

Inherited human group IVA cytosolic phospholipase A₂ deficiency abolishes platelet, endothelial, and leucocyte eicosanoid generation

Nicholas S. Kirkby,^{*,†} Daniel M. Reed,^{*} Matthew L. Edin,[‡] Francesca Rauzi,[†] Stefania Mataragka,^{*} Ivana Vojnovic,[†] David Bishop-Bailey,[§] Ginger L. Milne,[¶] Hilary Longhurst,^{||} Darryl C. Zeldin,[‡] Jane A. Mitchell,^{*,1} and Timothy D. Warner^{†,1,2}

^{*}National Heart and Lung Institute, Imperial College London, London, United Kingdom; [†]William Harvey Research Institute, Queen Mary University of London, London, United Kingdom; [‡]National Institutes of Health, National Institute of Environmental Health Sciences, Research Triangle Park, North Carolina, USA; [§]Department of Comparative Biomedical Sciences, Royal Veterinary College, London, United Kingdom; [¶]Department of Pharmacology and Department of Medicine, Vanderbilt University, Nashville, Tennessee, USA; and ^{||}Immunology Department, Barts Health and the London National Health Service Trust, London, United Kingdom

ABSTRACT Eicosanoids are important vascular regulators, but the phospholipase A₂ (PLA₂) isoforms supporting their production within the cardiovascular system are not fully understood. To address this, we have studied platelets, endothelial cells, and leukocytes from 2 siblings with a homozygous loss-of-function mutation in group IVA cytosolic phospholipase A₂ (cPLA₂α). Chromatography/mass spectrometry was used to determine levels of a broad range of eicosanoids produced by isolated vascular cells, and in plasma and urine. Eicosanoid release data were paired with studies of cellular function. Absence of cPLA₂α almost abolished eicosanoid synthesis in platelets (*e.g.*, thromboxane A₂, control 20.5 ± 1.4 ng/ml *vs.* patient 0.1 ng/ml) and leukocytes [*e.g.*, prostaglandin E₂ (PGE₂), control 21.9 ± 7.4 ng/ml *vs.* patient 1.9 ng/ml], and this was associated with impaired platelet activation and enhanced inflammatory responses. cPLA₂α-deficient endothelial cells showed reduced, but not absent, formation of prostaglandin I₂ (prostacyclin; control 956 ± 422 pg/ml *vs.* patient 196 pg/ml) and were primed for inflammation. In the urine, prostaglandin metabolites were selectively influenced by cPLA₂α deficiency. For example, prostacyclin metabolites were strongly reduced (18.4% of control) in patients lacking cPLA₂α, whereas PGE₂ metabolites (77.8% of control) were similar to healthy volunteer levels. These studies constitute a definitive account, demonstrating the fundamental role of cPLA₂α to eicosanoid formation and cellular responses within the human circulation.—Kirkby, N. S., Reed, D. M., Edin, M. L., Rauzi, F., Mataragka, S., Vojnovic, I., Bishop-Bailey, D., Milne, G. L., Longhurst, H., Zeldin, D. C., Mitchell, J. A., Warner, T. D. **Inherited**

human group IVA cytosolic phospholipase A₂ deficiency abolishes platelet, endothelial, and leucocyte eicosanoid generation. *FASEB J.* 29, 4568–4578 (2015). www.fasebj.org

Key Words: cardiovascular • thromboxane A₂ • prostacyclin • inflammation

In the cardiovascular system, eicosanoids have well-characterized roles in both normal function and a range of disease states (1, 2). For example, thromboxane A₂ (TXA₂), generated by platelets, drives thrombotic responses to particular stimuli (*e.g.*, collagen) and contributes to atherogenesis, whereas prostaglandin I₂ (prostacyclin), generated by endothelial cells, causes vasodilatation, inhibits platelet activation, and suppresses vascular inflammation. In leukocytes, eicosanoid formation [predominantly prostaglandin E₂ (PGE₂)] is induced by proinflammatory stimuli such as LPS that up-regulate cyclooxygenase (COX)-2 and other biosynthetic pathways (3) and so modulate the inflammatory response. In each case, although specific eicosanoid pathways such as TXA₂, PGE₂, and prostacyclin are well characterized, platelets, endothelial cells, and leukocytes synthesize substantial amounts of other arachidonic acid-derived mediators, the effect of which in combination remains poorly understood.

The arachidonic acid required for eicosanoid production is released from the sn-2 position of membrane

¹ These authors contributed equally to this work.

² Correspondence: The William Harvey Research Institute, Barts, and the London School of Medicine and Dentistry, Charterhouse Square, London EC1M 6BQ, United Kingdom. E-mail: t.d.warner@qmul.ac.uk

This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 International (CC BY 4.0) (<http://creativecommons.org/licenses/by/4.0/>) which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.
doi: 10.1096/fj.15-275065

This article includes supplemental data. Please visit <http://www.fasebj.org> to obtain this information.

Abbreviations: CD31, platelet endothelial cell adhesion molecule-1; COX, cyclooxygenase; cPLA₂, cytosolic phospholipase A₂; CXCL8, (C-X-C motif) ligand-8; DHA, docosahexaenoic acid; EET, epoxyeicosatrienoic acid; EPA, eicosapentaenoic acid; FLA, flagellin; FSL-1, bisacylated lipoprotein CGDPKHPKSF; HETE, hydroxyeicosatetraenoic acid; iPLA₂, calcium-independent

(continued on next page)

glycerophospholipids by the actions of phospholipase A₂ (PLA₂) enzymes. As reviewed (4, 5), >30 PLA₂ enzymes have been identified and currently classified into 6 broad families: secreted phospholipase A₂ (sPLA₂), Ca²⁺-dependent cytosolic phospholipase A₂ (cPLA₂), calcium-independent phospholipase A₂ (iPLA₂), platelet-activating factor acetylhydrolases, lysosomal PLA₂, and adipose-specific phospholipase. Of the known isoforms, group IVA cPLA₂ (also referred to as cPLA₂α), encoded by the PLA2G4A gene, is the most studied and has been characterized as a cytosolic enzyme, which upon Ca²⁺-dependent activation cleaves arachidonate-containing phospholipids to generate free intracellular arachidonic acid. This arachidonic acid is then used as a substrate by enzymes that synthesize the eicosanoid mediators, including COXs that produce prostanoids such as TXA₂ and prostacyclin, lipoxygenases (LOXs) that generate hydroxyeicosatetraenoic acids (*e.g.*, 12-HETE), and cytochrome P450 enzymes that generate epoxyeicosatrienoic acids (EETs) and HETEs (4, 5). cPLA₂α is widely expressed through the vasculature, in platelets, and in most types of blood leukocytes. Nonetheless, vascular and blood cells are known to express other PLA₂ enzymes, such as sPLA₂ enzymes including group II (platelets) and group V (endothelium) isoforms as well as iPLA₂ isoforms, which could also liberate arachidonic acid. For example, exogenous sPLA₂ has been demonstrated to activate platelets (6, 7) and endothelial cells (8). A role for endogenous sPLA₂ and iPLA₂ enzymes in eicosanoid generation by agonist-stimulated platelets (7, 9, 10), endothelial cells (11, 12), and leukocytes (13, 14) has also been described by several groups, calling into question the relative role of PLA₂ isoforms in eicosanoid generation and vascular protection. Indeed, the recent failure of the sPLA₂ inhibitor varespladib for the prevention of cardiovascular events in patients with acute coronary syndromes underlines our inadequate knowledge of the role of PLA₂ enzymes in vascular health and disease (15).

The key role of cPLA₂α in the generation of eicosanoid mediators is supported by data from cPLA₂α-knockout mice (9) and pharmacologic inhibitors (16, 17). Furthermore, we have recently reported 2 siblings with a homozygous mutation of the PLA2G4A gene that leads to a complete absence of cPLA₂α activity (18). Our work (18) and similar work from 2 other groups (19, 20) using tissue from patients with a heterozygous mutation of the PLA2G4A gene has shown that cPLA₂α regulates production of particular eicosanoids in platelets and in the urine. However, the relative role of cPLA₂α in endothelial cell and leukocyte eicosanoid function, as well as more broadly in platelets, has not thus far been addressed. By performing such studies, we have now definitively defined and compared the contribution of cPLA₂α with eicosanoid formation and inflammatory responses in leukocytes, platelets, and in endothelial cells. Our data

(continued from previous page)

phospholipase A₂; LOX, lipoxygenase; LTE₄, leukotriene E₄; Pam3CSK4, triacylated lipoprotein CSK4; PAMP, pathogen-associated molecular pattern; PGD₂, prostaglandin D₂; PGE₂, prostaglandin E₂; PLA₂, phospholipase A₂; Poly(I:C), polyinosinic polycytidylic acid; prostacyclin, prostaglandin I₂; sPLA₂, secreted phospholipase A₂; TRAP, thrombin receptor-activating peptide; TXA₂, thromboxane A₂; TX-M, metabolite of thromboxane A₂

show, for the first time, how loss of this fundamental enzyme system regulates phenotypes and inflammatory responses of these cardiovascular cells and associated urinary markers relevant to vascular disease.

MATERIALS AND METHODS

Blood collection and ethics

Blood was collected by venipuncture from healthy volunteers and from 2 patients (brother, patient B; sister, patient S) bearing a homozygous mutation in the PLA2G4A gene, which disrupts the active site of cPLA₂α (18). All experiments were subject to written informed consent, local ethical approval (healthy volunteer samples for platelet/leukocyte studies; St. Thomas's Hospital Research Ethics Committee, reference 07/Q0702/24: endothelial cell studies; Royal Brompton and Harfield Hospital Research Ethics Committee, reference 08/H0708/69: patient samples; South East National Health Service Research Ethics Committee), and in accordance with Declaration of Helsinki principles.

Whole-blood stimulation

Heparin-anticoagulated whole blood was incubated with vehicle (PBS), Horm collagen (Nycomed, St. Peter, Austria), thrombin receptor-activating peptide (TRAP)-6 amide (Bachem, Heidelberg, Germany), Ca²⁺ ionophore, A23187 (Sigma-Aldrich, Poole, United Kingdom), for 30 min, or with LPS (Sigma-Aldrich), triacylated lipoprotein CSK4 (Pam3CSK4; InvivoGen, Toulouse, France), bisacylated lipoprotein CGDPKHPKSF (FSL-1; InvivoGen), polyinosinic:polycytidylic acid [poly(I:C); Sigma-Aldrich], or IL-1β (Invitrogen, Life Technologies, Paisley, United Kingdom) for 18 h in the presence or absence of diclofenac (10 μM; Sigma-Aldrich). Levels of (C-X-C motif) ligand-8 (CXCL8; R&D Systems, Abingdon, United Kingdom), PGE₂ (Cisbio, Saclay, France), or TXB₂ (Cayman Chemical, Cambridge Bioscience, Cambridge, United Kingdom) were measured by immunoassay or total eicosanoids by gas chromatography–tandem mass spectrometry (see below) in the conditioned plasma.

Eicosanoid analysis

Basal and conditioned plasma was subject to eicosanomic analysis as previously described (21). Urinary prostanoid levels were determined by gas chromatography–tandem mass spectrometry as previously described (22, 23).

Light transmission aggregometry and ATP release

Platelet-rich plasma was preincubated with the COX inhibitor aspirin (30 μM; Sigma-Aldrich), the cPLA₂ inhibitor pyrrophenone (40 μM; Cayman Chemical, Cambridge Bioscience), or vehicle for 30 min at 37°C. Aggregation and ATP secretion responses to collagen (0.3–3 μg/ml), ADP (5 μM; Chrono-log; Labmedics, Abingdon, United Kingdom), U46619 (10 μM; Enzo Life Sciences, Exeter, United Kingdom), or arachidonic acid (1 mM; Sigma-Aldrich) were measured using a Chrono-log 560CA Lumi-Aggregometer (Chrono-log Corp., Havertown, PA, USA).

Platelet adhesion under flow

Whole blood was preincubated with aspirin (100 μM), pyrrophenone (40 μM), or vehicle before labeling of cells with

mepacrine (10 μ M; Sigma-Aldrich) for a further 30 min. This was then drawn through a slide chamber (Ibidi, Munich, Germany) coated with collagen (100 μ g/ml) by a syringe pump to achieve a shear rate of 1000 s^{-1} .

Endothelial cells

Blood outgrowth endothelial cells were grown out from progenitors in human blood as previously described (24–27). Once colonies emerged (between d 4 and 20), cells were expanded and maintained in Lonza EGM-2 medium (Lonza, Slough, United Kingdom) plus 10% fetal bovine serum, and experiments were performed between passages 2 and 8.

Endothelial cell immunocytochemistry

Endothelial cells were stained using anti-CD31 (platelet endothelial cell adhesion molecule-1)–Alexa Fluor 488 (BioLegend, London, United Kingdom) or anti-vascular endothelial-cadherin (Santa Cruz Biotechnology, Dallas, TX, USA) and imaged using a Cellomics VTI HCS ArrayScanner (Thermo Fisher Scientific, Hemel Hempstead, United Kingdom).

Endothelial cell eicosanoid and cytokine production

Cells were plated on 48- or 96-well plates. For eicosanoid measurements, endothelial cells were primed with IL-1 β (1 ng/ml) to up-regulate COX pathways, as described previously (28), before being treated for 30 min with A23187 or thrombin to activate PLA $_2$ or arachidonic acid to supply eicosanoid substrate directly. For inflammation studies, endothelial cells were treated with vehicle (Lonza EGM-2) or TLR ligands: heat-killed *Listeria monocytogenes* (10 7 cells/ml), Pam3CSK4 (1 μ g/ml), FSL-1 (1 μ g/ml), poly(I:C) (10 μ g/ml), LPS (1 μ g/ml), *Staphylococcus aureus*-derived flagellin (FLA; 100 ng/ml), imiquimod (1 μ g/ml), single-stranded RNA oligonucleotide-40 (1 μ g/ml), and oligodeoxynucleotide-2006 (5 μ M). After 24 h, media were collected for measurement of CXCL8 release by ELISA (R&D Systems).

Statistics and data analysis

Data are expressed as means \pm SE. Statistical analysis was performed by 1- or 2-way ANOVA or by unpaired Student's *t* test using GraphPad Prism 6 software (GraphPad Software, La Jolla, CA, USA). Patient eicosanomics data ($n = 1$ –2) were interpreted qualitatively without statistical testing.

RESULTS

Role of cPLA $_2\alpha$ in eicosanoid formation in platelets

Incubation of blood with collagen (Fig. 1A) or TRAP-6 (Fig. 1B) to specifically activate platelets increased levels of TXB $_2$ (the stable breakdown product of TXA $_2$) and 12-HETE, in particular. There were also increases in PGE $_2$, prostaglandin D $_2$ (PGD $_2$), 11-HETE, and 15-HETE. 12-HETE levels were somewhat lower in TRAP-6-stimulated blood as compared with collagen-stimulated blood. In blood treated with the Ca $^{2+}$ ionophore, A23187, to cause acute receptor-independent activation of platelets and leukocytes, a broadly similar pattern of eicosanoid formation was observed (Fig. 1C) with a marked production of 12-HETE, followed by TXB $_2$, 15-HETE, and 11-HETE.

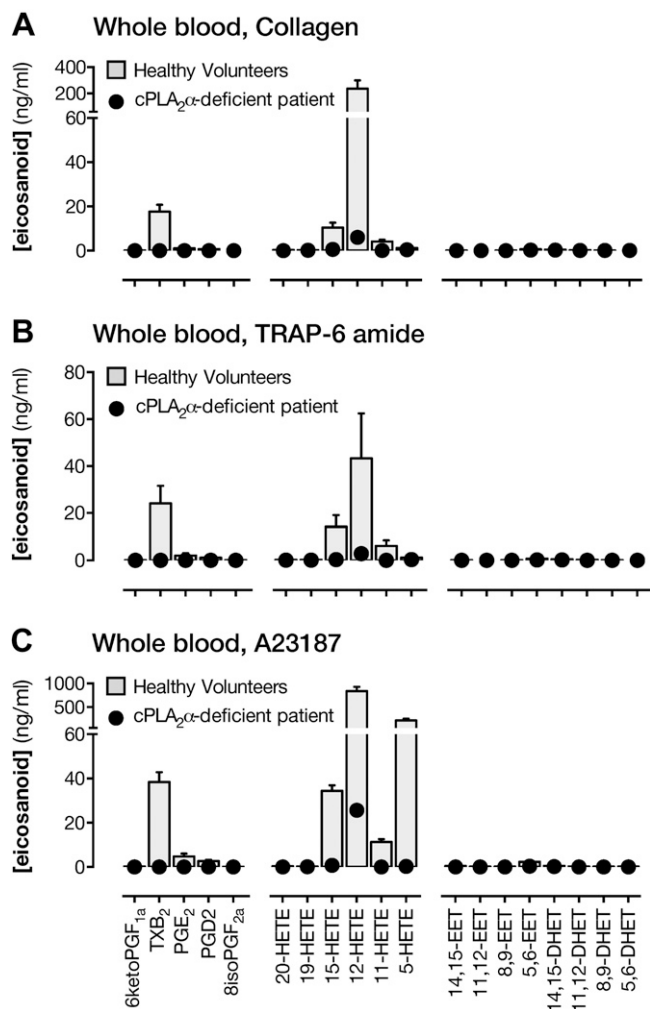


Figure 1. Contribution of cPLA $_2\alpha$ to eicosanoid synthesis in whole blood. Eicosanoid levels in whole blood from healthy volunteers or from a patient lacking cPLA $_2\alpha$ stimulated with collagen (30 μ g/ml) (A), TRAP-6 amide (30 μ M) (B), or A23187 Ca $^{2+}$ ionophore (50 μ M) (C). Levels are expressed as increase over levels in vehicle-treated blood. $n = 3$ –6 (healthy volunteers); $n = 1$ (patient S).

There were also greatly increased levels of 5-HETE, representing acute activation of leukocytes, and a more modest production of 5,6-EET. In each case, eicosanoid production to these stimuli was almost absent in blood from cPLA $_2\alpha$ -deficient patients (Fig. 1 and Supplemental Tables S1 and S2). Normal eicosanoid formation was observed in the presence of exogenous arachidonic acid in both healthy volunteer and cPLA $_2\alpha$ -deficient patient blood. In isolated platelets (platelet-rich plasma), TXB $_2$ formation induced by ADP, collagen, or the TXA $_2$ -mimetic U46619, but not exogenous arachidonic acid, was abolished by cPLA $_2\alpha$ deficiency (Supplemental Fig. S1).

Role of cPLA $_2\alpha$ in platelet aggregation, secretion, and adhesion responses

Absence of cPLA $_2\alpha$ or cPLA $_2$ inhibition by pyrrophenone produced a marked reduction in collagen-induced aggregation similar to that produced by aspirin (Fig. 2A) but had little effect upon responses to ADP or exogenous

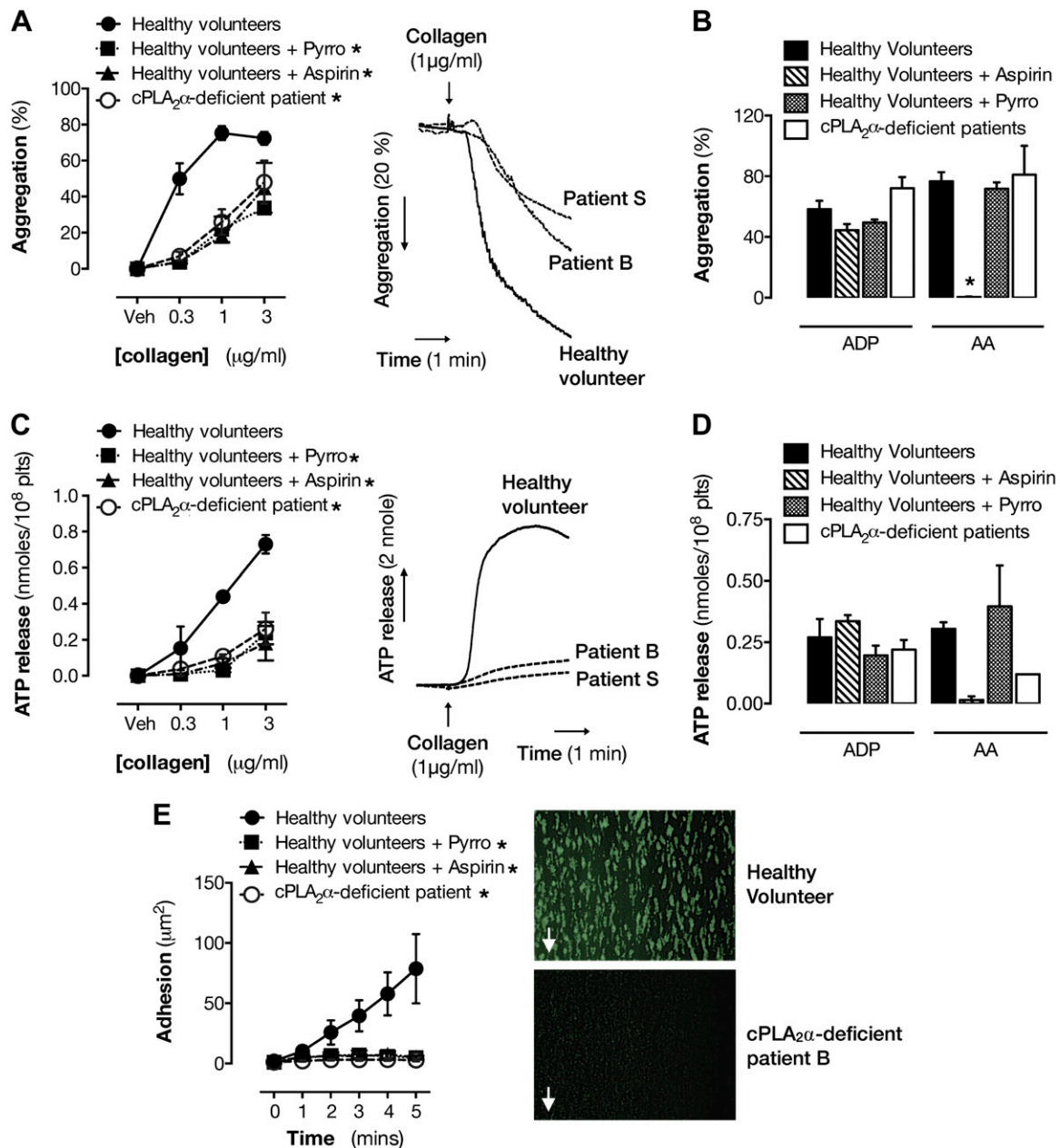


Figure 2. Effect of cPLA₂α deficiency on platelet aggregation, secretion, and adhesion responses. Effect of cPLA₂α deficiency, cPLA₂α inhibition, and aspirin on platelet aggregation to collagen (0.1–3 μg/ml) (A), ADP (5 μM) (B), and arachidonic acid (AA; 1 mM) is shown. Pyrro, pyrrophenone. ATP secretion to collagen (0.1–3 μg/ml) (C), ADP (5 μM) (D), and arachidonic acid (1 mM). E) Platelet adhesion to collagen under flow (1000 s⁻¹). n = 2–4 (healthy volunteers); n = 2 (patient B and patient S). *P < 0.05 by 2-way ANOVA with Dunnett's posttest.

arachidonic acid (Fig. 2B). ATP release induced by collagen (Fig. 2C), but not that induced by ADP or arachidonic acid (Fig. 2D), was strongly blunted by loss of functional cPLA₂α or aspirin treatment, and under flow conditions, platelet adhesion to collagen was almost abolished by cPLA₂α inhibition/deficiency (Fig. 2E).

Role of cPLA₂α in eicosanoid formation in endothelial cells

Endothelial cells from healthy volunteers or derived from cPLA₂α-deficient patients emerged in culture after 4–20 d,

grew with typical cobblestone morphology, expressed the endothelial cell markers CD31 and VE cadherin (Fig. 3A), and aligned when cultured under directional shear stress (29) (Fig. 3B). In the presence of A23187, endothelial cells from healthy volunteers released predominately prostacyclin (measured as the stable breakdown product 6-keto-PGF₁α) followed by PGE₂, PGD₂, 11-HETE, and 15-HETE. In each case, eicosanoid release was lower but not abolished in endothelial cells derived from cPLA₂α-deficient patients (Fig. 3C) (e.g., prostacyclin release from cPLA₂α-deficient endothelial cells was reduced by ~80%). Similarly, the cPLA₂ inhibitor, pyrrophenone, produced a concentration-dependent inhibition of prostacyclin release

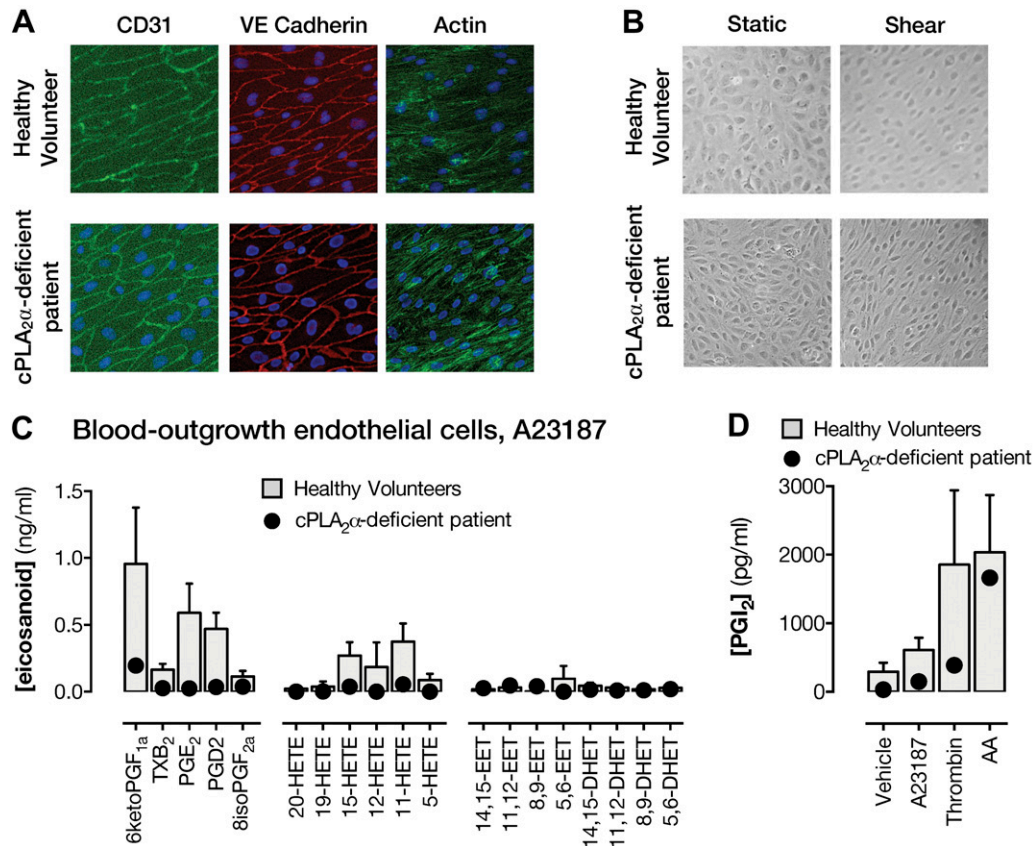


Figure 3. Phenotyping of and eicosanoid synthesis by endothelial cells grown out of blood progenitors from healthy volunteers and from a cPLA₂α-deficient patient. *A*) Endothelial-specific marker expression of CD31 (green) and VE cadherin (red) and actin staining (green). *B*) Morphologic response to shear stress after 3 d. *C*) Eicosanoid release in IL-1β (1 ng/ml)-primed endothelial cells stimulated with A23187 (50 μM). *D*) Prostacyclin release from IL-1β-primed endothelial cells stimulated for 30 min with A23187 (50 μM), thrombin (1 U/ml), or arachidonic acid (AA; 50 μM). Data are from at least 3 wells per condition. *n* = 3–6 (healthy volunteers); *n* = 1 (patient S).

from endothelial cells grown from healthy donors (Supplemental Fig. S2), with a maximal effect of ~80%. Prostacyclin was also released from endothelial cells of healthy volunteers when stimulated with the receptor-dependent activator, thrombin (Fig. 3D). As described for A23187-stimulated release above, thrombin-stimulated prostacyclin release was reduced but not abolished in cPLA₂α-deficient patient endothelial cells. Endothelial cells of both genotypes responded strongly to exogenous arachidonic acid (Fig. 3D).

Role of cPLA₂α in eicosanoid formation by leukocytes

When whole blood was stimulated (18 h) with the TLR4 agonist, LPS, to activate leukocytes and inducible biosynthetic pathways, the major eicosanoids produced were 12- and 15-HETE and PGE₂, and a smaller amount of 11-HETE (Fig. 4A and Supplemental Table S3). In cPLA₂α-deficient patient blood, LPS-induced production of PGE₂ and 15-HETE was greatly reduced, whereas the production of 12-HETE was little affected. Overall, productions were restored by acute addition of arachidonic acid (Fig. 4B and Supplemental Table S3). Pam3CSK4 (TLR2/1) and FSL-1 (TLR2/6), which activate pattern recognition receptors associated with gram-positive bacteria, as with LPS, activated

whole blood to release PGE₂, an effect that was abolished by cPLA₂α deficiency (Fig. 4C). Neither Poly(I:C), which stimulates the viral pathogen recognition receptor TLR3, nor IL-1β, which works independently of pattern recognition receptors, stimulated PGE₂ release from whole blood.

Role of cPLA₂α in inflammatory responses in endothelial cells and blood leukocytes

Whole blood from healthy volunteers treated with FSL-1, Poly(I:C), or LPS, but not with IL-1β, released the inflammatory chemokine CXCL8 (Fig. 5A). Blood from a cPLA₂α-deficient patient exhibited more than 5-fold greater responses to all agents except IL-1β as compared with matched controls (Fig. 5A). Treatment of blood from healthy volunteers with the COX inhibitor diclofenac suppressed the CXCL8 response to LPS but did not modify CXCL8 release stimulated by other tested agents (Fig. 5A).

Endothelial cells from healthy donors also released CXCL8 when stimulated with pathogen-associated molecular patterns (PAMPs) directed at TLR2, 3, or 4, or with IL-1β. Again, as with leukocytes in whole blood, endothelial cells from a cPLA₂α-deficient patient released elevated levels of CXCL8 when stimulated with inflammatory

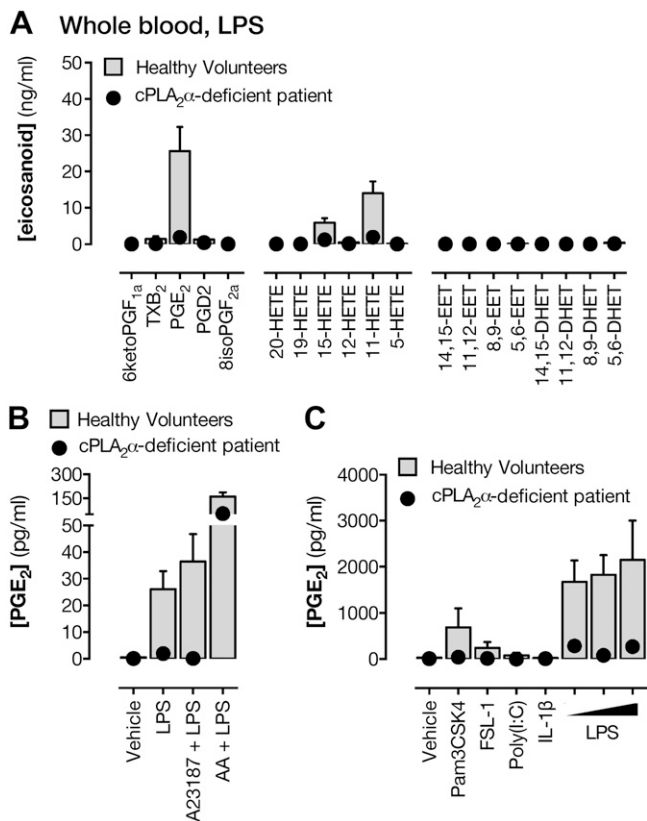


Figure 4. Contribution of cPLA₂α to eicosanoid synthesis in leukocytes. *A*) Eicosanoid levels in whole blood from healthy volunteers or from a patient lacking cPLA₂α treated with LPS (10 μg/ml) for 18 h. PGE₂ formation in whole blood treated with LPS (10 μg/ml) for 18 h followed by addition of A23187 (50 μM) or arachidonic acid (AA; 1 mM) for 30 min (*B*) or treated with agonists to TLR2/1 (Pam3CSK4; 1 μg/ml), TLR2/6 (FSL-1; 1 μg/ml), TLR3 [poly(I:C); 10 μg/ml], IL-1 receptor (IL-1β; 1 ng/ml), or TLR4 (LPS; 5–20 μg/ml) (*C*). *n* = 3–6 (healthy volunteers); *n* = 1 (patient S).

agents (Fig. 5*B*). Endothelial cells from either type of donor did not respond to ligands for TLR5, TLR7, or TLR8 (Fig. 5*B*).

Involvement of cPLA₂α in plasma and urinary eicosanoid levels

Plasma from healthy volunteers contained primarily metabolites of linoleic acid, eicosapentaenoic acid (EPA), and docosahexaenoic acid (DHA). Patients lacking cPLA₂α had reduced levels of these mediators compared with plasma from healthy volunteers (Fig. 6*A*). Basal plasma also contained substantial levels of 12-HETE, and this remained in cPLA₂α-deficient patient plasma.

Absence of cPLA₂α was associated with strong reductions in the levels of leukotriene E₄ (LTE₄) and prostacyclin metabolites (Fig. 6*B, C*), whereas substantial levels of PGD₂, PGE₂, and 8-isoprostane metabolites remained (Fig. 6*D–G*). Levels of the urinary metabolite of thromboxane A₂ (TX-M) were 50% lower in cPLA₂α-deficient patients as compared with healthy volunteers (0.202 ± 0.010 ng/mg creatinine *vs.* 0.101 ± 0.017 ng/mg creatinine; *P* < 0.01; Fig. 6*E*). In addition, substantial levels of PGD₂, PGE₂, and

8-isoprostane metabolites remained in urine samples from cPLA₂α-deficient patients.

DISCUSSION

Here, we have examined the contribution of cPLA₂α to eicosanoid formation, and thrombotic and inflammatory responses in platelets, blood leukocytes, and endothelial cells from 2 individuals with a unique genetic inactivation of this enzyme. Although we (18) and others (19, 20) have published reports of individuals lacking functional cPLA₂α, including limited analysis of platelet responses, this is the first time a full and systematic eicosanoid analysis has been undertaken on samples from these patients and considered in the context of the circulatory system in health and inflammation. These data demonstrate an absolute requirement for cPLA₂α in eicosanoid synthesis in the vascular compartment with a consequent loss of

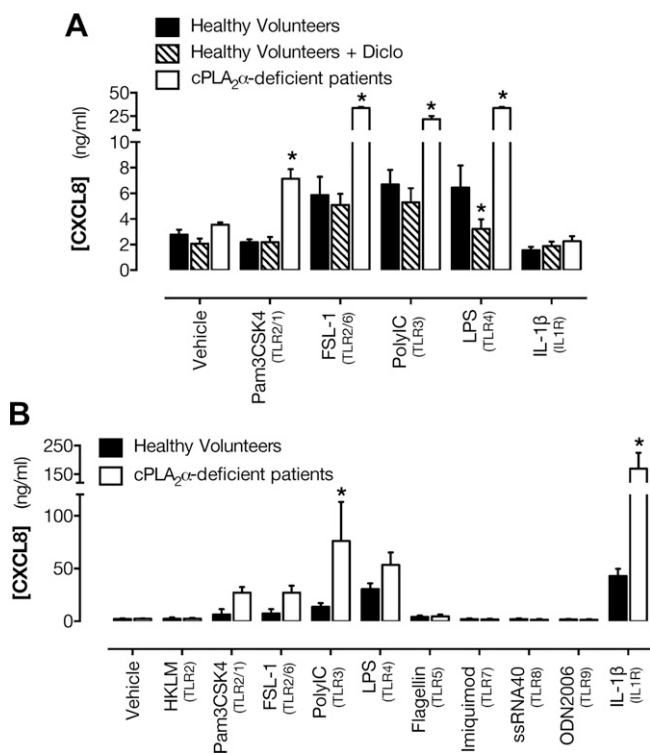


Figure 5. Effect of cPLA₂α deficiency on blood leukocyte and endothelial cell inflammatory responses. *A*) CXCL8 release in whole blood from healthy volunteers with or without pretreatment with the COX inhibitor diclofenac (Diclo; 10 μM) or a cPLA₂α-deficient patient in response to agonists to TLR2/1 (Pam3CSK4; 1 μg/ml), TLR2/6 (FSL-1; 1 μg/ml), TLR3 [poly(I:C); 10 μg/ml], TLR4 (LPS; 1 μg/ml), or IL-1 receptor (IL-1β; 1 ng/ml). *B*) CXCL8 release by endothelial cells to agonists of TLR2 [heat-killed *L. monocytogenes* (HKLM); 10⁷ cells/ml], TLR2/1 (Pam3CSK4; 1 μg/ml), TLR2/6 (FSL-1; 1 μg/ml), TLR3 [poly(I:C); 10 μg/ml], TLR4 (LPS; 10 μg/ml), TLR5 (FLA; 100 ng/ml), TLR7 (imiquimod; 1 μg/ml), TLR8 [single-stranded RNA oligonucleotide-40 (ssRNA40); 1 μg/ml], TLR9 [oligodeoxynucleotide-2006 (ODN2006); 5 μM], or IL-1 receptor (IL-1β; 1 ng/ml). *n* = 3–6 (healthy volunteers; 2 determinations each); *n* = 1 (patient S; 3 determinations each). **P* < 0.05 by 2-way ANOVA with Dunnett's posttest.

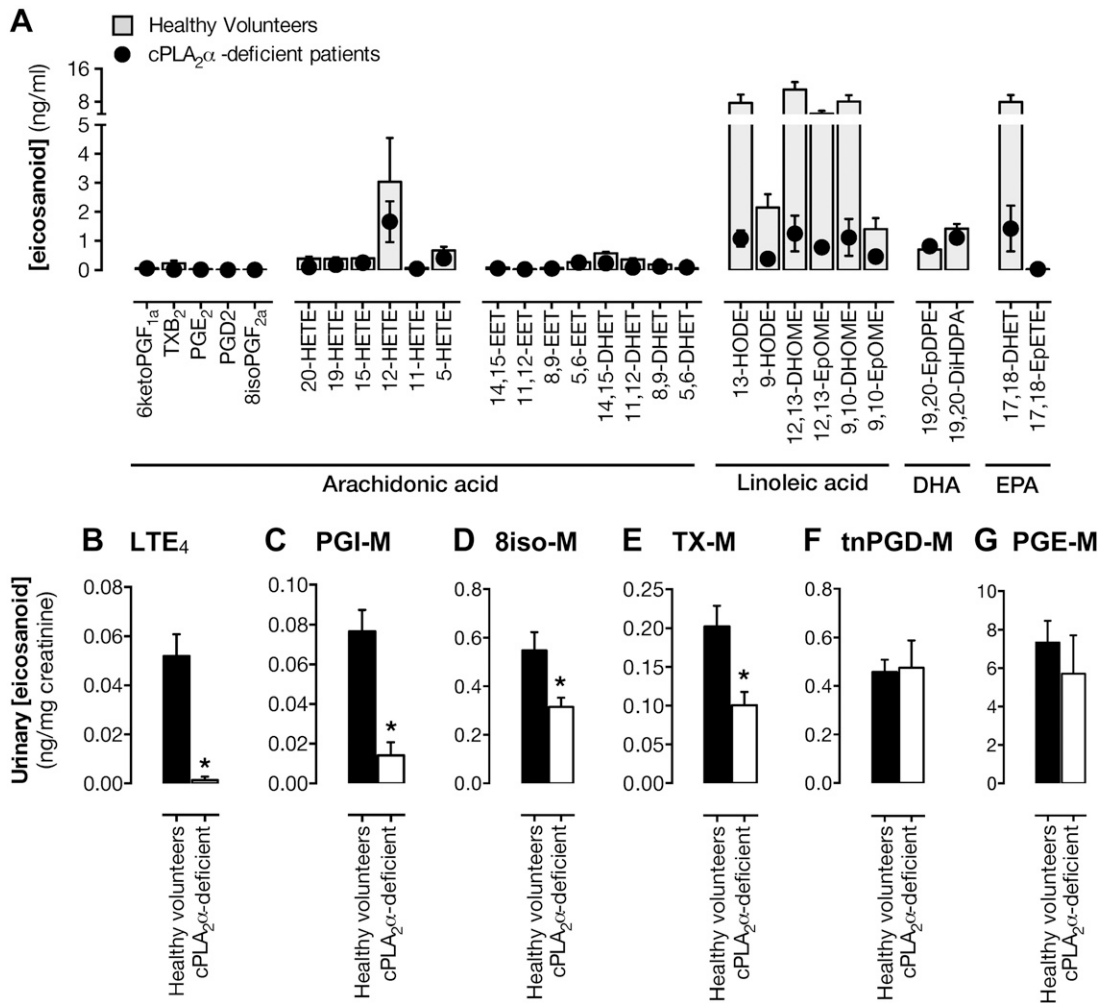


Figure 6. Contribution of cPLA₂α to basal plasma and urinary eicosanoid levels. A) Basal eicosanoid levels in plasma from healthy volunteers (circles) or from a patient lacking cPLA₂α. *n* = 8 (healthy volunteers; 2 determinations each); *n* = 2 (patient B and patient S; 2 determinations each). Urinary levels of LTE₄ (B) and metabolites of prostacyclin (PGI-M) (C), 8-isoprostane (8iso-M) (D), TXA₂ (TX-M) (E), PGD₂ [tetranor (tn)PGD-M] (F), and PGE₂ (PGE-M) (G) in healthy volunteers (filled columns) or from patients lacking cPLA₂α (unfilled columns). *n* = 7 (healthy volunteers; 2 determinations each); *n* = 2 (patient B and patient S; 4 determinations each).

platelet activation pathways, reduced antithrombotic prostacyclin, and increased inflammatory sensitivity of both endothelial cells and leukocytes.

Platelets

The PLA₂ system in platelets is among the most clearly defined in cardiovascular cell types. We and others have previously performed limited phenotyping of platelets from cPLA₂α-deficient individuals and found a requirement for this enzyme in TXA₂ formation and collagen-induced platelet aggregation, which is TXA₂ dependent. However, in addition to cPLA₂α, platelets also express sPLA₂, which others suggest contributes to eicosanoid formation in platelets (9, 10, 30). Here, for the first time, we have performed a full eicosanomic analysis of samples from cPLA₂α-deficient individuals to consider the role of this enzyme in synthesis of a range of functional distinct arachidonic acid-derived mediators. Stimulation of whole blood with the platelet activators collagen or

TRAP-6 resulted in greatly increased synthesis of TXA₂, in addition to PGE₂, PGD₂, and 11-, 12-, and 15-HETE, mediators primarily produced by COX and LOX pathways. 12-HETE levels were somewhat lower in TRAP-6-stimulated blood as compared to collagen-stimulated blood, consistent with reports that 12-LOX activation is linked to the platelet collagen receptor, glycoprotein VI (7). Stimulation of blood with the receptor-independent activator Ca²⁺ ionophore A23187 produced a similar platelet eicosanoid fingerprint, but unlike collagen and TRAP-6, increased levels of 5-HETE, reflecting acute activation of leukocytes. In each case, eicosanoid production was cPLA₂α dependent because it was lost in cPLA₂α-deficient patient blood but reversed by the addition of exogenous arachidonic acid, demonstrating its dependence on loss of endogenous substrate release. In agreement, isolated cPLA₂α-deficient platelets stimulated with a range of agonists (collagen, ADP, and U46619), but not exogenous arachidonic acid, exhibited a complete loss of TXA₂ synthesis, in contrast to reports that ADP-induced release is not altered in cPLA₂α-knockout mouse platelets (9). These data

illustrate the requirement for cPLA₂α in the full range of eicosanoids synthesized by platelets and that this is independent of the stimulus used (9, 10).

We next set out to establish the contribution of cPLA₂α-derived eicosanoids to functional platelet aggregation responses. Although it is well known that the platelet COX product TXA₂ is a powerful proaggregatory agent, this response may be modified by other eicosanoid mediators synthesized in parallel. Indeed, PGE₂ (31), 12-HETE (32–34), 15-HETE (35), and 5,6-EET (36) increase platelet activation, whereas PGD₂ (37) and higher levels of PGE₂ may limit platelet activation (31), meaning the net contribution of cPLA₂α-derived eicosanoids is unclear. Our studies using traditional light transmission lumi-aggregometry and 96-well plate aggregometry demonstrated that inhibition or absence of cPLA₂α produced a marked reduction in collagen-induced platelet aggregation and dense granule (ATP) secretion, in agreement with what we (18) and others (9, 19, 20) have previously reported. These defects were rescued by exogenous arachidonic acid, demonstrating that they were specifically associated with loss of endogenous substrate release. Similarly, platelet adhesion to a collagen-coated surface in flowing blood was abolished by cPLA₂ inhibition and absent in blood from cPLA₂α-deficient patients. These data are in agreement with reports of the importance of cPLA₂α and TXA₂ generation in platelet adhesion (38). In each of these functional assays, the reduction observed was similar to that produced by the COX inhibitor aspirin, suggesting that regulation of collagen-induced platelet responses by cPLA₂α is due to products of platelet COX-1, probably TXA₂. Overall, these data show that cPLA₂α is absolutely required for formation of eicosanoid mediators in platelets and that despite the synthesis of several eicosanoid families, the contribution of cPLA₂α to platelet aggregation, secretion, and adhesion responses can be entirely accounted for by generation of COX products. This reduction in platelet function is consistent with an increased tendency to bruising noted in the clinical care of these patients.

Endothelium

Through eicosanoid release, endothelial cells are key to health and disease of the circulation. Here, we have made use of endothelial cells isolated from blood progenitors providing the first opportunity to study genetic deficiency of cPLA₂α in human endothelium. Endothelial cells from a cPLA₂α-deficient patient expressed the normal endothelial cell markers CD31 and vascular endothelial-cadherin, had a cobblestone morphology, and when cultured under conditions of chronic (3 d) laminar shear stress (29), aligned with the direction of shear, demonstrating their endothelial phenotype. When we examined eicosanoid production by endothelial cells, A23187 stimulation increased production of several eicosanoid mediators, the most abundant of which was prostacyclin, with lower levels of PGE₂, PGD₂, and 11-, 12-, and 15-HETE. These were predominantly driven by cPLA₂α because they were strongly reduced in cPLA₂α-deficient endothelial cells. This was further confirmed by the ability of a selective cPLA₂ inhibitor to prevent the majority of A23187-stimulated prostacyclin production by endothelial cells and was

specific because cPLA₂α-deficient endothelial cells responded normally to exogenous arachidonic acid. However, cPLA₂α-deficient endothelial cells stimulated with either A23187 or thrombin continued to produce some prostacyclin, probably reflecting contribution of other PLA₂ isoforms [*e.g.*, group VIA iPLA₂ (also referred to as iPLA₂β)] to eicosanoid generation in endothelium (11, 12).

Leukocytes and inflammation

In parallel with platelet and endothelial cell studies, we examined the effect of addition of inflammatory stimuli (*e.g.*, LPS) to whole blood to investigate the role of cPLA₂α in blood leukocyte responses, an approach frequently applied in the eicosanoid field (3, 39). When whole blood was stimulated with A23187, in addition to platelet-derived mediators, 5-HETE was detected, which is associated with 5-LOX present in monocytes and neutrophils (40). When blood was stimulated with LPS to specifically activate leukocytes and inducible biosynthetic pathways such as COX-2, a more characteristic inflammatory eicosanoid profile was produced with PGE₂, 12- and 15-HETE being the most abundant products. In each case, eicosanoid synthesis was cPLA₂α mediated. In cPLA₂α-deficient patient blood, LPS-induced production of PGE₂ and 15-HETE was greatly reduced, and overall, productions were restored by acute addition of arachidonic acid, confirming that this defect is due to loss of free endogenous arachidonic acid. In contrast, in LPS-stimulated blood, the production of 12-HETE was little affected by cPLA₂α deficiency, suggesting that other PLA₂ isoforms specifically couple to 12-HETE synthesis in blood leukocytes. By its actions on TLR4, LPS mimics the effects of gram-negative bacteria. However, other pathogens activate immune and inflammatory responses in tissues through different pattern recognition receptor signaling pathways, each of which could theoretically drive eicosanoid production by different PLA₂ isoforms. To address this, we studied the effect of a full range of PAMPs that mimic gram-positive, as well as gram-negative, bacteria or viruses. Thus, Pam3CSK4 (TLR2/1) and FSL-1 (TLR2/6), which activate pattern recognition receptors associated with gram-positive bacteria, as with LPS, activated whole blood to release PGE₂, an effect that was abolished by cPLA₂α deficiency. Neither Poly(I:C), which stimulates the viral pathogen recognition receptor TLR3, nor IL-1β, which works independently of pattern recognition receptors, stimulated PGE₂ release from whole blood. Although these data demonstrate that cPLA₂α is central to leukocyte eicosanoid synthesis, particularly for PGE₂, there are clearly roles for other PLA₂ isoforms such as sPLA₂ (13).

To understand the implications of loss of eicosanoid production to the inflammatory response, we measured CXCL8 production, induction of which reflects both primary activation of inflammatory transcriptional pathways such as NF-κB pathways and subsequent secretion of TNF-α and IL-1β (20, 41). Whole blood from healthy volunteers treated with FSL-1, Poly(I:C), or LPS, but not with IL-1β, released the inflammatory chemokine CXCL8. Blood from a cPLA₂α-deficient patient exhibited more than 5-fold greater responses to all agents except IL-1β as

compared with matched controls (Fig. 5A). Treatment of blood from healthy volunteers with the COX inhibitor diclofenac suppressed the CXCL8 response to LPS but did not modify CXCL8 release stimulated by other tested agents (Fig. 5A), indicating that the effect was not mediated by loss of COX metabolites. Although it cannot be excluded that cPLA₂α-deficient patient blood contains altered leukocyte subsets, blood constituents, or other confounding influences, these data suggest that cPLA₂α-dependent mediators, other than COX products, act to suppress cytokine responses by blood leukocytes. This effect may reflect loss of 11- and/or 15-HETE synthesis because these were also detected in LPS-stimulated whole blood, and it has been previously reported that 15-HETE has anti-inflammatory activity (42, 43).

Similarly, endothelial cells from healthy donors released CXCL8 when stimulated with PAMPs directed at TLR2, 3, or 4, or with IL-1β. As with leukocytes in whole blood, endothelial cells from a cPLA₂α-deficient patient released elevated levels of CXCL8 when stimulated with inflammatory agents, consistent with activation of NF-κB pathways following treatment with inflammatory stimuli, as we have previously described (25). Endothelial cells from either type of donor did not respond to ligands for TLR5, the pattern recognition receptor for motile bacteria and fungi, TLR7 and TLR8, pattern recognition receptors for single-stranded RNA viruses, or TLR9, which is the pattern recognition receptor for bacterial DNA. Importantly, in contrast to blood leukocyte studies, these endothelial cells constitute a single, defined cell type in a controlled medium suggesting that any differences observed likely reflect changes in eicosanoid production as compared to confounding factors present in blood cells. Because prostacyclin was the most abundant eicosanoid produced by endothelial cells and is a powerful inhibitor of vascular inflammation, this proinflammatory phenotype of cPLA₂α-deficient endothelial cells is most easily explained by loss of this fundamental vascular hormone. CXCL8 is a potent neutrophil chemotactic factor, which has been implicated in atherosclerosis (44); thus, augmented production of CXCL8 and potentially other NF-κB-driven cytokines is likely to be detrimental to cardiovascular health. Moreover, taken together, these studies demonstrate that on a global level, blood leukocytes and endothelial cells require cPLA₂α to produce eicosanoids in response to a range of inflammatory stimuli, and this exerts both COX-dependent and possibly COX-independent regulation of cytokine production and, by inference, immunologic/inflammatory defenses, consistent with clinical manifestations in these patients (18, 19).

Production *in vivo*

Finally, to provide an overview of the contribution of cPLA₂α to eicosanoid formation from all sources in the body, we measured the eicosanoid profile in plasma and specific urinary eicosanoid metabolites. Plasma from healthy volunteers contained low levels of primarily metabolites of linoleic acid, EPA, and DHA. Patients lacking cPLA₂α had reduced levels of these mediators compared with plasma from healthy volunteers. Because cPLA₂α has strong specificity for arachidonate-containing

phospholipids, this may reflect altered physiology in these patients. Notably, basal plasma also contained substantial levels of 12-HETE, which may reflect platelet activation during blood sampling; as noted above, 12-HETE is the major product of activated platelets. However, surprisingly, a small 12-HETE peak was also seen in cPLA₂α-deficient patient plasma, suggesting possible cPLA₂α-independent eicosanoid formation in the body.

Interpretation of plasma eicosanoid data as representative of a circulating pool is difficult because levels may reflect local vascular activation during blood sampling, and many eicosanoids rapidly degrade/clear from the circulation. For this reason, many favor measurement of urinary metabolites to assess *in vivo* eicosanoid production. Using this approach, we observed that absence of cPLA₂α was associated with strong reductions in the levels of LTE₄, prostacyclin, and TXA₂ metabolites, consistent with the reductions in TXA₂ production by platelets, prostacyclin production by endothelial cells, and 5-HETE production by monocytes/neutrophils [LTE₄ is a downstream metabolite of 5-LOX products (40)] that we noted in our *in vitro* cell studies. Of particular relevance to platelet function was the urinary TXA₂ metabolite, TX-M. This has been often recommended as a marker of platelet activation *in vivo* that could be used to gauge the efficacy of aspirin treatment and the level of ongoing platelet activation (45). We noted that whereas platelets from cPLA₂α-deficient patients did not produce TXA₂, urinary levels of TX-M in the patients were reduced only by ~50%. This demonstrates that urinary TX-M does not specifically report production from platelets and adds to a growing body of evidence questioning the origin of TX-M and other urinary prostanoid metabolites (23, 46). In addition, substantial levels of PGD₂, PGE₂, and 8-isoprostane metabolites remained in urine samples from cPLA₂α-deficient patients, further suggesting that there are sites in the body where considerable cPLA₂α-independent prostanoid formation occurs.

CONCLUSIONS

Here, we have examined the contribution of cPLA₂α to eicosanoid formation, and thrombotic and inflammatory responses in platelets, blood leukocytes, and endothelial cells from individuals with a unique genetic inactivation of this enzyme. Our data demonstrate an absolute requirement for cPLA₂α in eicosanoid synthesis in the vascular compartment with a consequent loss of platelet activation pathways, reduced antithrombotic prostacyclin, and increased inflammatory sensitivity of both endothelial cells and leukocytes. This study unites many conflicting observations in the literature and provides a definitive account of the rate-limiting and perhaps most fundamental component of this system, cPLA₂α. FJ

The authors acknowledge the cytosolic phospholipase A₂-deficient individuals who made these studies possible by volunteering to provide blood samples. This research was supported by an Imperial College Junior Research Fellowship (to N.S.K.), Wellcome Trust program grant (0852551Z108/Z to J.A.M. and T.D.W.), British Heart Foundation Ph.D. studentship (FS/10/033/28271 to F.R.), British Heart Foundation project grant (PG/11/39/28890 to D.B.-B.), and by the Intramural Research Program of the U.S. National Institutes of

REFERENCES

- Warner, T. D., Nylander, S., and Whatling, C. (2011) Anti-platelet therapy: cyclo-oxygenase inhibition and the use of aspirin with particular regard to dual anti-platelet therapy. *Br. J. Clin. Pharmacol.* **72**, 619–633
- Mitchell, J. A., and Warner, T. D. (2006) COX isoforms in the cardiovascular system: understanding the activities of non-steroidal anti-inflammatory drugs. *Nat. Rev. Drug Discov.* **5**, 75–86
- Warner, T. D., Giuliano, F., Vojnovic, I., Bukasa, A., Mitchell, J. A., and Vane, J. R. (1999) Nonsteroid drug selectivities for cyclo-oxygenase-1 rather than cyclo-oxygenase-2 are associated with human gastrointestinal toxicity: a full in vitro analysis. *Proc. Natl. Acad. Sci. USA* **96**, 7563–7568
- Murakami, M., Taketomi, Y., Miki, Y., Sato, H., Hirabayashi, T., and Yamamoto, K. (2011) Recent progress in phospholipase A2 research: from cells to animals to humans. *Prog. Lipid Res.* **50**, 152–192
- Burke, J. E., and Dennis, E. A. (2009) Phospholipase A2 structure/function, mechanism, and signaling. *J. Lipid Res.* **50** (Suppl), S237–S242
- Polgár, J., Kramer, R. M., Um, S. L., Jakubowski, J. A., and Clemetson, K. J. (1997) Human group II 14 kDa phospholipase A2 activates human platelets. *Biochem. J.* **327**, 259–265
- Coffey, M. J., Jarvis, G. E., Gibbins, J. M., Coles, B., Barrett, N. E., Wylie, O. R., and O'Donnell, V. B. (2004) Platelet 12-lipoxygenase activation via glycoprotein VI: involvement of multiple signaling pathways in agonist control of H(P)ETE synthesis. *Circ. Res.* **94**, 1598–1605
- Houliester, R. A., and Wheeler-Jones, C. P. (2001) sPLA(2) cooperates with cPLA(2)alpha to regulate prostacyclin synthesis in human endothelial cells. *Biochem. Biophys. Res. Commun.* **287**, 881–887
- Wong, D. A., Kita, Y., Uozumi, N., and Shimizu, T. (2002) Discrete role for cytosolic phospholipase A(2)alpha in platelets: studies using single and double mutant mice of cytosolic and group IIA secretory phospholipase A(2). *J. Exp. Med.* **196**, 349–357
- Blache, D., Gautier, T., Tietge, U. J., and Lagrost, L. (2012) Activated platelets contribute to oxidized low-density lipoproteins and dysfunctional high-density lipoproteins through a phospholipase A2-dependent mechanism. *FASEB J.* **26**, 927–937
- Sharma, J., Turk, J., and McHowat, J. (2010) Endothelial cell prostaglandin I(2) and platelet-activating factor production are markedly attenuated in the calcium-independent phospholipase A(2)beta knockout mouse. *Biochemistry* **49**, 5473–5481
- Wong, M. S., Man, R. Y., and Vanhoutte, P. M. (2010) Calcium-independent phospholipase A(2) plays a key role in the endothelium-dependent contractions to acetylcholine in the aorta of the spontaneously hypertensive rat. *Am. J. Physiol. Heart Circ. Physiol.* **298**, H1260–H1266
- Marshall, J., Krump, E., Lindsay, T., Downey, G., Ford, D. A., Zhu, P., Walker, P., and Rubin, B. (2000) Involvement of cytosolic phospholipase A2 and secretory phospholipase A2 in arachidonic acid release from human neutrophils. *J. Immunol.* **164**, 2084–2091
- Balsinde, J., and Dennis, E. A. (1996) Distinct roles in signal transduction for each of the phospholipase A2 enzymes present in P388D1 macrophages. *J. Biol. Chem.* **271**, 6758–6765
- Nicholls, S. J., Kastelein, J. J., Schwartz, G. G., Bash, D., Rosenson, R. S., Cavender, M. A., Brennan, D. M., Koening, W., Jukema, J. W., Nambi, V., Wright, R. S., Menon, V., Lincoff, A. M., and Nissen, S. E.; VISTA-16 Investigators. (2014) Varespladib and cardiovascular events in patients with an acute coronary syndrome: the VISTA-16 randomized clinical trial. *JAMA* **311**, 252–262
- Riendeau, D., Guay, J., Weech, P. K., Laliberté, F., Yergey, J., Li, C., Desmarais, S., Perrier, H., Liu, S., and Nicoll-Griffith, D. (1994) Arachidonyl trifluoromethyl ketone, a potent inhibitor of 85-kDa phospholipase A2, blocks production of arachidonate and 12-hydroxyeicosatetraenoic acid by calcium ionophore-challenged platelets. *J. Biol. Chem.* **269**, 15619–15624
- Bartoli, F., Lin, H. K., Ghomashchi, F., Gelb, M. H., Jain, M. K., and Apitz-Castro, R. (1994) Tight binding inhibitors of 85-kDa phospholipase A2 but not 14-kDa phospholipase A2 inhibit release of free arachidonate in thrombin-stimulated human platelets. *J. Biol. Chem.* **269**, 15625–15630
- Brooke, M. A., Longhurst, H. J., Plagnol, V., Kirkby, N. S., Mitchell, J. A., Rüschemdorf, F., Warner, T. D., Kelsell, D. P., and MacDonald, T. T. (2014) Cryptogenic multifocal ulcerating stenosing enteritis associated with homozygous deletion mutations in cytosolic phospholipase A2- α . *Gut* **63**, 96–104
- Adler, D. H., Cogan, J. D., Phillips III, J. A., Schnetz-Boutaud, N., Milne, G. L., Iverson, T., Stein, J. A., Brenner, D. A., Morrow, J. D., Boutaud, O., and Oates, J. A. (2008) Inherited human cPLA(2alpha) deficiency is associated with impaired eicosanoid biosynthesis, small intestinal ulceration, and platelet dysfunction. *J. Clin. Invest.* **118**, 2121–2131
- Faioni, E. M., Razzari, C., Zulueta, A., Femia, E. A., Fenu, L., Trinchera, M., Podda, G. M., Pugliano, M., Marongiu, F., and Cattaneo, M. (2014) Bleeding diathesis and gastro-duodenal ulcers in inherited cytosolic phospholipase-A2 alpha deficiency. *Thromb. Haemost.* **112**, 1182–1189
- Newman, J. W., Watanabe, T., and Hammock, B. D. (2002) The simultaneous quantification of cytochrome P450 dependent linoleate and arachidonate metabolites in urine by HPLC-MS/MS. *J. Lipid Res.* **43**, 1563–1578
- Daniel, V. C., Minton, T. A., Brown, N. J., Nadeau, J. H., and Morrow, J. D. (1994) Simplified assay for the quantification of 2,3-dinor-6-keto-prostaglandin F1 alpha by gas chromatography-mass spectrometry. *J. Chromatogr. B Biomed. Appl.* **653**, 117–122
- Kirkby, N. S., Lundberg, M. H., Harrington, L. S., Leadbeater, P. D., Milne, G. L., Potter, C. M., Al-Yamani, M., Adeyemi, O., Warner, T. D., and Mitchell, J. A. (2012) Cyclooxygenase-1, not cyclooxygenase-2, is responsible for physiological production of prostacyclin in the cardiovascular system. *Proc. Natl. Acad. Sci. USA* **109**, 17597–17602
- Martin-Ramirez, J., Hofman, M., van den Biggelaar, M., Hebbel, R. P., and Voorberg, J. (2012) Establishment of outgrowth endothelial cells from peripheral blood. *Nat. Protoc.* **7**, 1709–1715
- Reed, D. M., Foldes, G., Gatheral, T., Paschalaki, K. E., Lendvai, Z., Bagyura, Z., Nemeth, T., Skopal, J., Merkely, B., Telcian, A. G., Gogsadze, L., Edwards, M. R., Gough, P. J., Bertin, J., Johnston, S. L., Harding, S. E., and Mitchell, J. A. (2014) Pathogen sensing pathways in human embryonic stem cell derived-endothelial cells: role of NOD1 receptors. *PLoS One* **9**, e91119
- Starke, R. D., Ferraro, F., Paschalaki, K. E., Dryden, N. H., McKinnon, T. A., Sutton, R. E., Payne, E. M., Haskard, D. O., Hughes, A. D., Cutler, D. F., Laffan, M. A., and Randi, A. M. (2011) Endothelial von Willebrand factor regulates angiogenesis. *Blood* **117**, 1071–1080
- Starke, R. D., Paschalaki, K. E., Dyer, C. E., Harrison-Lavoie, K. J., Cutler, J. A., McKinnon, T. A., Millar, C. M., Cutler, D. F., Laffan, M. A., and Randi, A. M. (2013) Cellular and molecular basis of von Willebrand disease: studies on blood outgrowth endothelial cells. *Blood* **121**, 2773–2784
- Boutaud, O., Aronoff, D. M., Richardson, J. H., Marnett, L. J., and Oates, J. A. (2002) Determinants of the cellular specificity of acetaminophen as an inhibitor of prostaglandin H(2) synthases. *Proc. Natl. Acad. Sci. USA* **99**, 7130–7135
- Potter, C. M., Lundberg, M. H., Harrington, L. S., Warboys, C. M., Warner, T. D., Berson, R. E., Moshkov, A. V., Gorelik, J., Weinberg, P. D., and Mitchell, J. A. (2011) Role of shear stress in endothelial cell morphology and expression of cyclooxygenase isoforms. *Arterioscler. Thromb. Vasc. Biol.* **31**, 384–391
- Coffey, M. J., Coles, B., Locke, M., Bermudez-Fajardo, A., Williams, P. C., Jarvis, G. E., and O'Donnell, V. B. (2004) Interactions of 12-lipoxygenase with phospholipase A2 isoforms following platelet activation through the glycoprotein VI collagen receptor. *FEBS Lett.* **576**, 165–168
- Iyü, D., Glenn, J. R., White, A. E., Johnson, A. J., Fox, S. C., and Heptinstall, S. (2010) The role of prostanoid receptors in mediating the effects of PGE(2) on human platelet function. *Platelets* **21**, 329–342
- Katoh, A., Ikeda, H., Murohara, T., Haramaki, N., Ito, H., and Imaizumi, T. (1998) Platelet-derived 12-hydroxyeicosatetraenoic acid plays an important role in mediating canine coronary thrombosis by regulating platelet glycoprotein IIb/IIIa activation. *Circulation* **98**, 2891–2898
- Yeung, J., Apopa, P. L., Vesci, J., Stolla, M., Rai, G., Simeonov, A., Jadhav, A., Fernandez-Perez, P., Maloney, D. J., Boutaud, O., Holman, T. R., and Holinstat, M. (2013) 12-lipoxygenase activity plays an important role in PAR4 and GPVI-mediated platelet reactivity. *Thromb. Haemost.* **110**, 569–581

34. Mais, D. E., Saussy, Jr., D. L., Magee, D. E., and Bowling, N. L. (1990) Interaction of 5-HETE, 12-HETE, 15-HETE and 5,12-diHETE at the human platelet thromboxane A₂/prostaglandin H₂ receptor. *Eicosanoids* **3**, 121–124
35. Vijil, C., Hermansson, C., Jeppsson, A., Bergström, G., and Hultén, L. M. (2014) Arachidonate 15-lipoxygenase enzyme products increase platelet aggregation and thrombin generation. *PLoS One* **9**, e88546
36. Ben-Amor, N., Redondo, P. C., Bartegi, A., Pariente, J. A., Salido, G. M., and Rosado, J. A. (2006) A role for 5,6-epoxyeicosatrienoic acid in calcium entry by de novo conformational coupling in human platelets. *J. Physiol.* **570**, 309–323
37. Song, W. L., Stubbe, J., Ricciotti, E., Alamuddin, N., Ibrahim, S., Crichton, I., Prempeh, M., Lawson, J. A., Wilensky, R. L., Rasmussen, L. M., Puré, E., and FitzGerald, G. A. (2012) Niacin and biosynthesis of PGD₂ by platelet COX-1 in mice and humans. *J. Clin. Invest.* **122**, 1459–1468
38. Prévost, N., Mitsios, J. V., Kato, H., Burke, J. E., Dennis, E. A., Shimizu, T., and Shattil, S. J. (2009) Group IVA cytosolic phospholipase A₂ (cPLA₂α) and integrin αIIbβ₃ reinforce each other's functions during αIIbβ₃ signaling in platelets. *Blood* **113**, 447–457
39. Patrignani, P., Panara, M. R., Greco, A., Fusco, O., Natoli, C., Iacobelli, S., Cipollone, F., Ganci, A., Créminon, C., and Maclouf, J. (1994) Biochemical and pharmacological characterization of the cyclooxygenase activity of human blood prostaglandin endoperoxide synthases. *J. Pharmacol. Exp. Ther.* **271**, 1705–1712
40. Rådmark, O., Werz, O., Steinhilber, D., and Samuelsson, B. (2015) 5-Lipoxygenase, a key enzyme for leukotriene biosynthesis in health and disease. *Biochim. Biophys. Acta* **1851**, 331–339
41. DeForge, L. E., Kenney, J. S., Jones, M. L., Warren, J. S., and Remick, D. G. (1992) Biphasic production of IL-8 in lipopolysaccharide (LPS)-stimulated human whole blood. Separation of LPS- and cytokine-stimulated components using anti-tumor necrosis factor and anti-IL-1 antibodies. *J. Immunol.* **148**, 2133–2141
42. Ternowitz, T., Fogh, K., and Kragballe, K. (1988) 15-Hydroxyeicosatetraenoic acid (15-HETE) specifically inhibits LTB₄-induced chemotaxis of human neutrophils. *Skin Pharmacol.* **1**, 93–99
43. Huang, Z. H., Bates, E. J., Ferrante, J. V., Hii, C. S., Poulos, A., Robinson, B. S., and Ferrante, A. (1997) Inhibition of stimulus-induced endothelial cell intercellular adhesion molecule-1, E-selectin, and vascular cellular adhesion molecule-1 expression by arachidonic acid and its hydroxy and hydroperoxy derivatives. *Circ. Res.* **80**, 149–158
44. Boisvert, W. A., Curtiss, L. K., and Terkeltaub, R. A. (2000) Interleukin-8 and its receptor CXCR2 in atherosclerosis. *Immunol. Res.* **21**, 129–137
45. FitzGerald, G. A., Oates, J. A., Hawiger, J., Maas, R. L., Roberts II, L. J., Lawson, J. A., and Brash, A. R. (1983) Endogenous biosynthesis of prostacyclin and thromboxane and platelet function during chronic administration of aspirin in man. *J. Clin. Invest.* **71**, 676–688
46. Smith, J. P., Haddad, E. V., Taylor, M. B., Oram, D., Blakemore, D., Chen, Q., Boutaud, O., and Oates, J. A. (2012) Suboptimal inhibition of platelet cyclooxygenase-1 by aspirin in metabolic syndrome. *Hypertension* **59**, 719–725

Received for publication May 21, 2015.

Accepted for publication July 6, 2015.

Inherited human group IVA cytosolic phospholipase A₂ deficiency abolishes platelet, endothelial, and leucocyte eicosanoid generation

Nicholas S. Kirkby, Daniel M. Reed, Matthew L. Edin, et al.

FASEB J 2015 29: 4568-4578 originally published online July 16, 2015

Access the most recent version at doi:[10.1096/fj.15-275065](https://doi.org/10.1096/fj.15-275065)

- Supplemental Material** <http://www.fasebj.org/content/suppl/2015/07/16/fj.15-275065.DC1>
- References** This article cites 46 articles, 23 of which can be accessed free at:
<http://www.fasebj.org/content/29/11/4568.full.html#ref-list-1>
- Subscriptions** Information about subscribing to *The FASEB Journal* is online at
<http://www.faseb.org/The-FASEB-Journal/Librarian-s-Resources.aspx>
- Permissions** Submit copyright permission requests at:
<http://www.fasebj.org/site/misc/copyright.xhtml>
- Email Alerts** Receive free email alerts when new an article cites this article - sign up at
<http://www.fasebj.org/cgi/alerts>
-

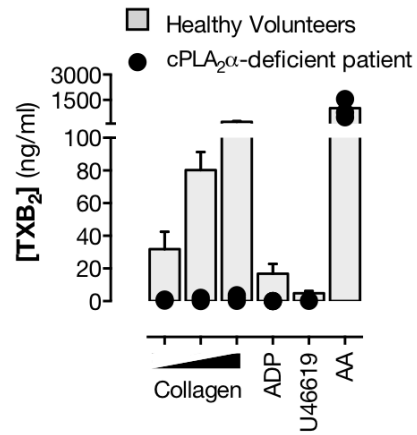


Figure S1. Effect of cPLA₂ deficiency on platelet TXA₂ release to a range of agonists. TXA₂ release (measured as TXB₂) by platelet-rich plasma from healthy volunteer or cPLA₂α-deficient patients (patient B + patient S) blood in response to stimulation by collagen (0.3-3 μg/ml), ADP (5 μmol/L), U46619 (10 μmol/L) or arachidonic acid (1mmol/L). n=4 (healthy volunteers), n=2 (patient).

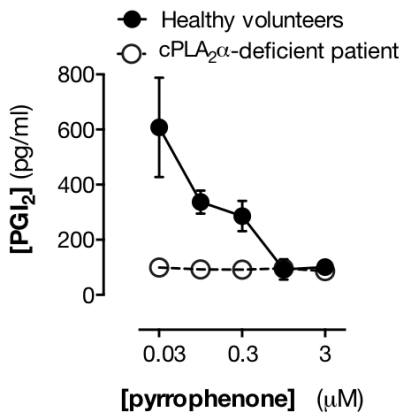


Figure S2. Effect of cPLA₂ inhibition on prostacyclin release by endothelial cells. Prostacyclin release (measured as 6-keto-PGF_{1α}) by endothelial cells isolation from healthy volunteer or cPLA₂α-deficient patient (patient S) blood in response to A23187 stimulation in the presence of increasing concentrations of the cPLA₂ inhibitor, pyrrophenone. n=4 (healthy volunteers), n=1 (patient).

Mediator (ng/ml)	Vehicle (PBS)		Collagen (30µg/ml)		TRAP6 (30µM)		A23187 (50µM)	
	Control	cPLA ₂ α deficient	Control	cPLA ₂ α deficient	Control	cPLA ₂ α deficient	Control	cPLA ₂ α deficient
6ketoPGF _{1α}	0.0 ± 0.0	0.0	0.1 ± 0.0	0.1	0.1 ± 0.0	0.0	0.2 ± 0.1	0.0
TXB ₂	0.1 ± 0.0	0.0	17.6 ± 3.1	0.1	24.1 ± 7.5	0.0	38.4 ± 4.4	0.1
PGE ₂	0.0 ± 0.0	0.0	1.1 ± 0.2	0.0	2.0 ± 1.0	0.0	4.8 ± 1.2	0.0
PGD ₂	0.0 ± 0.0	0.0	0.8 ± 0.2	0.0	1.1 ± 0.5	0.0	2.7 ± 0.6	0.0
8isoPGF _{2α}	0.0 ± 0.0	0.0	0.1 ± 0.0	0.0	0.1 ± 0.0	0.0	0.1 ± 0.0	0.0
9,12,13-THOME	1.2 ± 0.9	0.2	0.3 ± 0.1	0.2	1.2 ± 1.0	0.2	1.2 ± 0.9	0.2
9,10,13-THOME	0.1 ± 0.0	0.1	0.1 ± 0.0	0.1	0.1 ± 0.0	0.0	0.2 ± 0.0	0.0
12,13-DHOME	13.8 ± 4.6	2.5	15.0 ± 5.7	3.0	15.8 ± 5.5	1.9	17.3 ± 7.0	1.8
9,10-DHOME	7.6 ± 2.4	2.4	8.1 ± 2.7	2.7	8.8 ± 3.1	1.7	9.8 ± 3.2	1.2
19,20-DiHDPA	0.9 ± 0.2	1.2	0.9 ± 0.2	1.3	1.0 ± 0.2	0.9	0.9 ± 0.2	0.8
14,15-DHET	0.5 ± 0.0	0.3	0.5 ± 0.0	0.3	0.5 ± 0.0	0.2	0.5 ± 0.0	0.2
11,12-DHET	0.3 ± 0.0	0.1	0.3 ± 0.0	0.1	0.3 ± 0.0	0.1	0.3 ± 0.0	0.1
8,9-DHET	0.1 ± 0.0	0.1	0.1 ± 0.0	0.1	0.2 ± 0.0	0.1	0.2 ± 0.0	0.1
5,6-DHET	0.1 ± 0.0	0.1	0.1 ± 0.0	0.1	0.1 ± 0.0	0.0	0.2 ± 0.1	0.0
13-HODE	1.8 ± 0.3	0.6	2.0 ± 0.3	0.7	2.0 ± 0.3	0.6	3.4 ± 0.6	0.6
9-HODE	0.8 ± 0.1	0.2	1.0 ± 0.1	0.2	1.1 ± 0.2	0.1	1.5 ± 0.1	0.1
20-HETE	0.3 ± 0.1	0.1	0.3 ± 0.1	0.0	0.3 ± 0.1	0.1	0.4 ± 0.1	0.0
19-HETE	0.3 ± 0.0	0.2	0.3 ± 0.0	0.2	0.3 ± 0.0	0.1	0.3 ± 0.0	0.1
15-HETE	0.7 ± 0.1	0.5	10.5 ± 2.2	0.6	14.2 ± 4.9	0.4	34.5 ± 2.5	0.7
12-HETE	8 ± 5	2.8	235 ± 64	6.0	43 ± 19	2.9	838 ± 90	25.7
11-HETE	0.1 ± 0.0	0.1	4.1 ± 0.8	0.1	6.2 ± 2.4	0.0	11.3 ± 1.3	0.1
5-HETE	0.7 ± 0.2	0.4	1.3 ± 0.3	0.5	1.2 ± 0.2	0.3	217 ± 39	0.3
12,13-EpOME	2.9 ± 0.6	0.8	2.6 ± 0.4	0.8	2.8 ± 0.5	0.8	2.8 ± 0.5	1.0
9,10-EpOME	0.3 ± 0.1	0.2	0.3 ± 0.0	0.2	0.3 ± 0.1	0.2	0.4 ± 0.1	0.3
19,20-EpDPE	0.4 ± 0.1	0.8	0.5 ± 0.1	1.0	0.5 ± 0.1	0.8	0.6 ± 0.1	0.7
17,18-EpETE	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0
14,15-EET	0.1 ± 0.0	0.0	0.1 ± 0.0	0.1	0.1 ± 0.0	0.0	0.4 ± 0.0	0.1
11,12-EET	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0	0.2 ± 0.0	0.0
8,9-EET	0.1 ± 0.0	0.0	0.1 ± 0.0	0.1	0.1 ± 0.0	0.0	0.2 ± 0.0	0.0
5,6-EET	0.4 ± 0.1	0.2	0.6 ± 0.1	0.3	0.7 ± 0.1	0.2	2.4 ± 0.1	0.4

Table S1. Contribution of cPLA₂α to eicosanoid synthesis in whole blood stimulated with platelet activators. Total eicosanoid levels in whole blood from healthy volunteers ('control') or from patient S lacking cPLA₂α ('cPLA₂α deficient') stimulated with vehicle (PBS), collagen (30µg/ml), TRAP-6 amide (30µM) or Ca²⁺ ionophore A23187 (30µM). n=4 (healthy volunteers), n=1 (patient).

Mediator (ng/ml)	Vehicle (PBS)		AA (1mM)	
	Control	cPLA ₂ α deficient	Control	cPLA ₂ α deficient
6ketoPGF ₁ α	0.0 \pm 0.0	0.0	2.5 \pm 0.6	0.5
TXB ₂	0.1 \pm 0.0	0.0	168.0 \pm 12.1	83.3
PGE ₂	0.0 \pm 0.0	0.0	144.7 \pm 38.8	30.1
PGD ₂	0.0 \pm 0.0	0.0	143.1 \pm 46.5	35.0
8isoPGF ₂ α	0.0 \pm 0.0	0.0	14.3 \pm 5.9	0.8
9,12,13-THOME	1.2 \pm 0.9	0.2	1.4 \pm 0.5	0.2
9,10,13-THOME	0.1 \pm 0.0	0.1	0.9 \pm 0.4	0.1
12,13-DHOME	13.8 \pm 4.6	2.5	18.0 \pm 2.9	2.4
9,10-DHOME	7.6 \pm 2.4	2.4	17.2 \pm 7.2	2.2
19,20-DiHDPA	0.9 \pm 0.2	1.2	1.0 \pm 0.1	1.1
14,15-DHET	0.5 \pm 0.0	0.3	23.8 \pm 6.0	6.7
11,12-DHET	0.3 \pm 0.0	0.1	14.1 \pm 2.9	6.4
8,9-DHET	0.1 \pm 0.0	0.1	8.0 \pm 2.7	2.1
5,6-DHET	0.1 \pm 0.0	0.1	41.9 \pm 16.7	11.6
13-HODE	1.8 \pm 0.3	0.6	13.3 \pm 5.3	0.7
9-HODE	0.8 \pm 0.1	0.2	12.4 \pm 5.6	0.4
20-HETE	0.3 \pm 0.1	0.1	6.3 \pm 0.4	5.1
19-HETE	0.3 \pm 0.0	0.2	9.6 \pm 3.8	1.7
15-HETE	0.7 \pm 0.1	0.5	1945.3 \pm 484.6	813.3
12-HETE	8.5 \pm 5.1	2.8	1448.3 \pm 228.7	1060.0
11-HETE	0.1 \pm 0.0	0.1	301.1 \pm 83.3	129.3
5-HETE	0.7 \pm 0.2	0.4	520.0 \pm 148.3	298.7
12,13-EpOME	2.9 \pm 0.6	0.8	4.1 \pm 0.8	0.7
9,10-EpOME	0.3 \pm 0.1	0.2	1.2 \pm 0.5	0.2
19,20-EpDPE	0.4 \pm 0.1	0.8	1.1 \pm 0.5	1.0
17,18-EpETE	0.0 \pm 0.0	0.0	0.2 \pm 0.1	0.1
14,15-EET	0.1 \pm 0.0	0.0	149.8 \pm 36.8	66.3
11,12-EET	0.0 \pm 0.0	0.0	113.5 \pm 34.9	38.7
8,9-EET	0.1 \pm 0.0	0.0	115.9 \pm 42.2	40.9
5,6-EET	0.4 \pm 0.1	0.2	471.8 \pm 201.7	200.3

Table S2. Contribution of cPLA₂ α to eicosanoid synthesis in whole blood stimulated with exogenous arachidonic acid (AA). Total eicosanoid levels in whole blood from healthy volunteers ('control') or from patient S, lacking cPLA₂ α ('cPLA₂ α deficient') stimulated with vehicle (PBS) or exogenous AA (1mM). n=4 (healthy volunteers), n=1 (patient).

Mediator (ng/ml)	Vehicle (DMEM)		LPS (10µg/ml)		LPS + AA (1mM)	
	Control	cPLA ₂ α deficient	Control	cPLA ₂ α deficient	Control	cPLA ₂ α deficient
6ketoPGF _{1α}	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0	0.3 ± 0.0	0.1
TXB ₂	0.6 ± 0.3	0.0	2.5 ± 0.4	0.3	128.1 ± 12.9	35.2
PGE ₂	5.7 ± 5.3	0.1	26.2 ± 6.7	2.0	161.6 ± 25.3	55.4
PGD ₂	0.1 ± 0.0	0.0	0.2 ± 0.1	0.1	140.9 ± 5.2	105.1
8isoPGF _{2α}	0.0 ± 0.0	0.0	0.2 ± 0.0	0.0	2.7 ± 0.2	1.4
9,12,13-THOME	0.7 ± 0.5	0.2	0.7 ± 0.4	0.3	0.7 ± 0.4	0.3
9,10,13-THOME	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0	0.1 ± 0.0	0.1
12,13-DHOME	21.8 ± 5.6	3.0	19.6 ± 3.8	3.9	12.0 ± 6.3	3.6
9,10-DHOME	9.6 ± 3.2	2.5	8.5 ± 3.0	3.6	5.1 ± 3.0	0.9
19,20-DiHDPA	2.2 ± 0.5	2.9	2.1 ± 0.2	2.8	1.4 ± 0.6	1.9
14,15-DHET	0.7 ± 0.1	0.6	0.8 ± 0.1	0.5	8.9 ± 6.2	5.7
11,12-DHET	1.1 ± 0.1	0.5	0.8 ± 0.1	0.7	7.7 ± 5.2	6.2
8,9-DHET	1.2 ± 0.1	0.9	1.3 ± 0.1	0.6	7.4 ± 3.3	5.1
5,6-DHET	1.3 ± 0.2	0.5	1.3 ± 0.1	0.8	50.1 ± 23.9	23.9
13-HODE	16.8 ± 3.8	7.3	30.9 ± 9.7	20.7	12.8 ± 4.3	19.0
9-HODE	4.9 ± 1.1	1.6	5.3 ± 1.5	4.1	3.8 ± 1.6	0.5
20-HETE	0.8 ± 0.4	0.0	0.6 ± 0.2	0.0	9.9 ± 3.9	9.6
19-HETE	0.5 ± 0.3	0.0	0.8 ± 0.4	1.2	18.1 ± 6.3	18.8
15-HETE	11.9 ± 4.7	5.9	34.8 ± 8.6	12.5	2787 ± 1116	3386.7
12-HETE	215.0 ± 50.2	71.5	229.2 ± 70.6	98.1	1758 ± 1029	2082.0
11-HETE	3.6 ± 2.1	0.7	8.8 ± 4.2	2.5	400 ± 213.9	595.3
5-HETE	4.5 ± 2.0	2.3	4.8 ± 1.0	2.4	797.1 ± 359.8	1045.3
12,13-EpOME	2.8 ± 0.5	1.7	2.7 ± 0.7	1.9	1.8 ± 0.5	1.8
9,10-EpOME	0.6 ± 0.1	0.5	0.6 ± 0.2	0.6	0.4 ± 0.1	0.6
19,20-EpDPE	1.3 ± 0.2	4.4	1.6 ± 0.2	4.8	1.4 ± 0.5	5.2
17,18-EpETE	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0
14,15-EET	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0	63.3 ± 18.1	61.0
11,12-EET	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0	25.8 ± 9.1	33.9
8,9-EET	0.0 ± 0.0	0.0	0.0 ± 0.0	0.0	32.2 ± 11.6	40.3
5,6-EET	0.3 ± 0.3	0.0	0.5 ± 0.3	0.0	119.1 ± 36.6	125.8

Table S3. Contribution of cPLA₂α to eicosanoid synthesis in whole blood stimulated with LPS. Total eicosanoid levels in whole blood from healthy volunteers ('control') or from patient S lacking cPLA₂α ('cPLA₂α deficient') stimulated for 18 hours with vehicle (DMEM) or LPS (10µg/ml) with or without addition of exogenous arachidonic acid (AA; 1mM) for the final 30 mins. n=4 (healthy volunteers), n=1 (patient).