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FAK-heterozygous mice display enhanced tumour angiogenesis

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Abstract

Genetic ablation of endothelial Focal Adhesion Kinase (FAK) can inhibit pathological angiogenesis, suggesting that loss of endothelial FAK is sufficient to reduce neovascularisation. Here we show that reduced stromal-FAK expression in FAK-heterozygous mice unexpectedly enhances both B16F0 and CMT19T tumour growth and angiogenesis. We further demonstrate that cell proliferation and microvessel sprouting, but not migration, are increased in serum-stimulated FAK-heterozygous endothelial cells. FAK-heterozygous endothelial cells display an imbalance in FAK phosphorylation at pY397 and pY861 without changes in Pyk2 or Erk1/2 activity. By contrast, serum-stimulated phosphorylation of Akt is enhanced in FAK-heterozygous endothelial cells and these cells are more sensitive to Akt inhibition. Additionally, low doses of a pharmacological FAK inhibitor, although too low to affect FAK autophosphorylation *in vitro*, can enhance angiogenesis *ex vivo* and tumor growth *in vivo*. Our results highlight a potential novel role for FAK as a non-linear, dose-dependent regulator of angiogenesis where heterozygous levels of FAK enhance angiogenesis.

Introduction

The non-receptor tyrosine kinase, Focal Adhesion Kinase (FAK), can signal downstream of integrins and growth factor receptors¹. FAK is expressed ubiquitously and plays essential roles in a number of cellular processes, including proliferation and migration, which are important angiogenic processes. FAK is autophosphorylated at Y397 following integrin

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Author contributions VK and TL designed, performed and analysed experiments. DTJ performed proliferation assays, LER and DML assisted with signalling experiments, MB did the 3D tube assay, ARR did the VEGF-stimulated signalling experiments, SDR and BT helped with *in vivo* experiments. GB and AMR performed 2D tubulogenesis assays and MP carried out migration assays. IRH provided intellectual input. VK and KMHD intellectually conceived and supervised the research and wrote the manuscript. All authors had final approval of the submitted manuscript.

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activation and additionally this site acts to recruit members of the Src-family kinases (SFKs) ²⁻⁴. The proximity of SFKs to FAK is thought to result in its phosphorylation at other sites, including Y861, and this may cause activation of downstream effectors including Erk and Akt^{5,6}. Previous reports have highlighted that the kinase activity, and phosphorylation of FAK at Y397 and Y861, are important regulators of angiogenic responses *in vitro*^{3,7-9}. Nonetheless, how the activity of these sites affect the tumour stromal influence on tumorigenesis has never been addressed.

Genetic studies also have indicated a role for FAK in angiogenesis *in vivo*. For example, the global deletion of FAK results in an early embryonic lethal phenotype with defects in the vascular system¹⁰. Constitutive FAK-deletion specifically in vascular endothelial cells (ECs) caused embryos to die between E10.5 and E13.5, which coincides with the onset of embryonic vascular system development ^{11,12}, while over-expression of FAK in ECs has led to enhanced blood vessel density during various angiogenic processes¹³. Although one report suggests that the inducible deletion of endothelial-FAK in End-SCL-Cre^{ERT} adult mice failed to affect adult angiogenesis, this apparently was because of induced compensation by a related family member, Pyk2¹⁴. We have shown previously that endothelial-specific loss of FAK in Pdgfb-iCreER^{T2};FAK fl/fl mice inhibit tumour angiogenesis¹⁵. Together these data suggest that loss of FAK can be sufficient to inhibit pathological angiogenesis. Conversely, although FAK-knockdown inhibited cell migration and tube formation *in vitro*¹⁰, others have shown that FAK-phosphorylation¹⁶ and levels of expression are decreased during the early stage of an *in vitro* model of angiogenesis¹⁷. Such, apparently contradicting results suggest that FAK may have non-linear dose dependent roles in angiogenesis with varied FAK levels differentially affecting angiogenesis. Indeed, the embryonic development of FAK-heterozygous mice is severely retarded, but these defects are overcome by birth and FAK-heterozygous mice are born at normal Mendelian ratios with no gross abnormalities¹⁸. In addition, although FAK is expressed ubiquitously, its expression and activity levels can vary between cells and even within the same cell type^{4,19}, suggesting that examining the effects of partial loss of endothelial FAK may provide us with more physiologically relevant information about its roles *in vivo*.

Here, we have tested the influence that subtle changes in stromal-FAK levels may have on tumour growth and angiogenesis, by utilising FAK-heterozygous mice²⁰. FAK heterozygous mice display half the normal levels of FAK protein. We show that FAK-heterozygous mice exhibit: (1) enhanced B16F0 and CMT19T tumour growth, (2) increased formation of perfused tumour blood vessels and (3) elevated neovascularisation *in vivo*. At the molecular level, we have determined that the ratio of FAK phosphorylation at Y861:Y397 is misbalanced and that the relative phosphorylation of Akt is elevated in FAK-heterozygous ECs. In agreement with this, FAK-heterozygous ECs exhibit increased sensitivity to Akt inhibition. Lastly, low doses of the FAK inhibitor, PF-573228, can phenocopy the responses in FAK-heterozygotes. Taken together these data suggest that FAK has a non-linear dose dependent effect in angiogenesis with partial loss of FAK function being sufficient to enhance pathological angiogenic responses.

Results

Tumour growth and angiogenesis in FAK-heterozygous mice

Tumour growth has been associated with enhanced tumour cell FAK-expression, but the role of stromal-FAK is less well understood. Although some data indicate that total loss of FAK can inhibit angiogenic processes ^{10-12,15}, others suggest that it may play a non-linear dose dependent role in neovascularising events ^{4,16,17,19}. Using FAK-heterozygous mice ²⁰ we investigated the effect of partial loss of stromal FAK on tumour growth and angiogenesis. These mice carry only one FAK allele and western blot analysis confirmed that FAK protein

expression was reduced to 50% in FAK-heterozygous tissues when compared with wild-type (Supplementary Fig. S1). To assess whether decreased FAK levels play a role in tumour growth, we injected syngeneic B16F0 melanoma cells or CMT19T lung tumour cells into the flanks of age- and sex-matched FAK-heterozygous mice and wild-type littermates. Two weeks post-inoculation, both B16F0 and the CMT19T tumours from FAK-heterozygous animals were significantly larger than those from wild-type controls (Fig. 1A). Indeed, tumour growth was increased significantly in FAK-heterozygous mice when compared with wild-type controls over time (Supplementary Fig. S2). These data indicate that partial loss of FAK in the tumour stroma is sufficient to enhance tumour growth.

To examine whether the elevated tumour size in FAK-heterozygous mice correlated with any change in tumour angiogenesis midline sections of size-matched, age-matched tumours were analysed. Double immunostaining for PECAM and FAK, revealed that endothelial FAK levels were decreased significantly in tumour blood vessels in FAK-heterozygous mice when compared with wild-type controls and that the levels varied widely in the wild-type mice (Supplementary Fig. S3). In contrast, FAK levels were indistinguishable in the tumour cell compartment between both genotypes suggesting that stromal FAK did not affect tumour cell FAK levels (Supplementary Fig. S4). Tumour blood vessel density was defined by counting the total number of blood vessels across entire midline sections and dividing by the tumour section area. Results revealed that blood vessel density was increased in both B16F0 and CMT19T tumours grown in FAK-heterozygous mice when compared with control mice (Fig. 1B). Furthermore, the number of functional, PE-PECAM antibody perfused, blood vessels was also increased in B16F0 tumours grown in FAK-heterozygous mice (Fig. 1C). Enhanced tumour blood vessel perfusion has been associated with increased tumour growth previously^{24,25}. These data correlated with a significant increase in the number of BrdU-positive vessels in B16F0 tumours from FAK-heterozygous mice when compared with those from control mice (Fig. 1D). Importantly the blood vessel density in unchallenged skin was not changed between genotypes, suggesting that the enhanced tumour angiogenesis was not due to elevated blood vessel numbers in untreated FAK-heterozygous mice (Supplementary Fig. S5). Taken together these data point towards a role for stromal FAK in the regulation of angiogenesis.

Angiogenesis can be affected by various cell types other than ECs. For example, the formation of stable angiogenic vessels is associated with the recruitment of pericytes. Our data showed that the number of tumour blood vessels associated with α -smooth muscle actin-positive pericytes was similar between the two genotypes suggesting that vessel maturation was not affected by FAK-heterozygosity (Supplementary Fig. S6). Any changes in tumour vascularity did not appear to correlate with changes in tumour hypoxia, apoptosis, or levels of serum Vascular Endothelial Growth Factor (VEGF) between the two genotypes (Supplementary Fig. S7, S8 and S9). Although, changes in inflammatory cell infiltrate have been reported to enhance neovascularisation^{26,27} no significant differences in the relative level of macrophage (Gr1+ or F4/80+) or leukocyte (CD45+) infiltration was observed between tumours from FAK-heterozygous or control animals (Supplementary Fig. S10). Overall, although we cannot rule out a non-cell autonomous affect on the enhanced angiogenesis observed in FAK-heterozygous mice we have no evidence for changes in any of the cell types tested other than in ECs.

VEGF is a major pro-angiogenic growth factor. To determine whether the changes observed during tumour angiogenesis could be seen in a tumour free environment, VEGF impregnated sponges were implanted subcutaneously into FAK-heterozygous and wild-type mice to stimulate blood vessel growth. Results showed that FAK-heterozygous mice had a higher infiltration of blood vessels into sponges with or without VEGF when compared with similarly treated wild-type controls (Fig. 2). Indeed, there was no difference in the fold-

increase of *in vivo* blood vessel infiltration in wild-type mice when compared with FAK-heterozygous mice suggesting that the enhanced angiogenic response was not VEGF-specific. Thus our data suggested that *in vivo* VEGF-mediated responses were not enhanced in FAK-heterozygous mice *per se*. This notion was corroborated by Western blot analysis for VEGF-receptor 2 protein expression and downstream signalling activity in purified ECs. Our results showed that although FAK-heterozygous ECs displayed approximately half the levels of total FAK, as expected, there was no change in VEGF-stimulated levels of VEGF-receptor 2, phospho-VEGF-receptor 2, or downstream effectors such as p130Cas, phospho-p130Cas, total Erk1/2 or phospho-Erk1/2. In short our data indicated that partial loss of FAK in FAK-heterozygous ECs does not affect VEGF-mediated signalling specifically (Supplementary Fig. S11). In addition, VEGF-stimulation did not affect the migration speed of FAK-heterozygous cells (Supplementary Fig. S12). Therefore, although beyond the scope of the present study, other angiogenic factors may play a role in triggering the enhanced angiogenic response in FAK-heterozygous mice. Thus we focused on the effect of partial loss of FAK on serum-stimulated endothelial activity.

FAK-heterozygosity enhances cell survival and tube formation

In order to determine the cellular mechanisms underlying the enhanced pathological angiogenesis observed in FAK-heterozygous mice, ECs from FAK-heterozygous and control mice were isolated and subjected to a number of cellular assays. As an indication of relative survival, ECs grown in full growth medium were assayed for WST-1 metabolism. Interestingly, FAK-heterozygous ECs demonstrated significantly increased WST-1 metabolism when compared with control cells (Fig. 3A) suggesting that partial loss of FAK enhances endothelial survival. In contrast, using Dunn chamber assays to determine the effect of partial FAK loss on cell migration, no statistically significant differences in serum-mediated migration speed or persistence of cell migration were observed between genotypes (Fig. 3B).

Tube formation is a vital step of angiogenesis. ECs are known to form tubular structures when grown on a 2-dimensional (2D) Matrigel™ substrate²⁸. FAK-heterozygous cells spontaneously formed more tubular structures than wild-type control cells in 2D Matrigel™ assays (Fig. 3C). In addition, in 3-dimensional tubulogenesis assays, FAK-heterozygous ECs cultured on fibrinogen-coated beads, in the presence of full endothelial growth medium and a fibroblast feeder layer, developed significantly higher numbers of tubular sprouts than wild-type cells grown under similar conditions (Fig. 3D). In short, the partial loss of FAK in ECs confers a spontaneous ability to enhance angiogenic responses that is independent of effects on EC migration.

Pyk2 does not compensate FAK-heterozygosity

Changes in FAK activity have been correlated with changes in its levels of phosphorylation³. In addition, apart from FAK, a related family member, Pyk2 has also been reported to be expressed in ECs and has, in some circumstances, been implicated in compensating for FAK¹⁴. We next explored the expression and phosphorylation levels of FAK and Pyk2 in FAK-heterozygous and wild-type ECs. Immunofluorescence analysis of total FAK, pY397-FAK, and pY861-FAK showed that, although the intensity levels of total FAK and pY397-FAK were decreased in the FAK-heterozygous cells (Fig. 4A), the level of FAK-phosphorylation, specifically at Y861, was similar in FAK-heterozygous and wild-type ECs (Fig. 4A). Furthermore, western blot analysis of wild-type and FAK-heterozygous EC extracts confirmed that FAK levels were reduced in FAK-heterozygous ECs, and revealed a correlation with a decrease in pY397-FAK, but not pY861-FAK levels (Fig. 4B). Consequently, although the ratio of pY397-FAK/total FAK remained the same between the two genotypes, the ratio of pY861-FAK/total FAK displayed a two fold increase in FAK-

heterozygous cells when compared with wild-type ECs. Moreover, a reduction of FAK levels in ECs did not cause any changes in the expression or phosphorylation of Pyk2 (Fig. 4B). FAK is also known to play an important role in the dynamic regulation of focal contacts and the organisation of the actin cytoskeleton². Our data revealed that FAK-heterozygosity was not sufficient to alter the organisation of endothelial F-actin bundles (Fig. 5A). In addition, immunofluorescence and western blot studies suggested that paxillin, talin and vinculin levels were unchanged between genotypes (Fig. 5A and B). Taken together, FAK-heterozygosity does not appear to result in overt compensatory mechanisms involving Pyk2 or other focal contact proteins.

Effect of FAK-heterozygosity on Akt signalling

FAK can affect both integrin and growth factor signalling by regulating downstream targets, such as Erk1/2 and Akt^{1,3,6,8}. Our results show that serum-stimulated phosphorylation of Erk1/2 was not significantly altered in FAK-heterozygous ECs when compared with wild-type controls. In contrast, although the absolute expression levels of pAkt did not appear to change between genotypes (Fig 6A), the levels of total Akt were reduced significantly in FAK-heterozygous ECs compared to wild-type ECs (Supplementary Fig. S13). This resulted in a significant increase in the ratio of pAkt/ total Akt in FAK-heterozygous ECs when compared with wild-type controls (Fig. 6A). To establish whether this increase in pAkt/ total Akt ratio was functionally linked to the increased angiogenic activity of FAK-heterozygotes, we tested the affect of Akt inhibition on angiogenic responses *ex vivo*. FAK-heterozygous and wild-type aortic rings were treated with the Akt inhibitor VIII, which selectively inhibits the kinase activity of Akt1/2. High doses (10 μ M range) of the Akt VIII inhibitor were able to inhibit microvessel sprouting in both FAK-heterozygous and wild-type aortic rings. However, the enhanced Akt-sensitivity of FAK-heterozygotes was illustrated by their decreased sprouting response to lower doses (1 μ M range) of the Akt inhibitor that did not affect wild-type aortic rings (Fig. 6B). These data suggest that the elevated Akt activation could contribute to the pro-angiogenic phenotype of FAK-heterozygous mice.

Low-dose FAK inhibition increases tumour growth and angiogenesis

To determine whether inhibition of FAK can enhance microvessel sprouting, we tested the effect of treatment with various concentrations of FAK inhibitor PF-573228²⁹ in *ex vivo* aortic ring assays. Interestingly, although high concentrations (1 μ M) of PF-573228 reduced microvessel sprouting from aortic rings, low concentrations (0.01 μ M) of the inhibitor enhanced the numbers of microvessel sprouts significantly (Supplementary Fig. S14), phenocopying the enhanced angiogenic activity observed in FAK-heterozygous mice. PF-573228 has not been reported to have been used previously in *in vivo* experiments. However, the related PF-562271 FAK inhibitor, that display similar inhibitory activity in *in vitro* functional assays²⁹⁻³¹, has been shown to inhibit tumour growth in mice at 50mg/kg dose³⁰⁻³². Therefore, we treated wild-type mice bearing B16F0 tumours with 12.5mg/Kg PF573228 – a suboptimal dose of this inhibitor. Tumour growth was enhanced in mice treated with 12.5mg/Kg PF573228 compared to placebo treated mice (Supplementary Fig. S15). Further analysis showed that tumour blood vessel density was also enhanced significantly in mice treated with 12.5mg/Kg PF573228 (Supplementary Fig. S15). Taken together these findings indicate that FAK can have a non-linear dose dependent function in regulating tumour growth and angiogenesis.

Discussion

Although FAK is ubiquitously expressed, its actual levels of expression can vary within tissues and even within individual cell types^{4,19}. Indeed here we have observed a wide range of FAK expression levels in ECs from wild-type mice. These observations lead to the

question of how different levels of FAK might regulate biological outcome - a relatively unexplored field of research. Some reports have suggested that an increase in FAK within the whole tumour correlates with tumour progression, but in contrast high levels of FAK do not always predict outcome^{3,32-39}. Given the emerging importance of the tumour stroma in regulation of cancer progression, we speculate that the influence of stromal-FAK on tumour growth may be important. Indeed with the development of FAK inhibitors as potential anti-cancer drugs⁴⁰, and the fact that complete pharmacological inhibition of FAK may be difficult to maintain *in vivo*, elucidating the effects of different levels of stromal-FAK in cancer progression becomes a priority⁴¹.

Our data show that stromal-FAK heterozygosity is sufficient to enhance tumour growth and tumour angiogenesis. This does not correlate with any obvious changes in endothelial expression of focal adhesion proteins or cellular migration, but does correlate with enhanced EC proliferation and relative Akt activity levels. Thus our results establish a new role for FAK in the regulation of angiogenesis.

Previous genetic ablation work has highlighted an apparent requirement for FAK during developmental and postnatal angiogenesis^{10-12,15}. However, in adult organisms the total deletion of FAK is unlikely to be of physiological relevance. Thus, studying the effect of reduced levels of FAK may provide us with new information about the roles of this signalling effector. Our data suggest that heterozygous levels of FAK can elevate angiogenic responses and our results are corroborated by other reports^{16,17}. For example, FAK-heterozygous embryos show retarded embryonic development but, by birth, appear normal¹⁸, suggesting that heterozygous levels of FAK might allow for some compensation that overcomes the embryonic defect. Additionally, *in vitro* studies have indicated that, during the first, but not latter, phases of angiogenesis, FAK levels are reduced, suggesting that FAK may have a non-linear dose-dependent effect in angiogenic processes *in vitro*^{16,17}.

The tumour stroma consists of various cell types including endothelial cells, smooth muscle cells/pericytes and inflammatory cells. It is conceivable that the enhanced angiogenic responses in FAK-heterozygous mice may be due to the up-regulation of some or all of these non-EC types. However we found no evidence for the increase in the numbers of mural cells or immune infiltrating cells in tumours grown in FAK-heterozygous mice or the number of vessels with associated pericytes. Thus, although we cannot rule out a non-cell autonomous affect here we have focused on the endothelial compartment where we have observed FAK-dependent changes in molecular profiles and EC function.

Our data demonstrate that FAK-heterozygosity is sufficient to affect EC cells, by increasing endothelial proliferation and sprout formation, but without affecting overall migration speed of these cells. Pyk2, a member of the FAK family, has been reported previously to compensate for the loss of FAK¹⁴. However we did not observe any changes in Pyk2 levels or phosphorylation in the FAK-heterozygous ECs. In contrast, in FAK-heterozygous ECs, although phosphorylation of FAK at Y397 is reduced, the levels of pY861-FAK surprisingly, remain high. The ratio of pY397/ total FAK remains the same whereas the ratio of pY861/total FAK is increased. Although it has been proposed that phosphorylation of Y397 is required for conformational changes that trigger the phosphorylation of Y861^{5,6,42}, other studies have reported enhanced phosphorylation of Y861-FAK without alterations in pY397 levels or FAK activity, suggesting that the uncoupling of these two FAK phosphorylation sites can occur^{7,43-45}. In addition, FAK-heterozygous ECs have an increase in both basal and serum-stimulated activation of Akt. Previous genetic studies have indicated a pro-angiogenic role for Akt⁴⁶. More importantly, it has been reported that sustained activation of Akt in ECs resulted in enhanced blood vessel size and permeability and increased tumour growth⁴⁷. Our findings, although correlative, suggest that the

increased activity of Akt in FAK-heterozygous ECs could be responsible for the enhanced cellular proliferation and cause elevated angiogenic responses in the FAK-heterozygous mice. In the *ex vivo* aortic ring assay, a sub-optimal dose of the Akt VIII inhibitor had a greater impact on the angiogenic responses of FAK-heterozygotes when compared with wild-types, suggesting increased sensitivity to Akt inhibition. A recent report demonstrated that FAK can bind Akt1 independently of the phosphorylation status of Y397 and that partial depletion of FAK, using siRNA, stimulated basal phosphorylation of Akt⁴⁸. Thus we speculate that there may be a link between reduced FAK expression levels, a misbalance of FAK-phosphorylation and Akt activity that may enhance angiogenic responses.

The mechanisms by which endothelial FAK levels control survival are not clear. It is tempting to speculate that since FAK has both kinase and non-kinase functions⁴⁹, and the non kinase-adaptor functions of FAK can regulate cellular proliferation via p53 without affecting cell migration^{18,50}, that heterozygous FAK expression may bias for such scaffolding FAK functions. FAK can also regulate internalisation of surface molecules and affect cell survival⁵¹. Thus, we postulate that signalling downstream of FAK, in FAK-heterozygous cells, regulates the expression or trafficking of pro-angiogenic cell surface markers, thereby affecting survival but not migration of these cells. Lastly, other studies have shown that loss of FAK expression or function can induce a change in Src induced autophagy and thus enhance cell viability⁵². Although beyond the scope of this study it would be of interest to test these ideas in future experiments.

The enhanced angiogenic affect of stromal FAK-heterozygosity raises the possibility that low doses of FAK inhibitors may phenocopy the response in FAK-heterozygous mice. Our data indicate that this indeed maybe the case since low doses of FAK inhibitor PF573228 enhanced microvessel sprouting *ex vivo*, although 0.01 μ M PF573228 does not appear to affect autophosphorylation of FAK at Y397 *in vitro* (unpublished data). Furthermore low doses of PF573228 increased tumour growth and angiogenesis. We have no evidence that such low doses prevail when such inhibitors are given at the maximal tolerated doses, where they have been shown to inhibit tumour growth and angiogenic effects³⁰⁻³², or indeed how low doses may affect the tumour cells themselves. Off target effects of this inhibitor cannot be excluded or if this applies to other FAK inhibitors. Instead our results reveal new information about the basic biology behind the role of FAK in angiogenesis and provide proof-of-principle that reducing stromal-FAK either by genetic ablation or by pharmacological intervention can elevate angiogenic responses. This work highlights a novel function of FAK in the regulation of angiogenesis that depends on the levels of FAK expression, and not simply its presence.

Methods

Primary cell cultures

Primary mouse lung ECs were isolated and cultured as described previously²¹ from either FAK-heterozygous or wild-type control mice. Tissue culture plates for all experiments were coated with a mixture of collagen (30 μ g/ml), gelatin (0.1%), and fibronectin (10 μ g/ml).

In vivo tumour growth

Age- and sex-matched FAK-heterozygous²⁰ and wild-type littermate controls on a mixed C57BL/129 background were given a single subcutaneous injection into the flank of either 1×10^6 B16F0 melanoma cells, or 0.5×10^6 CMT19T lung carcinoma cells (ATCC). Tumour dimensions were measured everyday and the tumour growth was determined using the formula length \times width² \times 0.52. 14 days after injection or when tumour reached the

maximum legal size allowed, mice were sacrificed; tumour sizes measured, and tumour samples were either snap-frozen or fixed in 4% paraformaldehyde for histological analysis.

Immunohistochemical analysis

Tumours were bisected and midline sections immunostained for endomucin or PECAM to detect blood vessels ECs and smooth muscle actin conjugated to Cy3 to detect smooth muscle cells as described²². Blood vessel density was calculated by counting the number of blood vessels per mm² of midline section. Smooth muscle cell coverage was determined by calculating the ratio of smooth muscle cell positive blood vessels over total blood vessels¹⁵. Perfused blood vessels were detected by intravenously injection of Phycoerythrin-labelled PECAM antibody (100µg) 15 min before B16F0 tumour excision. The percentage of perfused/functional blood vessels was calculated by the ratio of PE-PECAM-positive perfused vessels over the total blood vessels, detected by staining for endomucin across whole tumour sections. As a control, transverse sections of unchallenged skin were also immunostained for endomucin or PECAM and blood vessel density was calculated by counting the number of blood vessels per mm² of skin dermis/ section. To examine tumour ECs proliferation mice received an intraperitoneal injection of 200µl BrdU (5mg/ml) and 30 minutes later were killed and B16F0 tumours were dissected and processed for histological analysis. Double staining for endomucin and BrdU incorporation was carried out, and endothelial proliferation was calculated as the number of BrdU positive cells per perimeter of blood vessels.

Subcutaneous tumour free angiogenesis assay

Briefly, two autoclaved synthetic sponges (a kind gift from Daryl Harmon, Caligen Foam Ltd.) of approx. 1cm³ in size were implanted subcutaneously in the flanks of anaesthetised mice. Every 2 days, sponges were injected *in situ* with either 100µl of PBS alone or 100µl of PBS containing 10ng/ml VEGF. After 14 days, sponges were excised and fixed for paraffin embedding. Sections of sponges were immunostained for vWF to identify blood vessels. Briefly, paraffin sections were dewaxed in xylene, hydrated by sequential immersion in 100%, 80%, 70% and 50% ethanol solutions and treated with protein kinase for 15 minutes at 37°C. Sections were blocked with 2% normal goat serum, 1% BSA and 0.1% TritonX-100 and incubated with anti-vWF (AbD Serotec) overnight at 4°C, followed by a Alexa@-488-conjugated secondary antibody (Invitrogen) for 1 hour at room temperature. Blood vessel density was assessed by counting the number of vWF-positive blood vessels/mm² of sponge section. The nuclear marker DAPI was used as a counterstain.

WST-1 assay

Primary mouse lung ECs were seeded at a density of 2,500 cells/well in coated 96-well plates. EC medium was changed every 2 days. At each time point, 1 day, 2 days and 4 days, the cell proliferation reagent, WST-1 (Roche Applied Science) was applied to each well. Following 3 hours incubation at 37°C, the absorbance of the samples was measured at 450nm using an Anthos 2001 microplate reader. Wells without cells but containing complete medium and WST-1 acted as baseline absorbance controls. Results were normalised to data from day 1.

Cell migration Assay

Primary ECs grown on glass coverslips were serum starved for 4 hours in Opti-MEM™. The coverslips were then placed 'face-down' onto Dunn chambers that contained Opti-MEM™ within the central chamber, and Opti-MEM™ with serum or VEGF in the outer chamber. Cells were then imaged at the 'bridge' between the two chambers, and tracked to determine speed and persistence of cellular movement.

2-Dimensional tubulogenesis assay

Primary mouse lung ECs were serum starved for 4 hours in Opti-MEM™. Cells were then detached from the plate and resuspended in Opti-MEM™ (supplemented with 1% FCS). 50,000 cells were seeded onto solid Matrigel™ (3 wells per condition) in a 48-well plate. Images of the tubes were recorded using a phase contrast 10x objective lens at 7 hours after seeding. The area captured by the camera was considered the field of view and 3-4 complete fields per well were quantified. The total sum length of tube network per field was quantified.

3-Dimensional endothelial cell sprouting assay

Primary mouse lung ECs were incubated with a suspension of Cytodex™ glass beads (GE Healthcare) in EGM-2 medium (Lonza) at 37°C for 4 hours with agitation every 20 minutes. This cell-bead suspension was then plated into flasks and left overnight. The following day, the coated beads were resuspended gently in 1 mg/ml fibrinogen solution (Invitrogen) with 0.15 units/ml of aprotinin solution (Invitrogen) and transferred to 24-well plates with each well containing a drop of thrombin solution (Invitrogen). Plates were left at room temperature for 15 minutes and then placed at 37°C for 1 hour to allow the combined solution to clot. Finally, mitomycin-inactivated NIH-3T3 fibroblasts were then added to each well and allowed to settle for 3 hours before changing the medium. Cells were fed with EGM-2 medium supplemented with 2% FCS with 30ng/ml VEGF or PBS every two days and sprouts counted daily from day 3 to day 6.

Immunofluorescence of cultured cells

Cells grown on 10-mm glass coverslips were washed once in PBS, fixed in acetone for 5 minutes at -20°C, washed in PBS three times and then blocked with 5% normal goat serum for 30 minutes at room temperature. After washing the cells once in PBS, cells were incubated with anti-FAK (1:100, BD-Biosciences cat. no 610087), anti-phospho-FAK (1:100, Invitrogen cat. no 44624, 44626) and anti-talin (1:100, Sigma T3287) antibodies for 1 hour at room temperature. Alternatively, cells were washed once in PBS, fixed in 4% paraformaldehyde for 10 minutes, rinsed in PBS, permeabilised in 0.5% NP40 for 10 minutes and blocked with 0.1% BSA/0.2% Triton-X100 for 10 minutes before incubation with anti-paxillin (1:100 BD-Biosciences 610051), anti-vinculin (1:400 Sigma cat. no V9131) and Rhodamine-phalloidin for F-actin (1: 1000 Invitrogen R415). Cells were then washed with PBS and incubated with Alexa-Fluor®-conjugated secondary antibodies (1:100, Invitrogen) for 45 minutes at room temperature before being mounted on microscope slides using Prolong® Gold anti-fade reagent (Invitrogen). Fluorescence microscopy was performed using a CCD camera Hamamatsu C4742-95 and a Zeiss Axioplan microscope. Identical settings were used to acquire the images in both wild-type and FAK-heterozygous cells. The mean fluorescent intensity of total FAK, pY397-FAK and pY861-FAK in the whole cell area was calculated using the ImageJ™ software.

Western blot analysis

For analysis of protein levels, cells were either lysed directly, or serum starved for 4 hours and then serum-stimulated for 0, 5, 15, 30 and 60 minutes before lysis. Western blot analysis was performed as described in supplementary methods. The anti-, total FAK, p-Pyk2, total-Pyk2, p-Erk, total-Erk, p-Akt (Ser473) and total Akt antibodies were from Cell Signaling Technologies (cat no. 3285, 3291, 3292, 9101, 9102, 9271, 4685, respectively) The phospho-tyrosine FAK antibodies were from Invitrogen (cat. no. 44626, 44624). The anti-talin and anti-vinculin were from Sigma-Aldrich (cat no. T3287 and V9131, respectively). The anti-HSC70 and anti-paxillin were from Santa Cruz Biotechnology (cat. no. sc-24 and sc-5574). All primary antibodies were used in 1:1000 dilution in 3% BSA or 5% skimmed

milk besides the anti-HSC-70 and the anti-talin that were used in 1:5000 dilution and 1:500 respectively. Densitometric readings of band intensities were obtained using the ImageJ™ software.

Ex vivo aortic ring assay with Akt inhibitor

Thoracic aortas were isolated and prepared for culture as described previously²³. Briefly, 1mm thick rings were serum starved for 18 hours at 37°C before being embedded in growth factor-reduced Matrigel™ (BD Biosciences) and cultured in medium supplemented with 2.5% foetal calf serum (FCS) with or without Akt kinase inhibitor VIII (Calbiochem®, Millipore), at concentrations indicated. Rings were fed every 2 days with fresh medium containing 2.5% FCS and the indicated concentration of the inhibitor. Sprouting microvessels were counted after 6 days.

Ethical regulations

All animal experiments were performed in accordance with UK Home Office regulations and the European Legal Framework for the Protection of animals used for scientific purposes (European Directive 86/609/EEC).

Statistical analysis

Statistical significance was calculated with unpaired Student's t-test and $P < 0.05$ was considered statistically significant, unless otherwise indicated. For tumour growth statistics, non-parametric Mann-Whitney U rank sum test was performed to compare tumour volumes of each day. For Akt inhibition studies of aortic ring sprouting, data is presented using the log scale for the concentration of Akt inhibitor and statistical analysis between wild-type and FAK-heterozygous aortic rings at each concentration of Akt inhibitor was performed using multiple Student's t-test and $P < 0.05$ was considered statistically significant.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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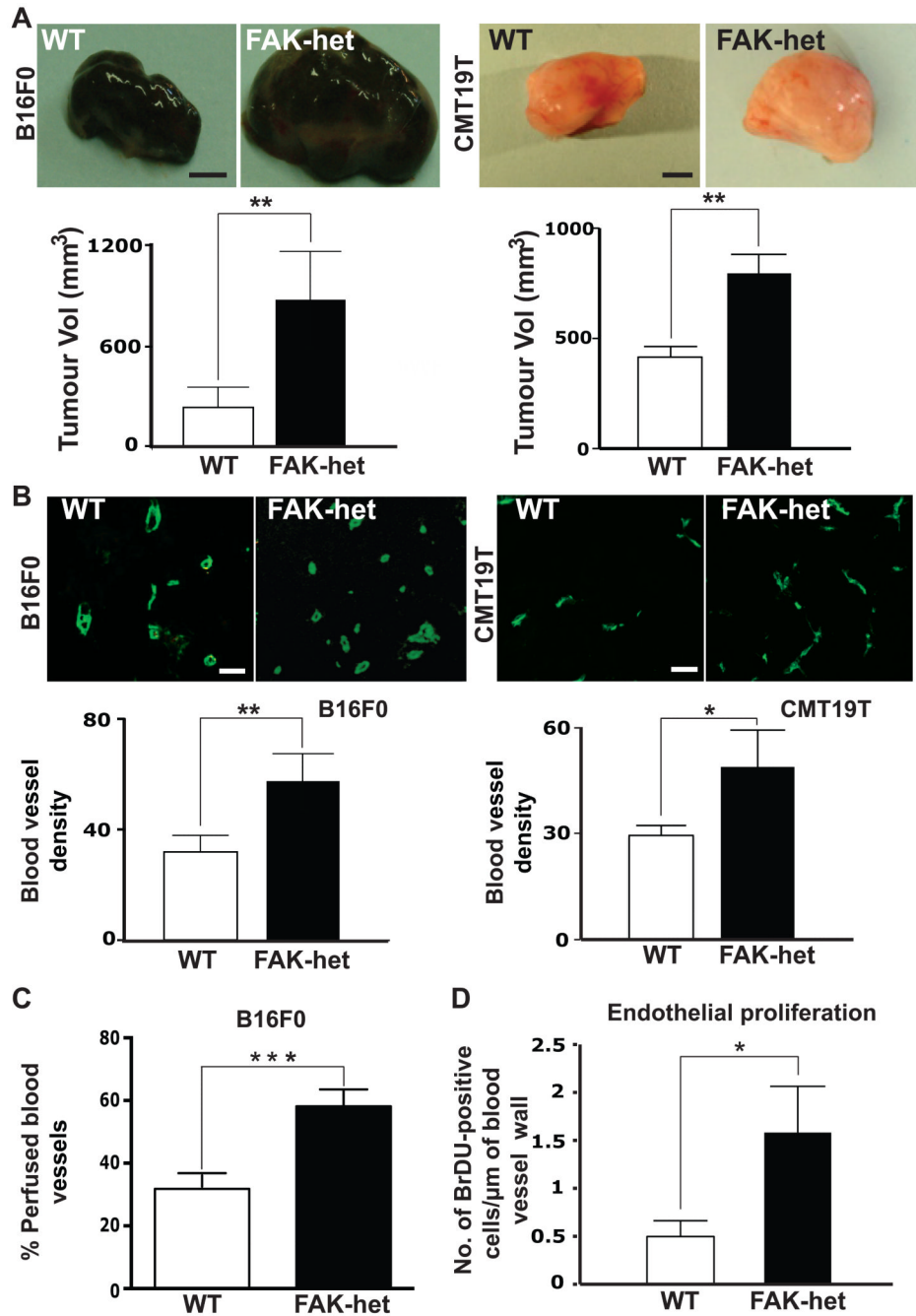


Figure 1. Increased tumour size and angiogenesis in FAK-heterozygous mice

(A) Subcutaneous 14 day-old B16F0 and CMT19T tumour sizes were elevated significantly in FAK-heterozygous (FAK-het) mice when compared with similar tumours in age- and sex-matched wild-type (WT) control littermates. *Top panels*, representative images of macroscopic appearance of tumours. Bar charts represent mean tumour volumes, +s.e.m; n=20 mice per genotype for the CMTs studies and n=30 mice per genotype for the B16F0 studies, Mann Whitney rank sum test, $**P<0.01$. (B) Endomucin staining patterns of midline sections of age-matched, size-matched B16F0 and CMT19T tumours from WT and FAK-het mice. Microvessel density is given as the number of blood vessels/mm² of midline tumour section. Microvessel density in B16F0 and CMT19T tumours was significantly greater in

FAK-het mice when compared with WT controls. Bar charts represent mean tumour blood vessel density of age-matched, size-matched tumours + s.e.m. n=20 CMTs tumours and n=30 B16F0 tumours per genotype, Student's t-test, $**P<0.01$, $*P<0.06$. **(C)** The number of functional tumour blood vessels was assessed by i.v. injection of PE-PECAM antibody before tumour excision in FAK-het and WT control mice. B16F0 tumours grown in FAK-het mice had an increased number of functional tumour blood vessels compared to WT littermates. Bar chart represents the % percentage of perfused vessels over total blood vessels per midline tumour section from WT and FAK-het mice + s.e.m. n=10 tumours per genotype, Student's t-test, $***P<0.001$. **(D)** Tumour endothelial proliferation was enhanced in FAK-het mice. Bar chart represents mean number of BrdU-positive ECs per blood vessel perimeter length (μm) in midline B16F0 tumour sections from WT and FAK-het mice + s.e.m. n= 5 tumours per genotype, Student's t-test, $*P<0.04$ Scale bars: 0.3cm **(A)**, 30 μm **(B)**.

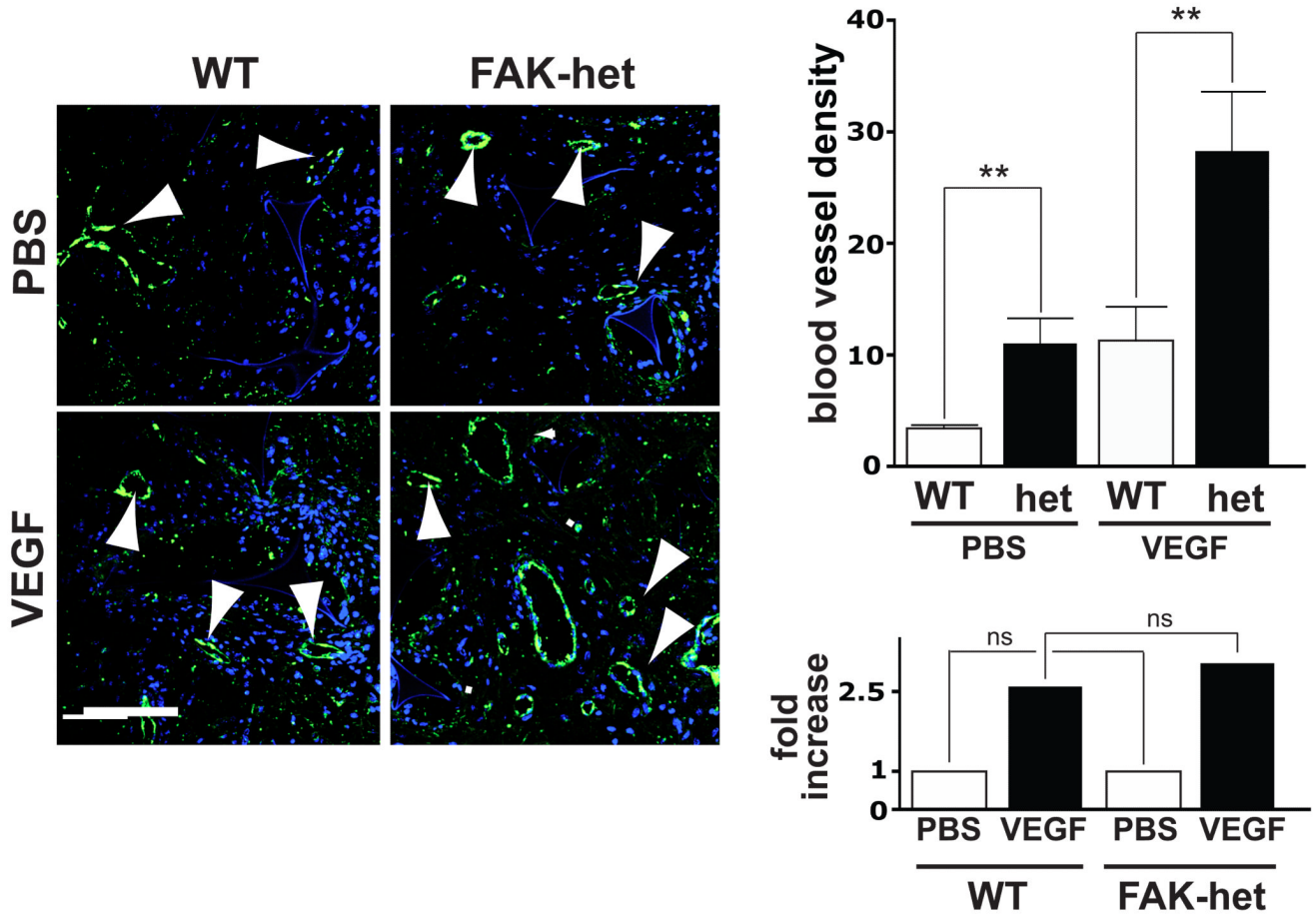


Figure 2. Tumour-free angiogenesis is enhanced in FAK-heterozygous mice

Sponges implanted into the flanks of either FAK-het mice or WT controls were injected with VEGF or PBS every two days for two weeks. At two weeks post implantation, *in vivo* angiogenesis was assessed by counting the numbers of blood vessels per mm² in sponge sections from WT and FAK-het mice. VEGF-treatment enhanced angiogenesis in both WT and FAK-het mice. However, quantitation showed that angiogenesis is elevated both at the baseline level, PBS-treated, (PBS) and after VEGF-stimulation (VEGF) in FAK-het mice. Upper bar chart represents mean numbers of blood vessels/ mm² of sponge section + s.e.m. Lower bar chart shows that the relative fold-increase in microvessel density of FAK-het mice over WT controls was similar for both PBS and VEGF-treated sponges. Arrowheads indicate blood vessels. DAPI-stained nuclei, blue, n=7 WT and n=10 FAK-het mice, Student's t-test, ns, not statistically significant different, ***P*<0.05. Scale bar: 100µm.

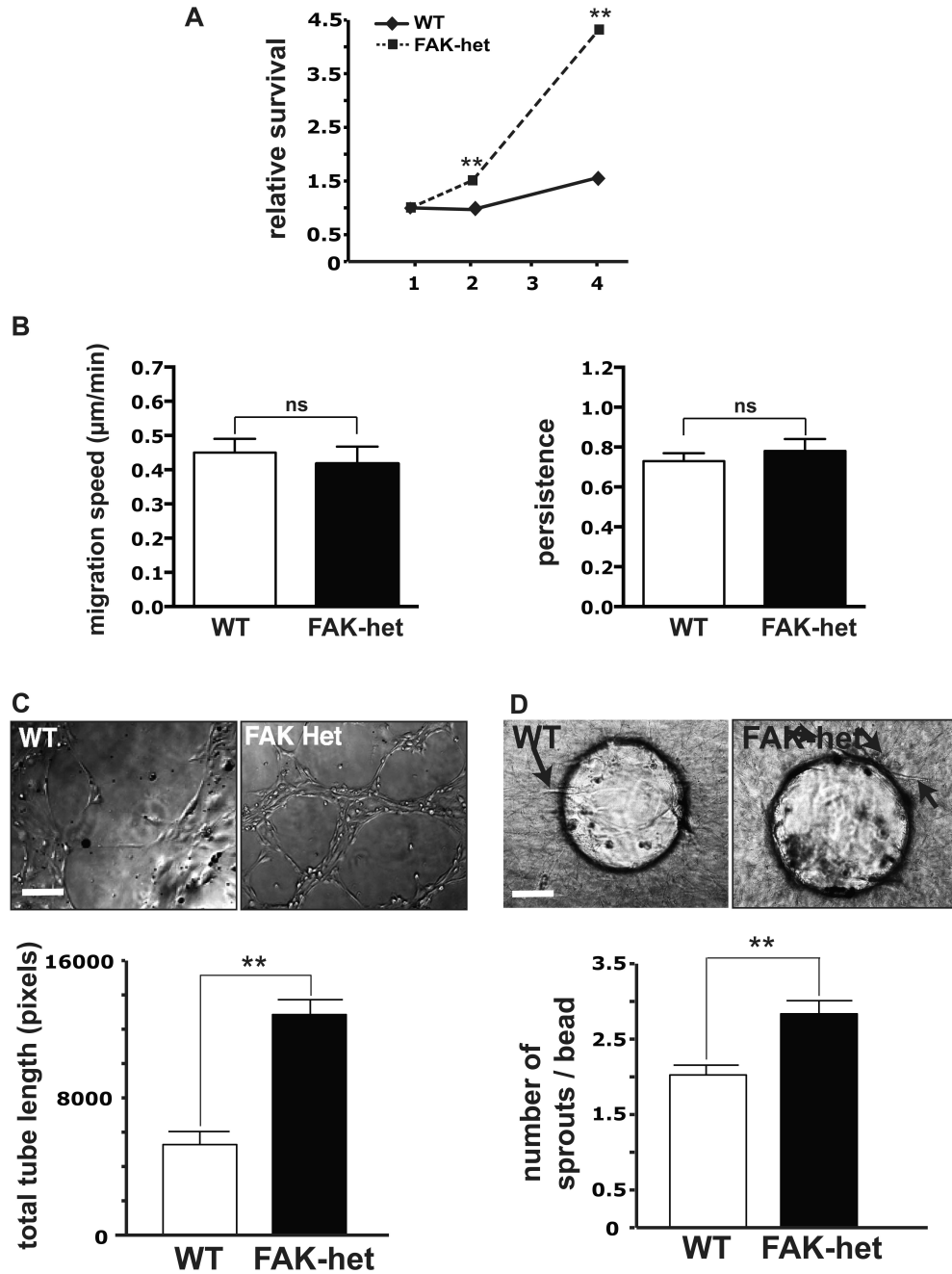


Figure 3. Characterisation of endothelial cells isolated from FAK-heterozygous mice (A) Using the WST-1 assay, primary EC survival *in vitro* was assessed. Graph shows that, over 4 days, survival of FAK-heterozygous (FAK-het) ECs was enhanced significantly over wild-type (WT) EC survival, n=44 wells/condition, 2500 cells/well, Student’s t-test, ****P<0.001**. (B) Serum stimulated speed of cell migration and persistence was assessed in Dunn chambers for WT and FAK-het ECs. No difference in speed or persistence was detected. Values are given as means + s.e.m. n=57 WT and n=62 FAK-het cells, Student’s t-test, ns, not statistically significant different. (C) Two-dimensional tube formation (total tube length) of FAK-het ECs was enhanced significantly when compared with WT cells.

Upper panels, representative images of endothelial tubes formed by WT and FAK-het primary ECs on Matrigel™, n=3, Student's t-test, $**P<0.01$. **(D)** Three-dimensional tube formation of FAK-het ECs was elevated significantly when compared with WT ECs. Representative images of endothelial sprouts from beads coated with WT and FAK-het cells cultured on beads in fibroblast-containing fibrin gel. Bar chart represents mean number of endothelial sprouts per bead + s.e.m., for WT and FAK-het ECs cultured on beads in fibroblast-containing fibrin gel. *Arrows* endothelial sprouts. n=54 WT and n=103 FAK-het beads, 3 independent experiments, Student's t-test, $***P<0.0005$ Scale bars in **(C)** and **(D)**, 50μm.

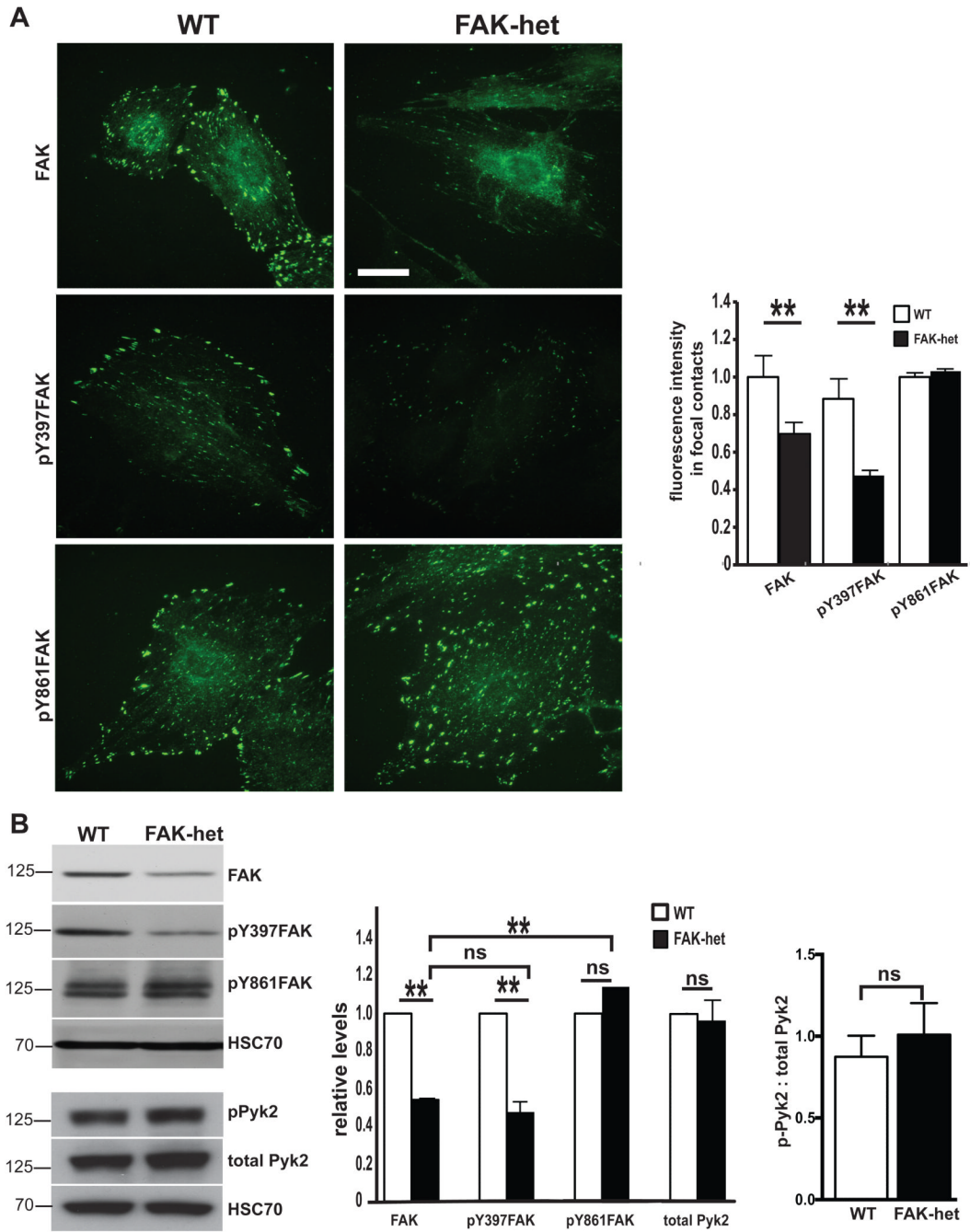


Figure 4. Imbalance between pFAK-Y397 and pFAK-Y861 in FAK-heterozygous cells
(A) Immunocytochemistry for FAK, pY397-FAK and pY861-FAK in cultured wild-type (WT) and FAK-heterozygous (FAK-het) ECs. Results show that the expression of FAK and pY397-FAK were reduced in FAK-het cells but that the expression of pY861-FAK was unchanged. Bar chart represents mean relative fluorescence signal intensity of FAK, pY397-FAK and pY861-FAK in FAK-het and WT ECs *in vitro* + s.e.m. n=5, Student's t-test, **P<0.01 **(B)** Western blot analysis of FAK, pY397-FAK, pY861-FAK, Pyk2 and phosphorylated Pyk2 (pPyk2) in cultured WT and FAK-het ECs lysates. HSC70 acted as a loading control. Results show that expression levels of total FAK and pY397-FAK are

similarly reduced in FAK-het cells but that pY861-FAK levels are unchanged between genotypes, resulting in significantly elevated ratio of pY861-FAK/total FAK while the ratio of pY397-FAK/total FAK remains the same in FAK-het ECs. The expression of Pyk2 and the ratio of pPyk2: total Pyk2 are similar in FAK-het and WT ECs. Bar chart represents densitometric readings relative to WT + s.e.m. n=3 independent experiments, Student's t-test $**P<0.01$, ns, not statistically significant different. Scale bar in (A), 25 μ m.

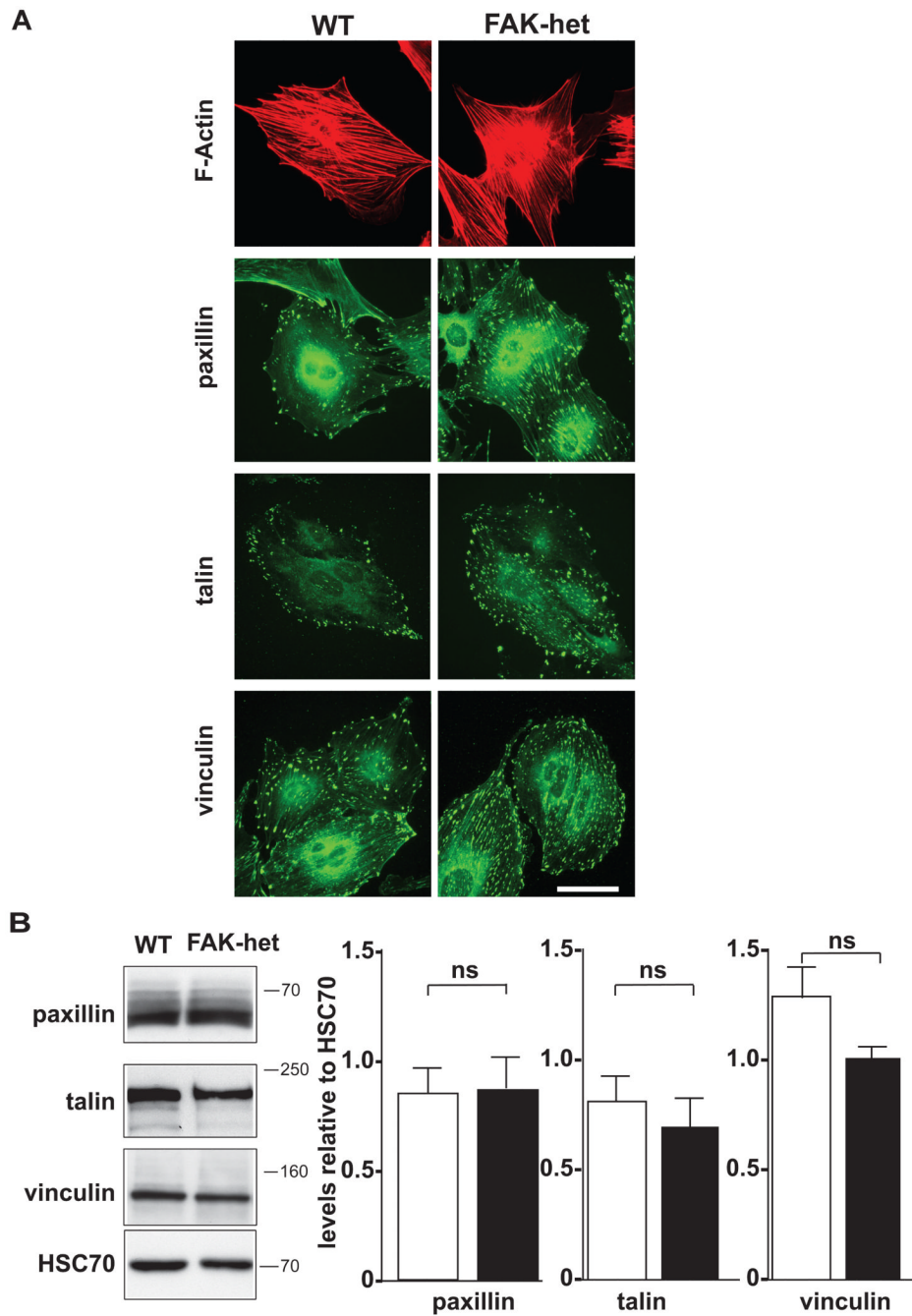


Figure 5. Expression of focal contact proteins in FAK-heterozygous endothelial cells
(A) Immunocytochemistry for F-actin, paxillin, talin and vinculin in cultured wild-type (WT) and FAK-heterozygous (FAK-het) ECs. **(B)** Western blot analysis of paxillin, talin and vinculin in cultured WT and FAK-het EC lysates. HSC70 acted as a loading control. Results show that overall expression levels of paxillin, talin and vinculin are unchanged between genotypes. Bar chart represents densitometric readings + s.e.m. n=4 independent experiments, Student's t-test, ns, no statistically significant difference. Scale bar in (A), 25 μ m.

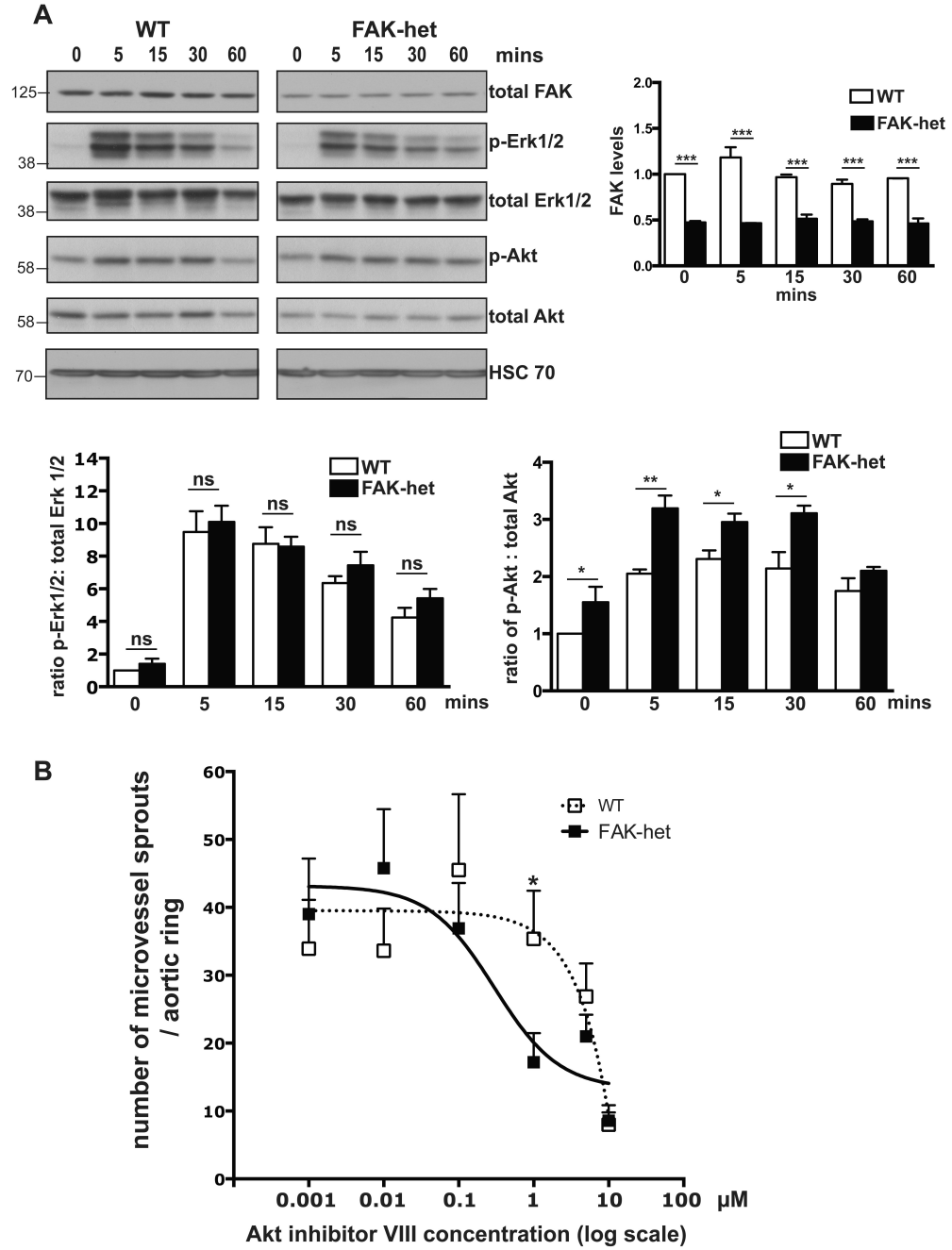


Figure 6. Effect of FAK heterozygosity on Akt signalling
(A) Western blot analysis of total FAK, phosphorylated-Erk1/2, total Erk1/2, phosphorylated-Akt and total Akt, in cultured wild-type (WT) and FAK-heterozygous (FAK-het) EC lysates after serum-stimulation for 0, 5, 15, 30 and 60 mins. HSC70 acted as loading control. Bar charts show mean densitometric readings of total FAK levels; ratios of p-Erk1/2:total Erk1/2; and ratios of p-Akt:total Akt from WT and FAK-het EC lysates after serum-stimulation for 0, 5, 15, 30 and 60 mins, + s.e.m. n=5 (FAK), n=3 (Erk) and n=4 (Akt) independent experiments, Student's t-test, *** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$, ns, no statistically significant difference. **(B)** Microvessel sprouting of FAK-het aortic rings displays higher sensitivity to inhibition of Akt activity than WT controls with lower dose of

Akt inhibitor causing a reduction in microvessel sprouting without affecting wild-type aortic rings. Bar chart represents the fitting curves of the average number of microvessel aortic sprouts + s.e.m. relative to the log concentrations of Akt inhibitor VIII. n= 12 aortic rings per treatment, Student's t-test, * $P < 0.05$