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8 **Novel blood pressure locus and gene discovery using GWAS and expression**
9 **datasets from blood and the kidney**

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12 The list of authors can be found at the end of the manuscript.

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1 **ABSTRACT**

2 Elevated blood pressure is a major risk factor for cardiovascular disease and has a substantial genetic
3 contribution. Genetic variation influencing blood pressure has the potential to identify new
4 pharmacological targets for the treatment of hypertension. To discover additional novel blood
5 pressure loci, we used 1000 Genomes Project-based imputation in 150,134 European ancestry
6 individuals and sought significant evidence for independent replication in a further 228,245
7 individuals. We report 6 new signals of association in or near *HSPB7*, *TNXB*, *LRP12*, *LOC283335*, *SEPT9*
8 and *AKT2*, and provide new replication evidence for a further 2 signals in *EBF2* and *NFKBIA*. Combining
9 large whole-blood gene expression resources totaling 12,607 individuals, we investigated all novel
10 and previously reported signals and identified 48 genes with evidence for involvement in BP
11 regulation that are significant in multiple resources. Three novel kidney-specific signals were also
12 detected. These robustly implicated genes may provide new leads for therapeutic innovation.
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1 INTRODUCTION

2 Genetic support for a drug target increases the likelihood of success in drug development (1) and
3 there is clear unmet need for novel therapeutic strategies to treat individuals with hypertension (2).
4 A number of large studies have described blood pressure (BP) variant identification by genome-wide
5 and targeted association approaches (3-19). Clinically the most predictive BP traits for cardiovascular
6 risk are systolic blood pressure (SBP) and diastolic blood pressure (DBP), reflecting roughly the peak
7 and trough of the BP curve, and pulse pressure (PP), the difference between SBP and DBP (20)
8 reflecting arterial stiffness. Using these three traits, we undertook a meta-analysis of 150,134
9 individuals from 54 genome-wide association studies of European ancestry with imputation based
10 on the 1000 Genomes Project Phase 1. To minimize reporting of false positive associations, we
11 sought stringent evidence for significant independent replication in a further 228,245 individuals.
12 We further followed up novel and previously reported association signals in multiple large gene
13 expression databases and the largest kidney tissue gene expression resource currently available.
14 Finally, we searched for enrichment of associated genes in biological pathways and gene sets and
15 identified whether any of the genes were known drug targets.

16 RESULTS

17 The stage 1 discovery meta-analysis included 150,134 individuals (**Online Methods; Supplementary**
18 **Tables 1-4, Supplementary Figures 1 and 2**) and 7,994,604 variants with minor allele frequency
19 (MAF) >1% and an effective sample size of at least 60% of the total (**Online Methods**). We identified
20 61 signals in the discovery analysis that were candidates for novel BP signals ($P < 10^{-6}$ for any trait;
21 **Supplementary Table 5**). To ensure robustness of signals, we examined BP associations in an
22 additional 228,245 individuals from 15 independent studies for replication, including 140,886
23 individuals from UK Biobank (19) (**Supplementary Table 6 and Online Methods**). We used the most
24 significant ("sentinel") SNP and trait for each locus in replication (61 tests). Twenty-two putatively
25 novel association signals were initially confirmed showing significant evidence of replication in the
26 independent stage-2 studies ($P < 8.2 \times 10^{-4}$, Bonferroni correction for 61 tests) and genome-wide
27 significance ($P < 5 \times 10^{-8}$) in a meta-analysis across all 378,376 individuals (**Online methods, Table 1,**
28 **Supplementary Table 7**). Of these, 14 were subsequently published in two other studies (18, 19)
29 which presented genome-wide significant associations with evidence of replication. A further two
30 were highlighted as putative novel signals in one of those studies (18) but had not been confirmed by
31 replication. In our study, we report the 6 remaining novel signals, and the 2 previously unconfirmed
32 signals (in *EBF2* and in *NFKBIA*), as novel signals. The 8 novel signals included 7 signals at 7
33 independent loci (**Supplementary Figure 3**) and one novel independent signal near a previously

1 reported hit near *TNXB* (**Online Methods, Supplementary Table 8, Supplementary Figure 4**). The
2 novel signals show both significant evidence of replication in the independent stage-2 studies ($P <$
3 8.2×10^{-4} , Bonferroni correction for 61 tests) and genome-wide significance ($P < 5 \times 10^{-8}$) in a meta-
4 analysis across all 378,376 individuals. The sentinel variants at all 8 signals were common (MAF>5%)
5 and the novel secondary signal at *TNXB* was in high linkage-disequilibrium ($r^2 > 0.8$) with a non-
6 synonymous SNP. With the exception of rs9710247, which was only significant for association with
7 DBP, all signals were significantly associated ($P < 0.006$, Bonferroni corrected for 8 tests) with all 3
8 traits (**Table 1 and Supplementary Table 9**).

9 We next sought to identify which genes might have expression levels that were associated with
10 genotypes of the BP-associated variants reported in this study and others. Strong evidence of an
11 association with expression of a specific gene may provide clues as to which gene(s) might be
12 functionally relevant to that signal. We took the 139 BP association signals reported prior to these
13 studies (18, 19), and 22 novel signals of association identified and confirmed in this study and two
14 contemporaneous studies (3-19, 21) (**Supplementary Table 10**), and searched for evidence of
15 association with gene expression in whole-blood (four studies, total $n=12,607$; **Online Methods**) and
16 in kidney tissue ($n=134$, the largest kidney eQTL resource currently available). Although of unclear
17 direct relevance to BP, whole-blood was studied due to the availability of large data sets enabling a
18 powerful assessment of expression patterns that are likely present across multiple cell and tissue
19 types. Kidney was chosen because of the many renal pathways that regulate BP and outstanding
20 questions about the relevance of kidney pathways to the genetic component of BP regulation in the
21 general population (3, 15). Expression quantitative trait loci (eQTL) signals were filtered by false
22 discovery rate (FDR<5%) and we examined *cis* (within 1Mb) associations only (**Online methods and**
23 **Supplementary Material**).

24 The four blood eQTL data sets were NESDA-NTR (22, 23), SABRe (15), the BIOS resource (24) and
25 GTEx(25) (**Online Methods and Supplementary Material**). The BIOS resource ($n=2,116$) has not
26 previously been utilized in the analysis of BP associations, findings from NESDA-NTR and SABRe have
27 been reported for a subset of the previously published signals (16, 17). For a total of 369 genes,
28 gene-expression was associated with the BP SNP in one or more of the 4 blood datasets at
29 experiment-wide significance (**Supplementary Table 11**). This included 14 genes for 6 of the 8 novel
30 signals. For 110 genes, we found eQTL evidence in 2 out of 4 datasets (**Figure 1**), including 4 genes
31 for 2 of the novel signals; *EIF4B* and *TNS2* for rs73099903 and *MAP3K10* and *PLD3* for rs9710247.
32 SNP rs73099903 was in strong linkage disequilibrium (LD $r^2 > 0.9$) with the SNP most strongly

1 associated with *TNS2* expression in the BIOS resource. *TNS2* encodes a tensin focal adhesion
2 molecule and may have a role in renal function (26).

3 For 48 genes, we found evidence in 3 out of the 4 resources (**Table 2**), suggesting robustness of the
4 SNP-gene expression correlation signal and highlighting those genes as potential candidates in
5 genetic BP regulation. Of the 48 genes, 28 have not previously been described in eQTL analyses using
6 BP associated SNPs and all were correlated with previously reported BP association signals.

7 In the kidney dataset (TransplantLines) (27), there was association of gene expression and genotype
8 for nine SNPs and 13 genes (**Table 2, Figure 1 and Supplementary Table 12**). Nine of the SNP-gene
9 expression associations were also observed in the whole-blood eQTL datasets, suggesting that those
10 signals may not be unique to the kidney. We report three signals that were unique to the kidney and
11 not previously reported (*C4orf34*, *HIP2* and *ASIC1*) and confirm a previously reported kidney eQTL
12 signal for an anti-sense RNA for *PSMD5* (15). The same SNP was also an eQTL for *PSMD5* itself in
13 both blood and kidney. *ASIC1* encodes the Acid Sensing Ion Channel Subunit 1 which may interact
14 (and be co-expressed) with ENaC subunits which mediate trans-epithelial Na transport in the kidney
15 (28). The comparatively small number of signals using kidney tissue (**Table 2 and Figure 1**) compared
16 to whole-blood could be due to the small sample size.

17 For genes implicated by eQTL information from whole-blood, we tested for enrichment of biological
18 pathways and gene ontologies (**Online Methods**). We noted enrichment of the 48 genes implicated
19 by 3 or 4 blood eQTL resources, **Table 2**, and a further 53 genes containing a non-synonymous
20 variant with $r^2 > 0.5$ with the top SNP (**Supplementary Table 13**), in pathways and ontology terms
21 related to actin and striated muscle (**Supplementary Tables 14 and 15, Online Methods**). Network
22 analysis using the same genes highlighted further GO terms relating to muscle function, particularly
23 cardiac muscle (**Online Methods, Supplementary Table 16**). We tested the overlap of 161 non-HLA
24 BP associated variants with DNase Hypersensitivity sites identified in the Roadmap and ENCODE cell
25 lines (**Online Methods**) and identified an overall enrichment in multiple cell and tissue types
26 including heart, kidney and smooth muscle (**Supplementary Figure 5**).

27 We next investigated these genes for potential suitable drug targets using the drug gene interaction
28 database (DGIdb) (29) and found 19 genes with known drug-gene interactions and 17 additional
29 genes with predicted druggability (**Supplementary Table 17**). These findings highlight potential
30 opportunities for novel therapeutic development and possible drug re-purposing, given that a large
31 number of the genes is already now targetable.

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2 **DISCUSSION**

3 Enhanced discovery of BP loci increases the potential targets for therapeutic advances. After major
4 advances in the number of BP loci known over the last years and months, we report 8 novel signals
5 that implicate 5 regions of the genome not previously connected to blood pressure regulation.

6 Six of the 8 novel signals we report had not previously been reported. Two signals (in *EBF2* and
7 *NFKBIA*) have been suggested previously but without evidence for replication (18). For these two
8 signals we present, for the first time, stringent evidence of replication, confirming their relevance to
9 blood pressure genetics.

10 The path from signal to genes is the essential next step towards realizing the therapeutic potential of
11 a genetic locus and understanding the mechanisms of BP regulation. We have used several large
12 eQTL resources as a first step to realize this objective. As expected, we observed that even across
13 eQTL studies of the same tissue, there is limited overlap in experiment-wide significant signals
14 suggesting either biologic variability, technology-specific differences in coverage of genes, or the
15 possibility of false positive results despite stringent within-experiment significance thresholds. By
16 selecting genes only significant in at least three resources, we identified 48 genes as candidates for
17 further study. These results are limited by the availability of large eQTL resources for whole-blood
18 only, which precludes well-powered comparisons across tissue types, particularly as the origin of
19 blood pressure control is unlikely to be located in the blood. Enrichment and pathway analyses using
20 these genes, and genes containing a correlated functional variant, highlight the potential relevance
21 of muscular tissue and pathways, compatible with a vascular and cardiac origin of BP genetics,
22 extending previous evidence (15). We identify a number of potential drug targets in the pathways
23 identified, providing, together with previous results, a possible avenue for development of
24 pharmacological interventions modulating blood pressure.

25 In summary, our study reports novel BP association signals and reports new candidate BP genes,
26 contributing to the transition from variants to genes to explain BP variation.

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2 **MATERIALS AND METHODS**

3 **Studies Stage 1**

4 Results from 54 independent European-ancestry studies, totaling 150,134 individuals, were included
5 in the Stage 1 meta-analysis: AGES (n=3215), ARIC (n=9402), ASPS (n=828), B58C (n=6458), BHS
6 (n=4492), CHS (n=3254), Cilento study (n=999), COLAUS (n=5404), COROGENE-CTRL (n=1878),
7 CROATIA-Vis (n=945), CROATIA-Split (n=494), CROATIA-Korcula (n=867), EGCUT (n=6395), EGCUT2
8 (n=1844), EPIC (n=2100), ERF (n=2617), Fenland (n=1357), FHS (n=8096), FINRISK-ctrl (n=861),
9 FINRISK CASE (n=839), FUSION (n=1045), GRAPHIC (n=1010), H2000-CTRL (n=1078), HealthABC
10 (n=1661), HTO (n=1000), INGI-CARL (n=456), INGI-FVG (n=746), INGI-VB (n=1775), IPM (n=300),
11 KORAS3 (n=1590), KORAS4 (n=3748), LBC1921 (n=376), LBC1936 (n=800), LOLIPOP-EW610 (n=927),
12 MESA (n=2678), MICROS (n=1148), MIGEN (n=1214), NESDA (n=2336), NSPHS (n=1005), NTR
13 (n=1490), PHASE (n=4535), PIVUS (n=945), PROCARDIS (n=1652), SHIP (n=4068), ULSAM (n=1114),
14 WGHS (n=23049), YFS (n=1987), ORCADES (n=1908), RS1 (n=5645), RS2 (n=2152), RS3 (n=3018),
15 TRAILS (n=1262), TRAILS-CC (n=282) and TWINGENE (n=9789). Full study names and general study
16 information is given in **Supplementary Table 1**.

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18 **Study-level genotyping and association testing**

19 Three quantitative BP traits were analyzed: SBP, DBP, and PP (difference between SBP and DBP).
20 Within each study, individuals known to be taking anti-hypertensive medication had 15 mmHg
21 added to their raw SBP value and 10 mmHg added to their raw DBP values (30). A summary of BP
22 phenotypes in each study is given in **Supplementary Table 2**. Association testing was undertaken
23 according to a central analysis plan that specified the use of sex, age, age², and body mass index
24 (BMI) as covariates and optional inclusion of additional covariates to account for population
25 stratification (**Supplementary Table 3**). Trait residuals were calculated for each trait using a normal
26 linear regression of the medication-adjusted trait values (mmHg) onto all covariates. The genotyping
27 array, pre-imputation quality control filters, imputation software and association testing software
28 used by each study are listed in **Supplementary Table 4**. All studies imputed to the 1000 Genomes
29 Project Phase 1 integrated release version 3 [March 2012] all ancestry reference panel (31). Imputed
30 genotype dosages were used to take into account uncertainty in the imputation. Association testing
31 was carried out using linear regression of the trait residuals onto genotype dosages under an
32 additive genetic model. Methods to account for relatedness within a study were used where
33 appropriate (**Supplementary Table 3**). Results for all variants (SNPs and INDELS) were then returned
34 to the central analysis group for further quality control checks and meta-analysis.

1 **Stage 1 meta-analysis**

2 Central quality control checks were undertaken across all results sets. This included checks to ensure
3 allele frequency consistency (across studies and with reference populations), checks of effect size
4 and standard error distributions (i.e. to highlight phenotype issues) and generation of quantile-
5 quantile (QQ) plots and genomic inflation factor lambdas to check for over- or under-inflation of test
6 statistics. Genomic control was applied (if $\lambda > 1$) at study-level. Variants with imputation quality
7 < 0.3 were excluded prior to meta-analysis. Inverse variance weighted meta-analysis was undertaken.
8 After meta-analysis, variants with a weighted minor allele frequency of less than 1% or $N_{\text{effective}}$
9 (product of study sample size and imputation quality summed across contributing studies) $< 60\%$
10 were then excluded and meta-analysis genomic control lambda calculated and used to adjust the
11 meta-analysis results.

12 **Selection of regions for follow-up**

13 For each trait, regions of association were selected by ranking variants by P value, recording the
14 variant with the lowest P value as a sentinel variant and then excluding all variants $\pm 500\text{kb}$ from
15 the sentinel and re-ranking the remaining variants. This was undertaken iteratively until all sentinel
16 variants representing 1Mb regions containing associations with $P < 10^{-6}$ had been identified. To
17 identify additional signals represented by secondary sentinel variants within 500kb of each of the
18 sentinel variants, GCTA (32) was used to run conditional analyses (conditioned on the first sentinel
19 variant) on each of the 1Mb regions using GWAS summary statistics and LD information from ARIC.
20 This was done both for putatively novel regions and for regions that had previously been reported. A
21 chi-squared test of heterogeneity of effect sizes across the 54 studies was run for each sentinel
22 variant and those with $P < 0.05$ for heterogeneity were excluded from further follow-up. Variants
23 with $P < 10^{-6}$ after conditioning on the sentinel SNP (novel or known) in the region and for which any
24 attenuation of the $-\log_{10} P$ value was less than 1.5 fold, were also taken forward for replication.

25 **Studies stage 2**

26 Data from 14 independent studies, totaling 87,360 individuals, and the first release of UK Biobank,
27 totaling 140,886 individuals, were combined to replicate the findings from stage 1 (i.e. totaling
28 228,245 individuals). Stage 2 study details, including full study names, are given in **Supplementary**
29 **Table 6** and included 3C-Dijon ($n=4061$), Airwave ($n=14023$), ASCOT-SC ($n=2462$), ASCOT-UK
30 ($n=3803$), BRIGHT ($n=1791$), GAPP ($n=1685$), GoDARTs ($n=7413$), GS:SFHS ($n=9749$), HCS ($n=2112$),
31 JUPITER ($n=8718$), LifeLines ($n=13376$), NEO ($n=5731$), TwinsUK ($n=4973$), UK Biobank-CMC
32 ($n=140,886$) and UKHLS ($n=7462$). Analysis was undertaken using the same methods as described for
33 Stage 1 studies. UK Biobank-CMC utilized a newer imputation reference panel than the other studies

1 and where a requested variant was not available, a proxy was used (next most significant P value
2 with linkage disequilibrium $r^2 > 0.6$ with original top variant). Results from all stage 2 studies were
3 meta-analyzed using inverse-variance weighted meta-analysis. Two of the variants, rs1048238 and
4 chr1:243458005:l, were not available in the largest study in Stage 2 (UK Biobank-CMC) and so proxy
5 variants were selected (based on P value and LD).

6 **Stage 1 + Stage 2 meta-analysis**

7 Following meta-analysis of stage 1 and stage 2 results, signals with a $P > 5 \times 10^{-8}$ were excluded. Of
8 the signals with a final $P < 5 \times 10^{-8}$, support for independent replication within the stage 2 studies only
9 was sought. Any signals which had $P < 5 \times 10^{-8}$ and evidence for independent replication in stage 2
10 alone, indicated by $P < 8.2 \times 10^{-4}$ (Bonferroni correction for 61 tests) were reported as novel signals of
11 association with BP. Any signals which were subsequently reported by other BP GWAS that were
12 accepted for publication during the time this analysis was ongoing, or signals for which
13 independence from another known signal could not be established, were removed from our list of
14 novel signals at this stage (**Supplementary Table 5**).

15 **Genotype and gene expression**

16 We searched for signals of association of genotype with gene expression for the 22 signals (including
17 8 novel) signals described in this study (**Supplementary Table 7**) and all signals reported prior to our
18 study (**Supplementary Table 10**) (3-17, 21) in 3 whole-blood data sets, 1 kidney data set and the
19 GTEx multiple tissue data resource, which included whole-blood (25). We selected cis signals of
20 association which were significant after controlling for 5% False Discovery Rate (FDR). The 3 whole-
21 blood eQTL data sets were the NHLBI Systems Approach to Biomarker Research in Cardiovascular
22 Disease initiative whole-blood eQTL resource (SABRe) (microarray, $n=5257$), NESDA-NTR
23 (microarray, $n=4896$), BIOS (RNAseq, $n=2116$). The whole-blood data from GTEx was based on data
24 from 338 samples. The kidney data set comprised 236 donor-kidney samples from 134 donors (27).
25 Full details of each data set can be found in the **Supplementary Material**.

26 **LD lookup**

27 The 1000 Genomes Project phase 3 release of variant calls was used (Feb. 20th, 2015), using 503
28 subjects of European ancestry(31). r^2 between the sentinel SNPs and all other bi-allelic SNPs within
29 the corresponding 2 Mb area was calculated using the Tabix and PLINK software package (v1.07) (33,
30 34). Annotation was performed using the ANNOVAR software package(35).

31 **Gene-based pathway analysis**

1 All genes identified in 3 or 4 of the whole-blood eQTL resources above (**Table 2**), and genes
2 containing a non-synonymous variant with $r^2 > 0.5$ with the sentinel variant (**Supplementary Table**
3 **13**), were tested for enrichment of biological pathways and gene ontology terms using
4 ConsensusPathDB (36) using a FDR < 5% cut-off. Enriched pathways and GO terms containing genes
5 only implicated by a single BP-associated variant were not reported.

6 **Network analysis**

7 To construct a functional association network, we combined two prioritized candidate gene sets into
8 a single query gene set as (i) genes mapping to the non-synonymous SNPs (nsSNPs) in high LD
9 ($r^2 > 0.5$) with the corresponding sentinel BP associated SNP, and (ii) genes with eQTL evidence from 3
10 or 4 of the blood eQTL resources. Three sentinel SNPs (rs185819, rs926552 and rs805303) mapping
11 to the HLA region on chromosome 6 were excluded from downstream analyses. The single query
12 gene set was then used as input for the functional network analysis(37). We used the Cytoscape (38)
13 software platform extended by the GeneMANIA(39) plugin (Data Version: 8/12/2014)(40). All the
14 genes in the composite network, either from the query or the resulting gene sets, were then used
15 for functional enrichment analysis against Gene Ontology terms (GO terms) (41) to identify the most
16 relevant GO terms using the same plugin (40).

17 **DNase1 Hypersensitivity overlap enrichment across tissue and cell-types**

18 The Functional element Overlap analysis of the Results of Genome Wide Association Study (GWAS)
19 Experiments (Forge tool v1.1)(42) was used to test for enrichment of overlap of BP SNPs in tissues
20 and cell lines from the Roadmap and ENCODE projects. All 164 SNPs were entered and 143 were
21 included in the analysis. SNPs from 9 commonly used GWAS arrays were used to select background
22 sets of SNPs for comparison and 10,000 background repetitions were run. A Z-score threshold of
23 ≥ 3.39 (estimated false positive rate of 0.5%) was used to declare significance.

24 **Drug-gene interactions**

25 Genes used for pathway and gene ontology enrichment analyses were further investigated for
26 potential druggable targets using the drug gene interaction database (DGIdb). The known drug-gene
27 interactions search parameters were set investigate all 15 databases in DGIdb and include all types
28 of interactions. The analysis performed for druggability prediction included all 9 databases
29 exclusively inspecting expert curated data only.

30

1 **NOTE:** Supplementary Information and Source Data files are available in the online version of the
2 paper.

3 **ACKNOWLEDGEMENTS**

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5 funding sources is provided in the **Supplementary Material**.

6 **CONFLICTS OF INTERESTS**

7 The authors declare competing financial interests (see corresponding section in the Supplementary
8 Material).

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1 **FIGURE LEGENDS**

2 **Figure 1: Overlap of eQTL evidence from four whole-blood and one kidney resource**

3 The figure indicates overlap of evidence for eQTLs from four whole-blood studies (SABRe, NESDA-
4 NTR, BIOS, and GTEx) and from one kidney resource (TransplantLines). Every colored line indicates
5 that this gene was analysis-wide significant in a given resource (see **Online Methods**). Only genes
6 identified by at least two resources are shown. The genes are sorted by genomic position on the y-
7 axis.

8

1 **FIGURES**

2 **Figure 1**

3

4

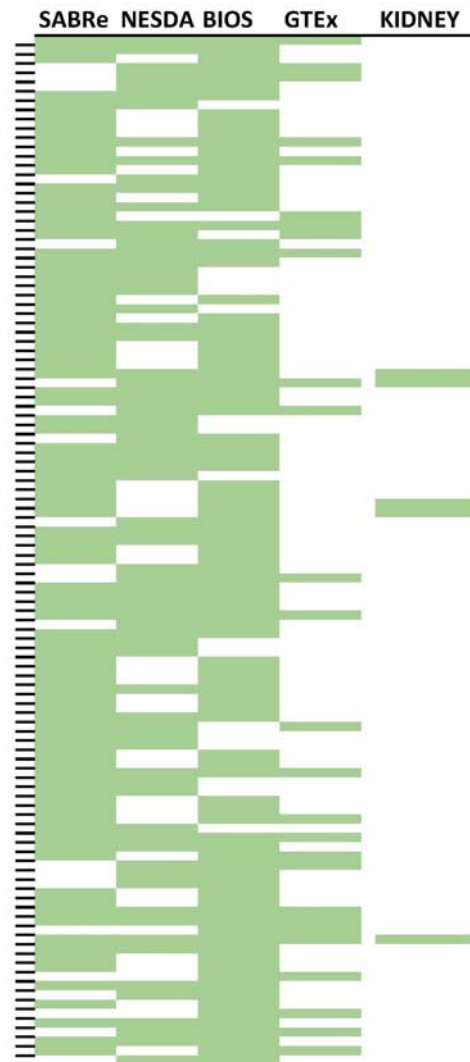


Table 1. Novel genome-wide significant signals of association

Results from stage 1 and stage 2, and the meta-analysis of stage 1 and stage 2, for all novel genome-wide significant signals of association. *P* values of association for all 3 traits from a meta-analysis of stages 1 and 2 are also presented. Genome-wide significant *P* values ($P < 5 \times 10^{-8}$) are in bold. Abbreviations: CAF: coded allele frequency se: standard error, Neff: effective sample size. #Novel signal at previously reported locus. ¹For intragenic variants the nearest genes are listed, all other variants are intronic unless indicated otherwise; ns= non-synonymous, s=synonymous, UTR= Untranslated Region. Results from proxy SNPs are indicated by (**proxy**); rs848309 was a proxy SNP for rs1048238 and rs10926988 was a proxy SNP for chr1:243458005:l.

Variant ID (noncoded/coded allele) chr:position, Nearest gene(s)(type ¹)	CAF	Results for most significant trait									Stage 1 + stage 2 meta-analysis P values for all traits			
		Stage 1		Stage 2			Stage 1+ stage 2				SBP	DBP	PP	
		Beta (se)	P value	Neff	Beta (se)	P value	Neff	Beta (se)	P value	Neff				
SBP														
rs1048238 (C/T) 1:16341649, <i>HSPB7</i> (3'UTR)	0.571	0.366 (0.074)	8.09E-07	140299	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
rs848309(proxy) (T/C) 1:16308447	0.567	0.347 (0.072)	1.70E-06	146755	0.347 (0.071)	9.10E-07	140462	0.347 (0.051)	7.07E-12	287217	7.07E-12	1.07E-10	5.48E-06	
#rs185819 (T/C) 6:32,050,067, <i>TNXB</i> (ns)	0.513	0.534 (0.073)	1.93E-13	142397	0.277 (0.053)	1.49E-07	221748	0.365 (0.043)	1.04E-17	364144	1.04E-17	2.24E-11	8.50E-15	
rs6557876 (C/T) 8:25,900,675, <i>EBF2</i>	0.252	-0.411 (0.084)	8.50E-07	143653	-0.350 (0.060)	5.66E-09	225803	-0.371 (0.049)	2.85E-14	369457	2.85E-14	2.50E-10	1.51E-08	
rs35783704 (G/A) 8:105,966,258, <i>LRP12/ZFPM2</i>	0.109	-0.609 (0.121)	4.96E-07	133924	-0.310 (0.089)	4.78E-04	215528	-0.414 (0.072)	7.08E-09	349452	7.08E-09	1.60E-06	2.92E-07	
rs73099903 (C/T) 12:53,440,779, <i>LOC283335</i>	0.074	0.768 (0.143)	8.05E-08	136064	0.396 (0.098)	5.32E-05	207253	0.515 (0.081)	1.95E-10	343318	1.95E-10	4.53E-06	5.46E-08	
rs8904 (G/A) 14:35,871,217, <i>NFKBIA</i> (3' UTR)	0.375	0.377 (0.076)	6.76E-07	140424	0.278 (0.054)	2.31E-07	224771	0.311 (0.044)	1.31E-12	365195	1.31E-12	1.13E-04	3.44E-12	
rs57927100 (C/G) 17:75,317,300, <i>SEPT9</i>	0.258	-0.489 (0.086)	1.10E-08	136624	-0.220 (0.061)	3.12E-04	210563	-0.310 (0.050)	4.04E-10	347188	4.04E-10	1.16E-10	1.81E-05	
DBP														
rs9710247 (A/G) 19:40,760,449, <i>AKT2</i>	0.447	0.252 (0.051)	8.11E-07	109695	0.129 (0.032)	5.76E-05	198332	0.164 (0.027)	1.61E-09	308028	3.82E-02	1.61E-09	5.03E-01	

Table 2: BP associated SNPs associated with expression of the same gene across 4 or 3 independent whole-blood eQTL resources and the kidney resource. Signals of association of SNP genotype and gene expression in other non-blood tissues in GTEx and in kidney are also indicated. Blood dataset order: (i) SABRe, (ii) NESDA-NTR, (iii) BIOS, (iv) GTEx (whole-blood). Top eQTL: Top GWAS SNP is top eQTL SNP (or in high LD, $r^2 > 0.9$, with top eQTL SNP) in at least one data set. eQTL signal previously reported: Genes for which eQTL signals have been previously reported for that sentinel SNP(15-17). For full list, see **Supplementary Table 12**.

Sentinel SNP	Chr	Position	Gene	Blood data sets	Top eQTL	Signal in other tissue(s) in GTEx	Signal in kidney	eQTL signal previously reported
Signal in 4 whole-blood eQTL resources								
rs17367504	1	11862778	<i>CLCN6</i>	YYYY		Y		Y
rs2169137	1	204497913	<i>MDM4</i>	YYYY	Y	Y		Y
rs10926988	1	243483279	<i>SDCCAG8</i>	YYYY		Y		
rs319690	3	47927484	<i>MAP4</i>	YYYY	Y	Y		Y
rs12521868	5	131784393	<i>SLC22A5</i>	YYYY		Y		
rs900145	11	13293905	<i>ARNTL</i>	YYYY		Y		Y
rs1060105	12	123806219	<i>CDK2AP1</i>	YYYY	Y	Y	Y	
rs1378942	15	75077367	<i>SCAMP2</i>	YYYY				
rs1126464	16	89704365	<i>CHMP1A</i>	YYYY		Y		Y
rs1126464	16	89704365	<i>FANCA</i>	YYYY				Y
rs12946454	17	43208121	<i>DKAKD</i>	YYYY		Y	Y	Y
Signal in 3 (out of 4) whole-blood eQTL resources								
rs17367504	1	11862778	<i>MTHFR</i>	YYYN		Y		Y
rs871524	1	38411445	<i>FHL3</i>	NYYY		Y		
rs871524	1	38411445	<i>SF3A3</i>	NYYY		Y		
rs4660293	1	40028180	<i>PABPC4</i>	YYYN	Y	Y		Y
rs6749447	2	169041386	<i>STK39</i>	YYYN	Y			
rs347591	3	11290122	<i>ATG7</i>	YYYN		Y		
rs319690	3	47927484	<i>ZNF589</i>	YYNY		Y		
rs12521868	5	131784393	<i>SLC22A4</i>	YYYN		Y		
rs1563788	6	43308363	<i>CRIP3</i>	YYYN	Y			Y
rs10943605	6	79655477	<i>PHIP</i>	YYYN	Y	Y		Y
rs4728142	7	128573967	<i>IRF5</i>	NYYY		Y	Y	Y
rs4728142	7	128573967	<i>TNPO3</i>	YYYN			Y	
rs2898290	8	11433909	<i>BLK</i>	YYYN		Y		
rs2898290	8	11433909	<i>FAM167A</i>	NYYY		Y		
rs2898290	8	11433909	<i>FDFT1</i>	YYYN		Y		
rs2071518	8	120435812	<i>NOV</i>	YYYN		Y		
rs76452347	9	35906471	<i>TPM2</i>	YYYN				

rs10760117	9	123586737	<i>MEGF9</i>	YYYN		Y		Y
rs4494250	10	96563757	<i>HELLS</i>	YYYN				Y
rs11191548	10	104846178	<i>NT5C2</i>	YYYN	Y			
rs661348	11	1905292	<i>TNNT3</i>	NYYY		Y		
rs2649044	11	9763969	<i>SBF2</i>	YYYN				
rs2649044	11	9763969	<i>SWAP70</i>	YYYN	Y	Y		?
rs7129220	11	10350538	<i>ADM</i>	YYYN				Y
rs7103648	11	47461783	<i>MYBPC3</i>	YYYN				
rs3741378	11	65408937	<i>CTSW</i>	YYYN				
rs7302981	12	50537815	<i>LIMA1</i>	YYYN				Y
rs7302981	12	50537815	<i>ATF1</i>	YNYN		Y		
rs1036477	15	48914926	<i>FBN1</i>	YNYN				
rs1378942	15	75077367	<i>CSK</i>	YYYN	Y	Y		Y
rs1378942	15	75077367	<i>MPI</i>	NYYY		Y		
rs1378942	15	75077367	<i>ULK3</i>	YNYN		Y		Y
rs12946454	17	43208121	<i>NMT1</i>	YYYN				Y
rs2304130	19	19789528	<i>GATAD2A</i>	YYYN				
rs867186	20	33764554	<i>EIF6</i>	NYYY		Y		
rs6095241	20	47308798	<i>PREX1</i>	YYYN				
rs9306160	21	45107562	<i>RRP1B</i>	YNYN	Y	Y		

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eQTL BIOS : Design of secondary analysis: R.Jansen Computation of secondary analysis: R.Jansen Study PI: R.Jansen

SABRe : Study phenotyping: Y.D., P.J.M., Q.T.N. Genotyping or analysis: R.Joehanes Design of secondary analysis: D.L. Study PI: D.L.

ICBP-Steering Committee

G.A., M.J.C., A.Chakravarti, D.I.C., G.B.E., P.E., T.F., M.R.J., A.D.J., M.Larson, D.L., A.P.M., P.B.M., C.N.C., P.O.R., W.P., B.M.P., K.R., A.V.S., H.Snieder, M.D.T., C.M.v.D., L.V.W., H.R.W.

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