0.4, 0.01	4940	0.0781	-0.503	0.6045
– insects – molluscs – rabbits	0.238	-1.57, 2.1	5148	0.7814	0.386	1.4711

Table SI.6 Step-wise model reduction with AIC and p-values of each assessed parameter in testing community-weighted mean genome size. This table shows interactions and variables that have been removed because they do not have a significant influence on the dependent variable. The beginning, most complex model included all four-way interactions between herbivore exclusion treatments and the fertilizers N and P.

Parameter	change in AIC	p-value
K	2	0.8217
Mg	2	0.8801
slope	2.1	0.3934
herbicide	1.1	0.2313
– rabbits – molluscs – insects : N	2	0.9931
– rabbits – molluscs – insects : P	1.9	0.7825
– rabbits – molluscs: N : P	1.3	0.3900
– rabbits – insects : N : P	1.6	0.5406
– molluscs – insects : N : P	1.8	0.6457
– rabbits – molluscs – insects	1.4	0.4503
– rabbits – insects : N	0.9	0.2899
– rabbits – molluscs: P	2	0.9833
– rabbits – insects : P	2	0.9911
– molluscs – insects : P	0.6	0.2342
– rabbits : N : P	1.1	0.3468
– molluscs : N : P	0.1	0.1699
– insects : N : P	1.1	0.3400
– rabbits – insects	2	0.8222
– rabbits : P	-0.5	0.1124
– molluscs : P	-0.7	0.0992
– insects : P	1.9	0.7700
N : P	1.4	0.4489

Table SI.7 Effects of experimental treatment (herbivore exclusion and N and P_input) on community-weighted mean genome size (CWM GS). CWM GS (1C-value) on plots with all herbivores and no nutrient input (control plots) = 5.76 pg. The model shows a significant decrease in CWM GS with N fertilizer and rabbit and mollusc exclusion (= +N plots with insect grazing only) to 4.22 pg; and with N fertilizer and mollusc and insect exclusion (= +N plots with rabbit grazing only) to 3.03 pg. CI= 95% confidence intervals. CWM GS estimated using PGLS.

	LME					ANOVA		
	Coef.	CI	Std. error	t-value	p-value	CWM GS (pg)	F value	Pr(>F)
Intercept	5.759	5.02, 6.50	0.453	12.721	< 0.00001	5.76	na	na
- rabbits	1.106	0.26, 1.95	0.485	2.280	0.0538	6.86	27.265	0.0019
- molluscs	-1.013	-2.05, 0.02	0.636	-1.593	0.1465	4.75	6.971	0.0566
- insects	-0.285	-1.15, 0.58	0.534	-0.533	0.6174	5.47	0.474	0.5283
+ N	-1.868	-2.42, -1.32	0.281	-6.636	< 0.00001	3.89	184.67	< 0.00001
+ P	-0.312	-0.54, -0.08	0.117	-2.669	0.0078	5.45	7.125	0.0078
- rabbits - molluscs	0.679	-0.52, 1.88	0.690	0.985	0.3544	6.53	0.017	0.9007
- rabbits : N	1.097	0.46, 1.73	0.325	3.374	0.0008	6.09	4.588	0.0327
- molluscs: N	-0.355	-1.14, 0.43	0.401	-0.885	0.3765	2.52	0.989	0.3206
- molluscs - insects	-0.330	-1.55, 0.90	0.760	-0.434	0.6826	4.13	0.286	0.6207
- insects : N	-0.315	-0.95, 0.32	0.325	-0.969	0.3329	3.29	2.927	0.0877
- rab - mol: N	-1.190	-2.10, -0.28	0.468	-2.540	0.0114	4.22	6.453	0.0114
- mol - ins : N	1.433	0.52, 2.35	0.469	3.054	0.0024	3.03	9.327	0.0024

Table SI.8 Means and standard deviations of community-weighted mean genome size (CWM GS) (1C-value, pg), total biomass, number of species and phylogenetic diversity of plots, shown for each of eight herbivore treatments and \pm nitrogen (N) treatment (n = the number of plots included in the analysis of each treatment, out of a total of 556 plots). Control = no treatment (i.e. all herbivores present and no fertilizer input).

Herbivore treatment	N	CWM GS (pg)	Total biomass (g)	Number of species	Phylogenetic diversity	n
Control (all herbivores)	–	5.59 \pm 1.43	33.14 \pm 15.17	9 \pm 2	57.03 \pm 12.56	36
– insects	–	5.33 \pm 1.68	59.79 \pm 23.72	8 \pm 3	50.42 \pm 14.12	36
– molluscs	–	4.77 \pm 1.53	29.79 \pm 10.34	9 \pm 3	54.03 \pm 16.64	36
– rabbits	–	6.72 \pm 1.34	43.29 \pm 11.23	7 \pm 2	41.36 \pm 13.05	36
– molluscs – rabbits	–	6.19 \pm 1.42	65.79 \pm 15.97	8 \pm 3	49.5 \pm 15.23	36
– insects – molluscs	–	3.78 \pm 1.33	48.72 \pm 18.59	11 \pm 2	65.37 \pm 11	30
– insects – rabbits	–	6.42 \pm 1.2	52.93 \pm 17.3	6 \pm 2	37.86 \pm 9.75	36
No herbivores	–	5.92 \pm 1.26	57.58 \pm 15.11	9 \pm 2	55.76 \pm 12.86	33
All herbivores present	+	3.6 \pm 1.96	46.4 \pm 24.39	8 \pm 3	50.97 \pm 15.13	36
– insects	+	3.27 \pm 1.7	78.23 \pm 31.46	7 \pm 3	43.61 \pm 14.87	36
– molluscs	+	2.44 \pm 1.09	43.61 \pm 16.7	8 \pm 3	48.72 \pm 14.54	36
– rabbits	+	6.07 \pm 2.33	76.92 \pm 21.44	5 \pm 2	28.42 \pm 11.19	36
– molluscs – rabbits	+	3.98 \pm 1.46	83.85 \pm 23.41	6 \pm 2	36.25 \pm 13.73	36
– insects – molluscs	+	2.82 \pm 1.27	74.63 \pm 31.13	8 \pm 2	54.07 \pm 12.08	29
– insects – rabbits	+	5.21 \pm 1.81	90.3 \pm 30.56	5 \pm 1	27.36 \pm 8.96	36
no herbivores	+	4.62 \pm 1.28	100.83 \pm 26.24	6 \pm 2	33.47 \pm 10.54	32

Table SI.9 Most reduced linear mixed effect model output showing the effects of experimental treatments on three alternative estimations of community-weighted mean genome size (CWM GS). Prior to model reduction, CWM GS was estimated for each plot: **A)** CWM GS was estimated without accounting for phylogenetic non-independence (i.e. it is simply weighted by species biomass). Similar to the community-weighted means estimated in the main text, GS was log₁₀-transformed prior to the estimation, and back-transformed for ease of interpretation and comparison; **B)** CWM GS estimated with PGLS as described in the text, however GS was *not* log-transformed; **C)** CWM GS was log₁₀-transformed and estimated with PGLS, however the lambda parameter was allowed to be estimated (rather than fixed at 1 which assumes a Brownian motion of evolution). The last column contains the estimated CWM GS (1C-value in pg) of plots under each of the treatments on left. This table shows that LME output is very similar between different estimations, the key results remaining unchanged. See also Fig. SI.4.

A) CWM GS, without phylogeny						
	Coef.	CI	Std. error	t-value	p-value	Mean GS (pg)
Intercept	5.669	5.02, 6.31	0.387	14.644	< 0.0001	5.67
– rabbits	0.962	0.21, 1.71	0.447	2.154	0.0650	6.63
– molluscs	-0.751	-1.65, 0.15	0.541	-1.388	0.1987	4.92
– insects	-1.530	-1.94, -1.12	0.213	-7.196	< 0.0001	4.14
+ N	-0.287	-1.02, 0.45	0.443	-0.648	0.5477	5.38
+ P	-0.197	-0.44, 0.05	0.126	-1.568	0.1175	5.47
– rabbits – molluscs	0.442	-0.60, 1.48	0.622	0.711	0.4997	6.32
– rabbits : N	1.050	0.57, 1.53	0.246	4.277	< 0.0001	7.39
– molluscs: N	-0.470	-1.06, 0.12	0.303	-1.554	0.1208	4.16
– molluscs – insects	-0.429	-1.47, 0.62	0.629	-0.683	0.5266	2.96
– insects : N	-0.365	-0.84, 0.11	0.246	-1.487	0.1376	3.49
– rabbits : P	-0.390	-0.73, -0.05	0.177	-2.208	0.0277	6.04
– rab – mol : N	-1.021	-1.71, -0.33	0.354	-2.885	0.0041	5.59
– mol – ins : N	1.188	0.50, 1.88	0.355	3.352	0.0009	3.02

B) CWM GS estimated with PGLS, GS untransformed						
	Coef.	CI	Std. error	t-value	p-value	Mean GS (pg)
Intercept	6.419	5.73, 7.11	0.410	15.676	< 0.0001	6.42
– rabbits	0.810	0.03, 1.59	0.469	1.728	0.1087	7.23
– molluscs	-0.996	-1.96, -0.03	0.575	-1.732	0.1078	5.42
– insects	-0.307	-2.55, -1.53	0.469	-0.654	0.5248	6.11
+ N	-2.040	-1.09, 0.48	0.263	-7.76	< 0.0001	4.38
+ P	-0.352	-0.56, -0.14	0.109	-3.217	0.0014	6.07
– rabbits – molluscs	0.643	-0.48, 1.76	0.667	0.964	0.3530	6.88
– rabbits : N	1.297	0.70, 1.89	0.304	4.272	< 0.0001	6.49
– molluscs: N	-0.517	-1.25, 0.21	0.374	-1.381	0.1680	2.87
– molluscs – insects	-0.250	-1.37, 0.87	0.667	-0.375	0.7136	4.87
– insects : N	-0.284	-0.88, 0.31	0.304	-0.935	0.3503	3.79
– rab – mol : N	-0.980	-1.83, -0.13	0.438	-2.241	0.0255	4.64
– mol – ins : N	1.309	0.46, 2.16	0.438	2.988	0.0029	3.33

Table SI.9 continued

c)	CWM GS estimated with PGLS, lambda parameter optimised						Mean GS (μg)
	Coef.	CI	Std. error	t-value	p-value		
Intercept	5.598	4.86, 6.33	0.445	12.569	< 0.0001	5.60	
– rabbits	0.890	0.04, 1.74	0.493	1.806	0.1081	6.49	
– molluscs	-0.981	-1.99, 0.03	0.615	-1.593	0.1462	4.62	
– insects	-0.405	-1.24, 0.43	0.511	-0.793	0.4658	5.19	
+ N	-1.300	-1.85, -0.75	0.282	-4.613	< 0.0001	4.30	
+ P	0.041	-0.32, 0.40	0.186	0.222	0.8246	5.64	
– rabbits – molluscs	0.806	-0.36, 1.97	0.682	1.182	0.2737	6.31	
– rabbits : N	1.086	0.51, 1.66	0.298	3.650	0.0003	6.27	
– molluscs: N	-0.252	-0.97, 0.47	0.370	-0.680	0.4965	3.07	
– molluscs – insects	-0.223	-1.40, 0.96	0.725	-0.308	0.7713	3.99	
– insects : N	-0.193	-0.77, 0.39	0.298	-0.649	0.5168	3.70	
– rabbits : P	-0.497	-0.92, -0.08	0.215	-2.315	0.0210	6.03	
+ N + P	-0.507	-0.92, -0.09	0.215	-2.355	0.0189	3.83	
– rab – molluscs: N	-1.484	-2.32, -0.65	0.430	-3.450	0.0006	4.36	
– mol – insects: N	1.065	0.23, 1.91	0.431	2.470	0.0139	5.18	

Table SI.10 Linear model output for each for the community properties in the path analysis model in Fig. 2 (\pm rabbits). CW = community-weighted.

Community-weighted mean genome size (1C-value, pg)					
	Estimate	Std. Error	95% CI	t value	Pr(> t)
Intercept	5.124	1.020	3.24, 7.06	5.021	0.0188
– rabbits	1.339	1.277	-1.08, 3.76	1.049	0.4664
N	-1.694	0.404	-2.47, -0.92	-4.195	0.0001
P	-0.100	0.412	-0.9, 0.7	-0.243	0.8088
K	-0.137	0.247	-0.61, 0.34	-0.552	0.5820
– grass (herbicide)	0.741	0.511	-0.24, 1.72	1.45	0.1973
– forbs (herbicide)	1.025	0.511	0.04, 2.01	2.007	0.0916
– rabbits : N	1.344	0.466	0.44, 2.24	2.881	0.0047
– rabbits : P	-0.433	0.466	-1.33, 0.47	-0.928	0.3550
N : P	-0.594	0.466	-1.49, 0.31	-1.274	0.2049

Community-weighted mean C-strategy					
	Estimate	Std. Error	95% CI	t value	Pr(> t)
Intercept	0.365	0.069	0.21, 0.52	5.311	0.0693
– rabbits	0.042	0.032	-0.02, 0.1	1.308	0.2034
N	0.089	0.028	0.04, 0.15	3.22	0.0016
P	0.011	0.027	-0.04, 0.06	0.43	0.6681
K	0.030	0.016	0, 0.06	1.84	0.0681
– grass (herbicide)	-0.063	0.028	-0.11, -0.01	-2.207	0.0648
– forbs (herbicide)	-0.002	0.029	-0.05, 0.05	-0.057	0.9560
CW mean genome size	0.025	0.005	0.02, 0.04	4.674	< 0.0001
– rabbits : N	-0.023	0.031	-0.09, 0.03	-0.743	0.4590
– rabbits : P	0.001	0.030	-0.06, 0.06	0.025	0.9799
N : P	0.009	0.030	-0.05, 0.07	0.286	0.7756

Total biomass (g/m²)					
	Estimate	Std. Error	95% CI	t value	Pr(> t)
Intercept	14.037	10.186	-5.47, 34.86	1.378	0.2688
– rabbits	10.212	8.795	-9.14, 29.35	1.161	0.4068
N	8.978	4.172	0.98, 16.92	2.152	0.0332
P	5.427	4.219	-2.63, 13.48	1.287	0.2005
K	3.614	2.551	-1.28, 8.47	1.417	0.1589
– grass (herbicide)	3.522	2.972	-2.23, 9.34	1.185	0.2381
– forbs (herbicide)	0.293	2.937	-5.45, 6	0.1	0.9207
CW mean C-strategy	37.640	12.292	14.06, 62.69	3.062	0.0027
– rabbits : N	19.974	4.773	10.85, 29.08	4.185	0.0001
– rabbits : P	-5.440	4.773	-14.55, 3.68	-1.14	0.2565
N : P	5.272	4.772	-3.83, 14.39	1.105	0.2713

Table SI.10 continued

Phylogenetic diversity

	Estimate	Std. Error	95% CI	t value	Pr(> t)
Intercept	65.081	4.336	56.05, 72	15.009	< 0.0001
– rabbits	-15.477	3.656	-22.16, -9.54	-4.233	0.0003
N	-3.143	3.240	-9.63, 2.88	-0.97	0.3339
P	-0.101	3.284	-6.5, 6.21	-0.031	0.9756
K	-2.653	1.981	-6.61, 1.05	-1.339	0.1830
– grass (herbicide)	7.998	3.042	3.77, 12.66	2.629	0.0398
– forbs (herbicide)	-5.469	3.027	-10, -1.18	-1.807	0.1223
CW mean C-strategy	-21.259	8.239	-30.36, -2.28	-2.58	0.0139
– rabbits : N	-6.666	3.715	-13.9, 0.47	-1.794	0.0752
– rabbits : P	2.620	3.715	-4.52, 9.85	0.705	0.4821
N : P	-3.964	3.714	-11.12, 3.25	-1.067	0.2879

Table SI.11 Linear mixed effect model output for each of the community properties in the path analysis in Fig. 3 (\pm insects \pm molluscs). CW= community-weighted, CI = confidence intervals.

Community-weighted mean genome size (1C-value, pg)					
	Estimate	Std. Error	95% CI	t-value	p-value
Intercept	6.943	0.777	5.70, 8.18	8.934	0.0001
– insects	-0.771	1.075	-2.48, 0.94	-0.718	0.5035
– molluscs	-0.938	1.075	-2.65, 0.77	-0.873	0.4208
N	-0.483	0.446	-1.34, 0.37	-1.084	0.2796
P	-0.697	0.449	-1.56, 0.16	-1.55	0.1223
K	-0.043	0.169	-0.37, 0.28	-0.253	0.8004
– grass (herbicide)	0.158	0.264	-0.36, 0.67	0.6	0.5583
– forbs (herbicide)	0.274	0.268	-0.24, 0.80	1.021	0.3244
– insects – molluscs	1.073	1.528	-1.37, 3.50	0.702	0.5117
– insects : N	-0.014	0.630	-1.22, 1.19	-0.023	0.9819
– molluscs : N	-1.448	0.630	-2.66, -0.24	-2.298	0.0224
– insects : P	0.938	0.630	-0.27, 2.15	1.489	0.1379
– molluscs : P	0.824	0.630	-0.38, 2.03	1.308	0.1922
N : P	-0.329	0.630	-1.54, 0.88	-0.523	0.6018
– insects – molluscs : N	0.155	0.907	-1.58, 1.90	0.171	0.8646
– insects – molluscs : P	-1.957	0.905	-3.69, -0.22	-2.162	0.0316
– insects : N : P	-1.096	0.891	-2.8, 0.61	-1.229	0.2202
– molluscs : N : P	-0.224	0.891	-1.93, 1.48	-0.252	0.8016
– insects – molluscs : N : P	2.571	1.284	0.10, 5.02	2.002	0.0464

Community-weighted mean C-strategy					
	Estimate	Std. Error	95% CI	t-value	p-value
Intercept	0.406	0.057	0.31, 0.50	7.106	0.0006
– insects	-0.041	0.079	-0.17, 0.08	-0.524	0.6223
– molluscs	-0.031	0.079	-0.16, 0.09	-0.395	0.7089
N	0.088	0.031	0.03, 0.15	2.827	0.0051
P	0.040	0.032	-0.02, 0.10	1.252	0.2118
K	0.014	0.012	-0.01, 0.04	1.207	0.2287
CW mean genome size	0.025	0.004	0.02, 0.03	5.77	0.0000
– grass (herbicide)	-0.088	0.017	-0.12, -0.05	-5.082	0.0002
– forbs (herbicide)	0.000	0.018	-0.04, 0.03	-0.022	0.9829
– insects – molluscs	0.048	0.112	-0.13, 0.23	0.427	0.6862
– insects : N	-0.041	0.044	-0.12, 0.04	-0.92	0.3583
– molluscs : N	-0.009	0.045	-0.09, 0.08	-0.213	0.8318
– insects : P	-0.043	0.044	-0.13, 0.04	-0.975	0.3306
– molluscs : P	-0.049	0.044	-0.13, 0.04	-1.1	0.2724
N : P	-0.036	0.044	-0.12, 0.05	-0.814	0.4165
– insects – molluscs : N	0.125	0.063	0, 0.25	1.966	0.0504
– insects – molluscs : P	0.105	0.064	-0.02, 0.23	1.647	0.1008
– insects : N : P	0.187	0.063	0.07, 0.31	2.987	0.0031
– molluscs : N : P	0.171	0.062	0.05, 0.29	2.745	0.0065
– insects – molluscs : N : P	-0.283	0.090	-0.46, -0.11	-3.129	0.0020

Table SI.11 continued ± insects ± molluscs

Total biomass (g/m²)					
	Estimate	Std. Error	95% CI	t-value	p-value
Intercept	14.923	6.949	2.46, 28.01	2.147	0.0339
– insects	9.157	6.737	-3.23, 21.48	1.359	0.1769
– molluscs	26.591	6.744	14.2, 38.93	3.943	0.0001
N	25.264	6.270	13.38, 37.33	4.029	0.0001
P	-2.659	6.265	-14.59, 9.35	-0.424	0.6717
K	3.595	2.348	-0.87, 8.09	1.531	0.1270
CW mean C-strategy	65.655	11.309	43.63, 85.97	5.806	0.0000
CW mean genome size	-1.570	0.866	-3.17, 0.08	-1.814	0.0709
– grass (herbicide)	4.147	3.638	-2.26, 10.49	1.14	0.2678
– forbs (herbicide)	0.023	3.560	-6.20, 6.24	0.006	0.9950
– insects – molluscs	-23.151	9.747	-41.01, -5.25	-2.375	0.0192
– insects : N	1.492	8.741	-15.25, 18.15	0.171	0.8646
– molluscs : N	-23.804	8.804	-40.62, -7.00	-2.704	0.0073
– insects : P	9.271	8.778	-7.54, 25.99	1.056	0.2919
– molluscs : P	-0.815	8.774	-17.63, 15.9	-0.093	0.9261
N : P	7.572	8.741	-9.16, 24.24	0.866	0.3872
– insects – molluscs : N	24.815	12.639	0.82, 49.08	1.963	0.0507
– insects – molluscs : P	-2.824	12.695	-26.92, 21.53	-0.222	0.8241
– insects : N : P	-2.374	12.567	-26.18, 21.82	-0.189	0.8503
– molluscs : N : P	6.648	12.499	-17.05, 30.69	0.532	0.5953
– insects – molluscs : N : P	-2.636	18.211	-37.68, 31.78	-0.145	0.8850
Phylogenetic diversity					
	Estimate	Std. Error	95% CI	t-value	p-value
Intercept	45.363	4.942	37.37, 53.36	9.179	0.0000
– insects	-5.111	6.654	-15.79, 5.57	-0.768	0.4711
– molluscs	7.722	6.654	-2.96, 18.40	1.16	0.2893
N	-12.111	3.353	-18.54, -5.68	-3.612	0.0004
P	1.584	3.380	-4.90, 8.06	0.469	0.6397
K	-3.585	1.272	-6.02, -1.15	-2.818	0.0052
– grass (herbicide)	-0.740	2.503	-5.61, 4.13	-0.295	0.7721
– forbs (herbicide)	-8.265	2.535	-13.18, -3.33	-3.261	0.0056
– insects – molluscs	8.768	9.490	-6.53, 23.99	0.924	0.3896
– insects : N	3.389	4.742	-5.70, 12.48	0.715	0.4755
– molluscs : N	3.167	4.742	-5.92, 12.26	0.668	0.5049
– insects : P	3.222	4.742	-5.87, 12.31	0.68	0.4975
– molluscs : P	0.833	4.742	-8.26, 9.92	0.176	0.8606
N : P	-1.667	4.742	-10.76, 7.42	-0.351	0.7255
– insects – molluscs : N	-10.602	6.827	-23.65, 2.52	-1.553	0.1217
– insects – molluscs : P	1.010	6.817	-12.00, 14.13	0.148	0.8823
– insects : N : P	-1.889	6.706	-14.74, 10.97	-0.282	0.7784
– molluscs : N : P	-6.944	6.706	-19.80, 5.91	-1.036	0.3014
– insects – molluscs : N : P	-1.797	9.674	-20.43, 16.65	-0.186	0.8528

Table SI.12 Change in species mean percent of above-ground biomass, measured as dry weight, per herbivore exclusion treatment, relative to the control plots (plots without herbivore exclusion i.e. plots with rabbits, molluscs, and insects), and by N treatment. Mean percentages were rounded, thus plants with very low biomass may equal to 0.

	- rab	- rab	- rab	- rab	- mol	- mol	- ins	- ins	- ins	- ins	- all	- all	- ins	- ins
	no N	+ N	- mol	- mol	no N	+ N	- rab	- rab	no N	+ N	no N	+ N	- mol	- mol
<i>Achillea millefolium</i>	0.4	0	-0.3	-0.2	0.1	-0.2	-0.6	-0.6	-0.6	-0.5	1.9	3.9	-0.6	-0.5
<i>Agrostis capillaris</i>	-7.1	-10.6	-8.5	-12.1	0.9	-1.7	-8.1	-11.8	-8.7	-10.6	-5	-11.9	-6.5	-10.3
<i>Anthoxanthum odoratum</i>	-0.4	-0.8	-0.7	-0.8	0.4	-0.3	-0.6	-0.7	-0.7	-0.8	-0.2	-0.8	-0.8	-0.7
<i>Aphanes microcarpa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Arrhenatherum elatius</i>	17.9	26.6	-3.3	-5.9	-3.3	-8.1	-0.5	0.6	6.3	2.1	0.7	-3.7	-1.6	-3
<i>Carex hirta</i>	0	0	0	0	0	0	0	0	0.3	0	0	0	0	0
<i>Carex muricata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Centaurea nigra</i>	0.3	0	0	0	0	1.1	0	0	0	0	0	0	0	0
<i>Cerastium fontanum</i>	0	-0.2	0	-0.2	0	-0.1	0	-0.2	0	-0.2	0	-0.2	0	-0.2
<i>Chenopodium album</i>	-0.3	0	-0.3	0.1	-0.3	0	-0.3	0	-0.3	0	-0.3	0	-0.3	0
<i>Cirsium arvense</i>	-1.1	-4.3	-0.2	-3.5	-1	-2.2	-1.1	-4	3.7	2.6	-0.8	-4	2.8	-0.6
<i>Crepis capillaris</i>	0	-0.1	1.3	-0.1	0.2	0	0.2	-0.1	0	0	0.2	-0.1	0	-0.1
<i>Dactylis glomerata</i>	0.3	0.3	7	7.4	0	0	3	2.7	2.8	0.8	2.3	1.8	0.1	0
<i>Epilobium ciliatum</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Festuca rubra</i>	-5.4	5.2	8.4	2.5	-6	-13.2	7.6	15	-4.5	-8.3	-7.6	-7.6	-24.3	-21.1
<i>Galium aparine</i>	0	0	0	0	0	0	0	0	0.2	0.1	0	0	0	0
<i>Galium saxatile</i>	0	0	0	0	0	0	0.7	2.3	0	0	0	0	0	0
<i>Galium verum</i>	0	0	0	0	0	0	0	0	0	0	0	0.7	0	0
<i>Heracleum sphondylium</i>	0	0	0	0.6	0	0	0	0	0	0	0	0	0	0
<i>Hieracium pilosella</i>	0	0	-0.1	0	-0.1	0	-0.1	0	-0.1	0	-0.1	0	-0.1	0
<i>Holcus lanatus</i>	-3.8	-13	-3.8	-4.1	6	18.2	-1.9	-15.3	1	3.3	1.1	-11.5	7.1	7.3
<i>Holcus mollis</i>	1.1	1.8	1.9	17.6	1.5	6.4	3	14.4	-0.5	7.4	6.7	34.5	8.1	15.9

Table SI.12 continued

	- rab	- rab	- rab	- rab	- mol	- mol	- ins	- ins	- ins	- ins	- all	- all	- ins	- ins
	no N	+ N	- mol	- mol	no N	+ N	- rab	- rab	no N	+ N	no N	+ N	- mol	- mol
<i>Hypochaeris radicata</i>	0	0	0	0	0	0	0	0	0	0.1	0	0	0	0
<i>Jacobaea vulgaris</i>	-1.3	-0.6	-1.3	-0.6	-1	-0.5	-1.3	-0.6	1.5	4	-1.3	-0.6	12.1	13
<i>Juncus effusus</i>	0	0	0	0	0	0	0	0	0.1	0	0	0	0	0
<i>Lotus corniculatus</i>	0.5	0	-0.1	0	-0.1	0	-0.1	0	-0.1	0	0.1	0	0.4	0
<i>Luzula campestris</i>	-0.7	-0.1	-1.3	-0.1	-0.5	-0.1	-1.3	-0.1	-1.3	-0.1	-1.3	-0.1	-1	-0.1
<i>Malva moschata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Medicago lupulina</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Phleum pratense</i> subsp. <i>bertolonii</i>	0	0	0.1	1.7	3.1	1.3	0	0.5	0.1	0.3	1.2	1.7	0.7	0
<i>Plantago lanceolata</i>	-0.4	-0.2	0.3	-0.1	-0.4	-0.2	0.2	-0.2	-0.4	-0.2	-0.1	-0.2	-0.3	-0.2
<i>Poa annua</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Poa pratensis</i>	0.7	0.1	-0.1	1.4	-0.1	0.4	0	2.1	0	0.9	0.8	1.7	0.6	0.3
<i>Poa trivialis</i>	0	0	0	0	0	0	0	0	0	0.2	0	0	0	0
<i>Potentilla erecta</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Quercus cerris</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Quercus robur</i>	0.3	0	0.1	0	0	0	0	0	0	0	0	0	0	0
<i>Ranunculus acris</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Ranunculus bulbosus</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Ranunculus repens</i>	-0.3	-0.1	-0.2	0	-0.2	0	-0.3	-0.1	-0.1	0.1	-0.3	-0.1	0.2	0.8
<i>Rubus fruticosus</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Rumex acetosa</i>	-0.1	-0.3	-0.1	-0.2	0.5	1.1	0	-0.1	0.9	1.3	0.2	-0.3	0.5	0.2
<i>Rumex acetosella</i>	-0.2	-2.2	-0.2	-2.2	-0.2	-1.1	-0.2	-2.2	-0.2	-2	-0.2	-2.2	0.5	-1.5
<i>Sambucus nigra</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Scorzoneroideis autumnalis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Stellaria graminea</i>	0.1	-0.2	0	0.1	0.4	0.2	-0.1	-0.3	0	-0.1	0.1	-0.2	0.2	1.1

Table SI.12 continued

	- rab	- rab	- rab - mol	- rab - mol	- mol	- mol	- ins - rab	- ins - rab	- ins	- ins	- all	- all	- ins - mol	- ins - mol
	no N	+ N	no N	+ N	no N	+ N	no N	+ N	no N	+ N	no N	+ N	no N	+ N
<i>Stellaria media</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Taraxacum officinale</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Trifolium dubium</i>	0	0	0	0	0	0	0	0	0	0	0.1	0	0	0
<i>Trifolium pratense</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Trifolium repens</i>	0	0	0	0	0.3	0	0	0	0	0	0.4	0	0.1	0
<i>Trisetum flavescens</i>	0	0	0	0	0	0	0.1	0	0	0	0	0	0	0
<i>Veronica arvensis</i>	0	0	0	0	0	0	0.1	0	0	0	0	0	0	0
<i>Veronica chamaedrys</i>	-0.9	-1.2	0.3	-1.1	-0.6	-0.9	-0.8	-1.2	0.6	0	-0.3	-0.8	2	-0.1
<i>Vicia sativa</i> subsp. <i>nigra</i>	0.3	0.1	1.1	0	0.1	0	2.6	0	0	0	1.7	0	0.2	0
<i>Viola riviniana</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0

References

1. Faith DP. 1992. Conservation evaluation and phylogenetic diversity. *Biol Conserv* **61**, 1-10.
2. Pellicer J, Leitch IJ. 2014. The application of flow cytometry for estimating genome size and ploidy level in plants. In *Molecular Plant Taxonomy* (ed. Besse P), pp. 279-307, Humana Press.
3. Bennett MD, Leitch IJ. 2012. Plant DNA C-values database (release 6.0, Dec. 2012). <http://data.kew.org/cvalues/>.
4. The Angiosperm Phylogeny Group. 2016. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. *Bot J Linn Soc* **181**, 1-20.
5. Canty AJ, Ripley B. 2014. Boot: Bootstrap R (S-Plus) functions. R package version 1.3-20.
6. von Hardenberg A, Gonzalez-Voyer A. 2013. Disentangling evolutionary cause-effect relationships with phylogenetic confirmatory path analysis. *Evolution* **67**, 378-387.
7. Gonzalez-Voyer A, von Hardenberg A. 2014. An introduction to phylogenetic path analysis. In *Modern phylogenetic comparative methods and their application in evolutionary Biology* (ed. Garamszegi LZ), pp. 201-229. Berlin Heidelberg, Springer-Verlag.