# **Materials Horizons**





# Photoelectrochemical response of carbon dots (CDs) derived from chitosan and their use in electrochemical imaging

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De-Wen Zhang,<sup>ab</sup> Nikolaos Papaioannou,<sup>cd</sup> Naomi Michelle David,<sup>a</sup> Hui Luo,<sup>a</sup> Hui Gao,<sup>a</sup> Liviu Cristian Tanase,<sup>e</sup> Thibault Degousée,<sup>a</sup> Paolo Samorì,<sup>f</sup> Andrei Sapelkin,<sup>cd</sup> Oliver Fenwick,<sup>a</sup> Maria-Magdalena Titirici,<sup>a</sup>\* Steffi Krause<sup>a</sup>\*

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We report a direct photoelectrochemical response from low cost carbon dots (CDs) prepared from chitosan via a solvothermal method. The carbon dots were covalently linked to an indium tin oxide (ITO) surface through a self-assembled silane monolayer. We attribute the photocurrent of the ITO-silane-CD surface to a photogenerated electron-transfer process by CDs under illumination with a wavelength of 420 nm to 450 nm. The selfassembled monolayer of CDs was used for ac-photocurrent imaging of the surface with micron scale lateral resolution. This discovery opens up new applications for CDs as biocompatible, light-addressable electrochemical sensors in bioanalytical and bioimaging applications.

Carbon dots (CDs) have recently generated excitement as a new generation of metal free fluorescent materials and semiconductor-like photocatalysts.<sup>1, 2</sup> CDs have been widely applied to enhance the photocatalytic activity of known semiconductor photocatalysts in photoelectric conversion,<sup>3</sup> pollutant photodegradation,<sup>4</sup> water splitting,<sup>5, 6</sup> CO<sub>2</sub> conversion,<sup>7, 8</sup> and organic synthesis.<sup>9</sup> CDs have played important roles in photoreactions by light absorption, electron-hole pair generation, photoexcited charge separation and migration, which showed both electron donor and acceptor abilities under illumination. However, the photoinduced electron transfer process of CDs alone is still unclear. Graphene quantum dots (GQDs),<sup>10</sup> which belong to the CD family, showed photoelectric conversion properties as

a semiconductor.<sup>11-13</sup> N- and O- doping in carbon sp<sup>2</sup> clusters of GQDs were shown to be responsible for n- and p-type domains, respectively.<sup>14</sup> To date the direct photoelectric conversion using pure carbon nanodots under illumination has reported. Herein, not been we report the photoelectrochemical behaviour of CDs prepared by simple bottom-up solvothermal approach from chitosan as carbon precursor containing nitrogen functionality and their use in electrochemical imaging sensors. Self-assembled monolayers of quantum dots (e.g. CdS) have been used in light-addressable electrodes for macroscopic electrochemical sensing.<sup>15</sup> The great advantage of CDs over these conventional quantum dots is that they are cheap, non-poisonous and biocompatible making them ideal for live-cell imaging applications with high spatial resolution. Many mature amperometric or impedimetric biosensor technologies for the detection of metabolites and proteins have traditionally been using carbon as electrode material and can, therefore, be easily adapted for use in the CD based imaging sensor introduced in this work.

CDs were prepared by a one-step solvothermal carbonization of chitosan dispersed in ethanol (4% w/v).<sup>16</sup> Figure 1(A) shows the transmission electron microscopy (TEM) image of CDs with spherical morphologies and an average diameter of around 4 nm. CDs were observed to be amorphous carbon particles without any lattices from the high-resolution transmission electron microscopy (HRTEM) image. The X-ray diffraction (XRD) pattern of CDs (Figure S1) displayed a broad peak centred at 21°, which corresponded to a set of sp2 carbons-graphitic carbons with stacking faults known as turbostratic carbons<sup>17</sup> and indicated a significant level of topological disorder in CDs.<sup>18</sup> The absence of other peaks in the XRD spectrum confirmed the amorphous nature of CDs. The CD suspension in ethanol was brown, transparent and clear under daylight and exhibited strong blue luminescence under 365 nm UV excitation (Figure 1 (B)). The UV/Vis absorption spectrum of the CD solution in ethanol shows an absorption peak at 263 nm (Figure S2 (A)). In agreement with results reported previously, excitationdependent emission was observed (Figure S3), where emission

<sup>&</sup>lt;sup>a.</sup> School of Engineering and Materials Science, Queen Mary University of London, Mile End Road, London, E1 4NS, UK. Email: <u>m.m.titirici@gmul.ac.uk</u>, <u>s.krause@gmul.ac.uk</u>

<sup>&</sup>lt;sup>b.</sup> Institute of Materials, China Academy of Engineering Physics, Jiangyou, 621908, Sichuan, P.R. China

<sup>&</sup>lt;sup>c.</sup> School of Physics and Astronomy, Queen Mary University of London, 327 Mile End Road, London, E1 4NS, UK

<sup>&</sup>lt;sup>d.</sup> Materials Research Institute, Queen Mary University of London, Mile End Road, E1 4NS, London, UK

<sup>&</sup>lt;sup>e.</sup> National Institute of Materials Physics, Atomistilor 405A, 077125, Magurele-Ilfov, Romania

<sup>&</sup>lt;sup>f.</sup> University of Strasbourg, CNRS, ISIS UMR 7006, 8 allée Gaspard Monge, F-67000 Strasbourg, France

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### COMMUNICATION

peaks shifted to higher wavelengths as the excitation wavelength increased from 340 nm to 500 nm.<sup>16</sup> The fluorescent quantum yield of the CD solution was calculated to be 5.7%. Figure S4 shows the X-ray photoelectron spectroscopy (XPS) C 1s and N 1s spectra of the CDs, which demonstrates the presence of amine groups in the CDs derived from chitosan. As shown in Figure S5, the Fourier transform infrared (FTIR) spectrum showed the presence of functional groups of CDs, which also demonstrated the presence of amine groups in the CDs.

To investigate the photoelectric conversion behaviour, CDs were covalently linked to an indium tin oxide (ITO) surface through a 3-glycidoxypropyldimethoxymethylsilane (G-silane) monolayer based on the epoxy-amine reaction as shown in Figure 2 (A).<sup>19</sup> An epoxy terminated monolayer on ITO was formed by silanization. Then, CDs from chitosan containing alkyl amine groups were covalently attached to the epoxy groups. After binding of CDs, the colourless ITO-silane surface became evenly light brown. As shown in Figure S6, the absorbance of the ITO-silane-CDs surface at wavelengths < 400 nm in the UV/Vis absorption spectrum increased compared to the ITO-silane surface, which indicated the binding of CDs to the ITO-silane surface. The X-ray photoelectron spectroscopy (XPS) survey spectrum in Figure 2 (B) confirms the linking of Gsilane and CDs by the appearance of a nitrogen peak at 399.5 eV (Figure S7 (D)) assigned to the amine in the CDs from chitosan and the increase in the carbon signal. The Si 2p peak at 101.2 eV (Figure S7 (C)) was assigned to the organic Si in silane and corresponded to G-silane bonded to the ITO surface. The C 1s peak at 285.6 eV on the ITO-silane-CDs surface (Figure S7 (B)) was assigned to C-N bonds, and its absence on the ITO-silane surface (Figure S7 (A)) further confirmed the binding of CDs onto the ITO-silane surface. Atomic Force Microscopy (AFM) was used to image the ITOsilane and ITO-silane-CD surface. The height variation changed from 1.0 nm of ITO-silane to 2.0 ~ 4.5 nm of ITO-silane-CDs (Figure S8), which proved that CDs were homogenously coated onto the ITO-silane surface. The ITO-silane-CD surface was used for the investigation of photoelectrochemical responses.

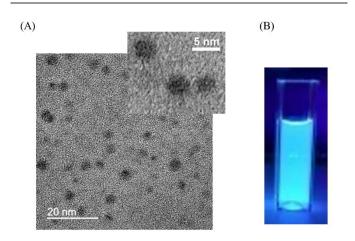
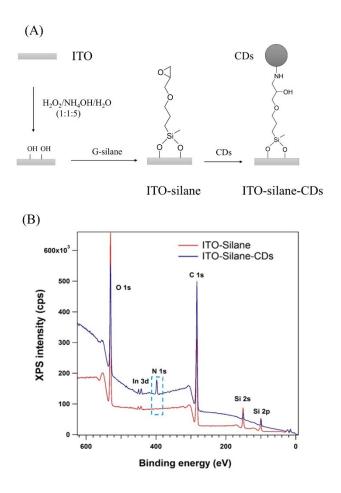


Figure 1. (A) TEM image of the CDs; (B) photograph of CDs in ethanol with 365 nm excitation.



**Figure 2.** (A) Scheme of CDs binding onto ITO via reaction with 3-glycidoxypropyldimethoxymethylsilane; (B) XPS full scan spectrum of the ITO-silane and ITO-silane-CDs.

Photoelectric response measurements of ITO-silane-CDs were carried out by linear sweep voltammetry (LSV) from -0.6 to 1 V with a 5 mV/s scan rate in 10 mM pH 7.4 phosphate buffer with 0.1 M KCl in an electrochemical cell consisting of a platinum electrode and an Ag/AgCl electrode as the counter and reference electrodes, respectively. To prevent the interference of the ITO substrate under UV illumination, an adjustable diode laser (wavelength 400 nm - 450 nm, max 500 mW) with a UV filter that blocked the light of wavelengths lower than 420 nm was used as the chopped back side illumination source. The CDs could be excited by the 420 - 450 nm wavelength light (also see the fluorescence spectra of CDs in Figure S3). The light intensity was measured as 10 mW/cm<sup>2</sup> with a calibrated power meter. As shown in Figure 3(A), the ITO-silane-CDs surface showed an obvious photocurrent at anodic potentials with chopped illumination compared to the background dark current. The photocurrent under illumination appeared from 0.25 V and increased with the applied potential. Figure 3 (B) shows the LSV curves of ITO, ITO-silane, and ITO-silane-CDs surfaces from 0.2 V to 1 V under chopped illumination. The dark current of the ITO-silane-CD surface

slowly increased at anodic potentials compared to the ITO and ITO-silane surfaces, suggesting that the CDs increased the conductivity of the surface. The photocurrent response of the ITO-silane-CD surface was significantly higher than that of ITO and ITO-silane surfaces. At 0.96 V, photocurrents (I<sub>illumination</sub> - $I_{dark}$ ) of ITO, ITO-silane, and ITO-silane-CD surfaces were 0.6 ± 0.1  $\mu$ A/cm<sup>2</sup>, 1.2 ± 0.1  $\mu$ A/cm<sup>2</sup>, and 2.8 ± 0.6  $\mu$ A/cm<sup>2</sup>, respectively. The results confirmed the direct photoelectrochemical response of CDs derived from chitosan under 420 - 450 nm laser illumination. Figure S9 shows the stability of the photocurrent of the ITO-silane-CD surface at 1.0 V with chopped illumination in 10 minutes. After 10 cycles, the photocurrent decayed to 68.4%. In this work, the photocurrent  $(I_{illumination} - I_{dark})$  of the ITO-silane-CD surface was 0.35 ± 0.02  $\mu$ A/cm<sup>2</sup> at 0.25 V, which is comparable to the reported performance of a covalently bound CdS quantum dots monolayer.15

We demonstrate here that solvothermally synthesized nitrogen doped CDs from chitosan have an n-type semiconductor-like photoelectrochemical behaviour. To better understand the mechanism for the photoelectrochemical response of the CDs, we used photoelectron spectroscopy in air to measure the ionization potential of CDs in powder form (Figure S10). We measured a value for the ionization potential of  $5.52 \pm 0.01 \text{ eV}$ , corresponding to the energy of highest occupied electronic state of CDs. This is significantly larger than the ionization potential of other carbon systems such as graphite  $(\sim 4.45 \text{ eV})^{20}$  and graphene  $(4.5 - 5.1 \text{ eV})^{21-23}$ . One reason for this might be nitrogen doping coming from the amine groups in the chitosan starting material. Nitrogen doping is commonly reported to reduce the work function of carbon systems,<sup>24-26</sup> but it can increase the work function of graphitic systems if the incorporation of nitrogen atoms is at pyrrolic or pyridinic sites.<sup>27</sup> We note that our XPS N1s spectra do indicate both pyrridinic and pyrrolic species in the CDs. Previous studies of nitrogen-doped CDs have reported large ionization potentials (6.16 eV),  $^{\rm 28}$  as in our case. We also note that partial oxidation of the CDs may also increase the ionization potential as is the case for graphene oxide.<sup>29</sup> The Tauc plot (Figure S2 (B)) derived from the UV/Vis absorption spectrum of CDs indicates an energy of gap (Egap) of 3.7 eV, therefore putting the energy of the lowest unoccupied states at 1.8 eV. The excitation-dependent photoluminescence (PL) of CDs (Figure S3) indicates individual energy states (due to surface defects, functional group, impurities, etc.) of CDs,<sup>30</sup> which allows a very broad excitation band extending up to 500 nm (2.48 eV). The possibility of excitations significantly below the energy gap (3.7 eV) indicates a significant number of intergap states as shown in Figure 4. There is a large energetic driving force for electron transfer to the ITO (Fermi energy,  $E_f \approx 4.6 \text{ eV}$ ). At anodic potentials, the holes in CDs accept electrons from an electron donor (D) in electrolyte solution, which is OH<sup>-</sup> in this case. The formation of D<sup>+</sup> from D using CDs as photocatalysts under illumination generates the photocurrent. The photocurrents of CDs related to their quantum yields were investigated. CDs with a higher quantum yields (QY) of 10.4% were synthesised by a one-step solvothermal carbonization of chitosan and ethylenediamine to increase the N content in the CDs. The ITO-silane-CDs with 10.4% QY showed higher photocurrents than those with 5.7% QY under the same binding and measurement conditions (Figure S11). This indicates that N doping plays an important role in the PL/QY and photocurrent response of CDs.

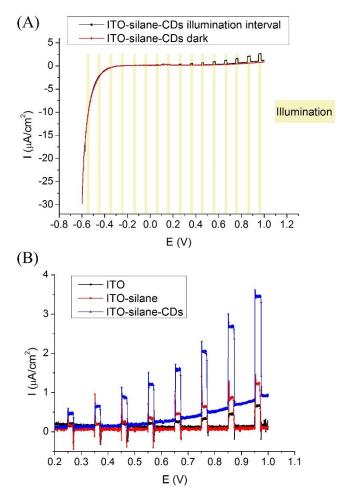


Figure 3. (A) LSV curves of ITO-silane-CDs in the dark and with chopped illumination (420 nm-450 nm, 10 mW/cm<sup>2</sup>, a UV blocking filter was used to block the light at wavelengths < 420 nm); (B) LSV curves of ITO, ITO-silane, and ITO-silane-CDs with chopped illumination.

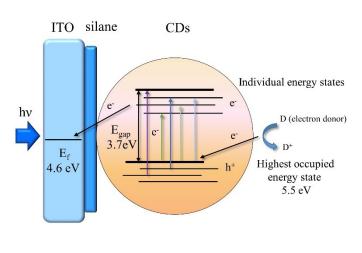


Figure 4. The mechanism of photoelectric conversion of CDs.

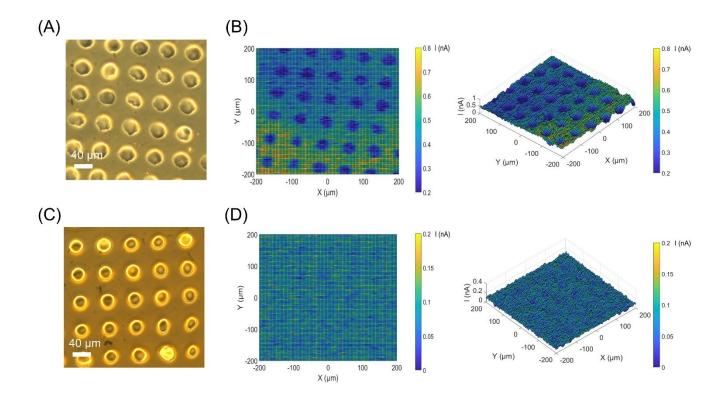


Figure 5. (A) Optical micrograph of a PMMA dot array on ITO-silane-CD surface by microcontact printing; (B) Corresponding ac-photocurrent image of (A) measured at 1.8 V; (C) Optical micrograph of a PMMA dot array on ITO-silane surface; (D) Corresponding ac-photocurrent image of (C) measured at 1.8 V.

The ITO-silane-CD surface was applied to ac-photocurrent imaging using the same experimental techniques usually applied to light-addressable potentiometric sensors (LAPS) and scanning photo-induced impedance microscopy (SPIM), which measure the local photocurrent by illuminating a semiconductor substrate with a focused, modulated laser beam. 2D images were obtained by moving the sensor in the XY plane with respect to the laser beam while recording photocurrent, providing local electrochemical information such as impedance,<sup>31</sup> ion concentrations,<sup>32</sup> and surface charge<sup>33</sup>. An array of poly (methyl methacrylate) (PMMA) dots was deposited onto the surface by PDMS stamp using microcontact printing ( $\mu$ CP). Figure 5 (A) shows the circular islands of PMMA with a diameter of 40  $\mu$ m and 30  $\mu$ m gaps. Figure 5 (B) shows the corresponding ac-photocurrent image measured at a bias voltage of 1.8 V and a modulation frequency of 10 Hz with a focused 420 - 450 nm laser beam. The polymer dot array is clearly visible as a reduction in photocurrent because of the high impedance of PMMA. An ITO-silane surface without CDs, but patterned with PMMA dots was imaged with ac-photocurrent imaging as the control experiment. The control image (Figure 5(D)) did not show any contrast, proving that contrast in the ac-photocurrent images comes directly from the photoelectrochemical response of CDs. To demonstrate the uniformity of the ITO-silane-CD surface, a small area ( $40 \times 40 \mu$ m) ac-photocurrent scan with a step size of 0.4  $\mu$ m at 1.8 V was performed. The ac-photocurrent image (Figure S12) proved that the surface was uniform and CDs were homogenously coated. As shown in Figure S13, the ac-photocurrent remained stable for 15 min at a bias voltage of 1.8 V and a modulation of 10 Hz with unfocused 420-450 nm laser illumination.

To demonstrate the feasibility of using the CD modified ITO surface for bioanalytical and bioimaging applications, a multilayer of yeast cells was immobilized on the ITO-silane-CD surface by low gelling temperature agarose gel (Figure S14 (A)). Figure S14 (B) shows the corresponding ac-photocurrent image at 1.5 V and 10 Hz. The yeast-agarose gel is visible as a reduction in the photocurrent because of the impedance and negative surface charge of yeast cells in pH 7.4 PBS solution<sup>34</sup>. The control image of pure agarose gel (Figure S14 (D)) did not show any photocurrent contrast, proving that signal came from the surface attached yeast cells.

# Conclusions

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In conclusion, CDs from chitosan were covalently linked to an ITO surface and showed a direct photoelectrochemical response. The photocurrent generated by CDs and the mechanism have been studied. The photocurrent was attributed to the enhanced electron-transfer process catalyzed by CDs under illumination. A homogeneous monolayer of CDs was successfully used for ac-photocurrent imaging. This opens up new applications for CDs as biocompatible, lightaddressable electrochemical sensors in bioanalytical and bioimaging applications. As for well-established photocurrent imaging techniques such as LAPS and SPIM, a CD modified surface could be used for electrochemical imaging in the cellsurface attachment area, which is not accessible to other electrochemical or electrophysiological techniques. However, in contrast to LAPS and SPIM, which are limited to the measurement of ion concentrations<sup>35</sup> or impedance<sup>34</sup>, a CD modified surface could image a much broader range of parameters as carbon has been a common electrode material for the amperometric detection of metabolites and lends itself to standard immobilization techniques for enzymes and other electroactive molecules. The lateral resolution of acphotocurrent imaging with a CD modified surface would only be limited by the quality of the focus of the light as there would be no lateral diffusion of photogenerated charge carriers as in the bulk semiconductors traditionally employed for LAPS and SPIM<sup>36</sup>.

# Author contributions

D. –W. Z., M. –M. T., and S. K. designed the experimental program and coordinated the project. N. P., H. L., H. G., and A. S. produced the CDs samples and performed characterizations of CDs. D. –W. Z. and N. M. D. performed the binding of CDs onto ITO surface and photocurrent measurements. L. C. T. performed XPS measurements and analysis. T.D. performed AFM measurements and analysis. P. S. and O. F. performed PESA measurements and provided the mechanism. D. –W. Z. performed the  $\mu$ CP and ac photocurrent imaging. All authors contributed to the interpretation of the results. D. –W. Z., O. F., M. –M. T., and S. K. wrote the paper.

# **Conflicts of interest**

There are no conflicts of interest to declare.

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# References

- 1 H. Yu, R. Shi, Y. Zhao, G. I. N. Waterhouse, L.-Z. Wu, C.-H. Tung and T. Zhang, *Adv. Mater.*, 2016, **28**, 9454-9477.
- K. A. S. Fernando, S. Sahu, Y. Liu, W. K. Lewis, E. A. Guliants, A. Jafariyan, P. Wang, C. E. Bunker and Y.-P. Sun, ACS Appl. Mater. Interfaces, 2015, 7, 8363-8376.
- 3 J. Bian, C. Huang, L. Wang, T. Hung, W. A. Daoud and R. Zhang, ACS Appl. Mater. Interfaces, 2014, **6**, 4883-4890.
- 4 H. Yu, H. Zhang, H. Huang, Y. Liu, H. Li, H. Ming and Z. Kang, New J. Chem., 2012, 36, 1031-1035.
- 5 X. Zhang, F. Wang, H. Huang, H. Li, X. Han, Y. Liu and Z. Kang, Nanoscale, 2013, 5, 2274-2278.
- 6 H. Yu, Y. Zhao, C. Zhou, L. Shang, Y. Peng, Y. Cao, L.-Z. Wu, C.-H. Tung and T. Zhang, *Journal of Materials Chemistry A*, 2014, **2**, 3344-3351.
- K. D. Yang, Y. Ha, U. Sim, J. An, C. W. Lee, K. Jin, Y. Kim, J.
  Park, J. S. Hong, J. H. Lee, H.-E. Lee, H.-Y. Jeong, H. Kim and K.
  T. Nam, *Adv. Funct. Mater.*, 2016, 26, 233-242.
- 8 L. Cao, S. Sahu, P. Anilkumar, C. E. Bunker, J. Xu, K. A. S. Fernando, P. Wang, E. A. Guliants, K. N. Tackett and Y.-P. Sun, J. Am. Chem. Soc., 2011, **133**, 4754-4757.
- 9 R. Liu, H. Huang, H. Li, Y. Liu, J. Zhong, Y. Li, S. Zhang and Z. Kang, ACS Catalysis, 2014, 4, 328-336.
- A. Ciesielski, S. Haar, A. Aliprandi, M. El Garah, G. Tregnago, G. F. Cotella, M. El Gemayel, F. Richard, H. Sun, F. Cacialli, F. Bonaccorso and P. Samorì, ACS Nano, 2016, **10**, 10768-10777.
- 11 J. Shen, Y. Zhu, X. Yang, J. Zong, J. Zhang and C. Li, New J. Chem., 2012, **36**, 97-101.
- 12 S. K. Lai, C. M. Luk, L. Tang, K. S. Teng and S. P. Lau, Nanoscale, 2015, 7, 5338-5343.
- 13 C.-B. Ma, Z.-T. Zhu, H.-X. Wang, X. Huang, X. Zhang, X. Qi, H.-L. Zhang, Y. Zhu, X. Deng, Y. Peng, Y. Han and H. Zhang, *Nanoscale*, 2015, **7**, 10162-10169.
- 14 T.-F. Yeh, C.-Y. Teng, S.-J. Chen and H. Teng, *Adv. Mater.*, 2014, **26**, 3297-3303.
- 15 W. Khalid, M. El Helou, T. Murböck, Z. Yue, J.-M. Montenegro, K. Schubert, G. Göbel, F. Lisdat, G. Witte and W. J. Parak, ACS Nano, 2011, 5, 9870-9876.
- 16 J. Briscoe, A. Marinovic, M. Sevilla, S. Dunn and M. Titirici, Angew. Chem. Int. Ed., 2015, 54, 4463-4468.
- 17 Z. Q. Li, C. J. Lu, Z. P. Xia, Y. Zhou and Z. Luo, *Carbon*, 2007, 45, 1686-1695.
- 18 S. Zhu, Q. Meng, L. Wang, J. Zhang, Y. Song, H. Jin, K. Zhang, H. Sun, H. Wang and B. Yang, *Angew. Chem. Int. Ed.*, 2013, 52, 3953-3957.
- 19 M. Barreiros dos Santos, S. Azevedo, J. P. Agusil, B. Prieto-Simón, C. Sporer, E. Torrents, A. Juárez, V. Teixeira and J. Samitier, *Bioelectrochemistry*, 2015, **101**, 146-152.
- 20 W. N. Hansen and G. J. Hansen, Surf. Sci., 2001, 481, 172-184.
- 21 S. M. Song, J. K. Park, O. J. Sul and B. J. Cho, *Nano Lett.*, 2012, 12, 3887-3892.
- 22 Y.-J. Yu, Y. Zhao, S. Ryu, L. E. Brus, K. S. Kim and P. Kim, *Nano Lett.*, 2009, **9**, 3430-3434.
- 23 G. Giovannetti, P. A. Khomyakov, G. Brocks, V. M. Karpan, J. van den Brink and P. J. Kelly, *Phys. Rev. Lett.*, 2008, **101**, 026803.
- 24 D. H. Lee, J. A. Lee, W. J. Lee and S. O. Kim, *Small*, 2011, **7**, 95-100.
- 25 J. O. Hwang, J. S. Park, D. S. Choi, J. Y. Kim, S. H. Lee, K. E. Lee, Y.-H. Kim, M. H. Song, S. Yoo and S. O. Kim, *ACS Nano*, 2012, **6**, 159-167.
- 26 D. G. Kvashnin, P. B. Sorokin, J. W. Brüning and L. A. Chernozatonskii, *Appl. Phys. Lett.*, 2013, **102**, 183112.
- 27 K. Akada, T.-o. Terasawa, G. Imamura, S. Obata and K. Saiki, *Appl. Phys. Lett.*, 2014, **104**, 131602.

- 28 Y. R. Park, H. Y. Jeong, Y. S. Seo, W. K. Choi and Y. J. Hong, *Scientific Reports*, 2017, **7**, 13.
- 29 F. Zheng, W.-L. Xu, H.-D. Jin, X.-T. Hao and K. P. Ghiggino, *RSC Advances*, 2015, **5**, 89515-89520.
- 30 A. Cayuela, M. L. Soriano, C. Carrillo-Carrion and M. Valcarcel, *Chem. Commun.*, 2016, **52**, 1311-1326.
- 31 D.-W. Zhang, F. Wu and S. Krause, *Anal. Chem.*, 2017, **89**, 8129-8133.
- 32 T. Yoshinobu, K.-i. Miyamoto, T. Wagner and M. J. Schöning, Sensors Actuators B: Chem., 2015, **207**, Part B, 926-932.
- 33 J. Wang, Y. Zhou, M. Watkinson, J. Gautrot and S. Krause, Sensors Actuators B: Chem., 2015, 209, 230-236.
- 34 D.-W. Zhang, F. Wu, J. Wang, M. Watkinson and S. Krause, Electrochem. Commun., 2016, **72**, 41-45.
- 35 K. Miyamoto and T. Yoshinobu, *Sensors and Materials*, 2016, **28**, 1091-1104.
- 36 Y. Guo, K.-i. Miyamoto, T. Wagner, M. J. Schöning and T. Yoshinobu, *Sensors Actuators B: Chem.*, 2014, **204**, 659-665.