Computationally Guided Design of a Readily Assembled Phosphite– Thioether Ligand for a Broad Range of Pd-Catalyzed Asymmetric Allylic Substitutions

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Abstract

A modular approach employing indene as the common starting material has enabled the straightforward preparation (in three reaction steps) of a set of ligands for the palladiumcatalyzed asymmetric allylic substitution. The optimization of the first generation ligand library on the basis of rational design and theoretical calculations has provided an anthracenethiol- derivative that displays excellent behavior in the reaction of choice. Improving most approaches reported to date, this streamlined ligand presents a broad substrate and nucleophile scope. Excellent enantioselectivities have been therefore achieved for a range of linear and cyclic allylic substrates using a large series of C-, N- and Onucleophiles (40 examples in total). The species responsible for the catalytic activity have been further investigated by NMR and the origin of the enantioselectivity have been clearly established. The resulting products have been derivatized by means of ring-closing metathesis or Pauson-Khand reactions to further prove the synthetic versatility of the method.

1. Introduction

The future of chemical production must keep up with the growing demand for fine chemicals while reducing the overall waste production and energy consumption demanded by international regulations (and common sense). Over the last decades, this need for sustainability has driven the shift from suboptimal non-catalyzed processes to highperforming catalytic processes for the production of all sorts of chemicals.¹ This has been especially noteworthy in the production of enantiopure compounds, which play a key role in many technologically and biologically relevant applications.² Amongst the toolkit of catalytic enantioselective transformations, asymmetric Pd-catalyzed allylic substitution stands out for its versatility (as it creates new C-C and C-heteroatom bonds starting from simple precursors), high functional group tolerance and mild reaction conditions. Moreover, the resulting products accept further derivatization thanks to the presence of an alkene functionality.³ The key role of the ligand in the induction of chirality in this process has motivated several studies concerning the generation and evaluation of myriad candidates in terms of yield, selectivity and substrate scope. Heterodonor compounds (phosphine/phosphinite-oxazolines being the paradigmatic example) have proven especially advantageous because the different trans influence of the two donor groups generates an efficient electronic differentiation between the two allylic terminal carbon atoms. Indeed, the nucleophilic attack is known to take place predominantly trans to the donor group with stronger trans influence. On the basis of this premise, we have contributed with mixed ligands bearing biaryl phosphite moieties, 3^{j,4} which flexible nature allows the catalyst chiral pocket to adapt to the steric demands of the substrate4^e and , therefore, they have significantly broaden the substrate scope.

The vast amount of Pd-catalyzed allylic substitution studies reported in the literature might give the wrong impression that this is a mature field. However, despite the remarkable advances in catalyst design, ligands are still rarely suitable for a wide range of substrates. Instead, the most common scenario is that each allylic precursor requires independent optimization to identify the optimal catalytic system, and a similar situation takes place with the various nucleophiles. Consequently, the identification of "privileged" ligands remains a central task in this type of chemistry. In addition to giving excellent results for a broad range of allylic precursors and nucleophiles (C, N or O-based), such privileged ligands must be readily prepared from available starting materials and be easy to handle (i.e. solid, robust and stable in air).

To this end, we recently started a research line aimed at identifying suitable alternatives to the labile oxazoline moiety. We were especially interested in the stable and easy to prepare thioether group, which allowed the preparation of a Pd/phosphite-thioether furanoside-based catalyst that creates C-C, C-N and C-O bonds with different substrates and a variety of nucleophiles.⁵ The yields and enantioselectivities obtained were comparable to the best catalytic systems reported in the literature. Although these furanoside ligands were prepared from inexpensive D-Xylose, their synthesis was tedious and required a large number of steps. Other researchers have demonstrated the utility of thioether-based *P-S* ligands.⁶ For instance, the pioneering work in Pd-catalyzed allylic substitution and other relevant asymmetric reactions of Pregosin⁷ and Evans,6^a among others, put the focus on this kind of ligands and spurred their development. Despite the many efforts devoted to develop P-S ligands, their impact has been limited for two main reasons: (a) even in the most successful cases, they were limited in substrate and nucleophile scope: enantioselectivities were mainly high for the allylic substitution of the standard (and hindered) rac-1,3-diphenyl-3-acetoxyprop-1-ene S1 using dimethylmalonate as nucleophile,6 and (b) thioether-based ligands are prone to producing mixtures of diastereomeric thioether complexes, which tend to interconvert in solution.⁸ However, if one could design a scaffold able to control the S-coordination of the P-S ligand, the chiral element would move closer to the metal, thus giving rise to simpler ligands that can be prepared in less steps than their oxazoline-phosphine counterparts.

Herein, we give a new push to the study of the catalytic potential of *P*,*S*-ligands by screening readily accessible but novel thioether-containing compounds, including a detailed study of the species responsible for the catalytic performance. For this purpose, we designed a small but structurally diverse library of *P*-thioether ligands L1-L8a-g (Figure 1) that was tested in the Pd-catalyzed allylic substitution of a broad range of substrates and nucleophiles. These new *P*,*S*-ligands are synthesized in only three steps from inexpensive indene (ca. 20 USD/kg in bulk) and, since the corresponding enantiopure epoxide is prepared through Jacobsen epoxidation, both enantiomeric series are equally available. This modular approach⁹ greatly expedites the evaluation of several thioether and phosphite/phosphinite moieties, which is deemed crucial for the iterative optimization of the most promising candidates. Consequently, the catalytic performance of the ligands depicted in Figure 1 has been studied by systematically varying: (i) the electronic and steric properties of the thioether (L1–L8)

group, (ii) the configuration of the biaryl phosphite moiety $(\mathbf{a}-\mathbf{c})$, and (iii) the P-containing group (phosphite versus phosphinite groups, $\mathbf{d}-\mathbf{g}$).



Figure 1. Phosphite/phosphinite-thioether ligand library L1–L8a–g.

An additional advantage of this set of ligands is the fact that their simplified backbone renders very simple NMR spectra, thus reducing signal overlap, as well as facilitating the identification of relevant intermediates and accelerating the DFT calculations performed to rationalize the behavior of the system. By combining theoretical studies and NMR spectroscopy, we have been able to rationally fine-tune the ligands, improve enantioselectivity and identify the species responsible for the catalytic performance. This optimized ligand has proven active in the Pd-catalyzed allylic substitution of both linear and cyclic substates with a broad range of C-, N-, and O-nucleophiles (26 examples in total), even with the environmentally friendly propylene carbonate as solvent. Finally, the applicability of the new Pd/P-thioether catalysts has been further demonstrated in the practical synthesis of chiral (poly)carbocyclic and heterocycles using straightforward sequences of allylic alkylation/ring-closing metathesis or allylic alkylation/Pauson-Khand reactions.

2. Results and discussion

2.1. Synthesis of the first generation ligand library L1-L7a-g

Phosphite/phosphinite-thioether ligands L1-L7a-g can be efficiently prepared in three steps as illustrated in Scheme 1. In the first step, epoxidation of inexpensive indene 1 with bleach using Jacobsen's catalyst, followed by low-temperature crystallization, yielded indene oxide with 99% ee.¹⁰ Next, the regio- and stereospecific ring opening of 2 with the corresponding thiol was carried out with sodium hydroxide in a dioxane/water mixture.¹¹ In order to ensure chemical diversity, eight thiols with markedly different steric and electronic properties were used at this stage. Finally, we took advantage of the hydroxy group in 3–9 to establish a representative set of phosphite and phosphinite moieties following standard procedures.¹² The resulting enantiopure ligands were isolated in good yields as white solids (phosphite-thioether ligands L1–L7a–c) or colorless oils (phosphinite-thioether ligands L1–L7d–g). Phosphite-thioether ligands were found to be stable in air and resistant to hydrolysis, whereas the phosphinite analogues, proved less stable, slowly decomposing after a month even when stored at low temperature.



Scheme 1. Three-step synthesis of phosphite/phosphinite-thioether ligands L1–L7a–g from indene. (i) (*R*,*R*)-Mn-salen catalyst, 4-PPNO, aq. NaClO, CH₂Cl₂;¹⁰ (ii) RSH, NaOH, dioxane/H₂O (10:1);¹¹ (iii) ClP(OR¹R²)₂; (OR¹R²)₂= a–c, Py, toluene and (iv) ClPR³₂; R³= d–g, NEt₃, toluene.

All ligands were characterized by ${}^{31}P{}^{1}H$, ${}^{1}H$ and ${}^{13}C{}^{1}H$ NMR spectroscopy and HRMS. All data were in agreement with assigned structures.ⁱ See experimental section for purification and characterization details.

2.2. Evaluation of the first generation ligand library in the allylic substitution of symmetrical 1,3-disubstituted allylic substrates

ⁱ The spectra assignments were supported by the information obtained from ${}^{1}H{-}^{1}H$ and ${}^{1}H{-}^{13}C$ correlation measurements. The ${}^{31}P{}^{1}H$, ${}^{1}H$ and ${}^{13}C{}^{1}H$ NMR spectra showed the expected pattern for the C_{1} -ligands. The VT-NMR in CD₂Cl₂ (+35 to -85 °C) spectra showed only one isomer in solution. In all cases, one singlet in the ${}^{31}P{}^{1}H$ NMR spectra was observed.

As already mentioned, the catalyst ability to adjust to the steric demands of the substrate is a key factor in transferring the chiral information to the product. To assess the potential of this ligand library in the allylic substitution, we first tested **L1–L7a–g** in the Pd-catalyzed allylic substitution of two substrates with different steric requirements: the model substrate **S1** and the more challenging cyclic **S2** (Table 1). Excellent yields were almost invariably obtained under mild reaction conditions (i.e. 1 mol% Pd, ligand-to-palladium ratio of 1.1 at room temperature) with TOF as high as 2000 mol (mol h)⁻¹. As for the enantioselectivities, up to 97% ee for **S1** and 88% ee for **S2** could be achieved by using ligands that combine an aryl thioether group with a chiral biaryl phosphite moiety.

In an effort to measure the contribution of the different P-donor groups, we analyzed the results of ligands **L1a–g** (entries 1-7). The trend was clearly pointing out to a superior performance of phosphite- over phosphinite-based structures (i.e. entries 2 vs. 4-7), even with very bulky ones. The possibility that chirality of the ligand could control the conformation around the biphenyl moiety was ruled out by comparing entries 1-3, where the superior performance of ligands bearing a phosphite with axial chirality was evident (entries 2-3). Actually, the chiral axis seems to be the major factor in controlling the enantioselectivity. Indeed, ligands differing only in the configuration of this chiral axis (but otherwise having the same stereocenters) give rise to products with opposite absolute configuration (entries 2-3). Considering the substrates independently, with **S1** a remarkable cooperative effect between the configuration of the biaryl phosphite moiety and the ligand backbone is observed, which results in a matched combination with ligand **L1b**, that bears an (*R*) chiral axis (entry 2 vs 3). This cooperative effect is less pronounced for cyclic substrate **S2**, and both enantiomers of the alkylated products are therefore easily accessible with similar levels of enantioselectivity by simply setting the configuration of the biaryl phosphite moiety (entry 2 vs 3).

Finally, by further comparing ligands L1-L7b, we found that the electronic and steric properties of the thioether substituent have a small but important effect on the enantioselectivities: ligands with aryl-thioether groups led to higher ee's (especially with cyclic substrate S2; i.e. entry 11 vs 2, 8 and 9) than their counterparts with alkyl thioether moieties, even for the bulky *tert*-butyl thiol derivative. In summary, the best enantioselectivities for S1 (ee's up to 97%) were obtained with ligands L4–L7b, built from a combination of any aryl thioether group with an *R*-biaryl phosphite group: the above-

mentioned matched combination (entries 11, 12, 16 and 17). On the other hand, the best enantioselectivities recorded for substrate **S2** (ee's up to 88%), in both enantiomers of the alkylated product, were obtained with ligands **L4–L7b–c**, containing either an *R* or *S*-biaryl phosphite group with an aryl thioether moiety (*i.e.* entries 11–13 and 16–17).

With the aim of improving the sustainability profile of the process, we studied the reactions in 1,2-propylene carbonate (PC), an environmentally friendly alternative to standard organic solvents because of its high boiling point, low toxicity, and "green" synthesis.¹³ However, it has been scarcely used in asymmetric Pd-catalyzed allylic substitution and mainly limited to the standard **S1** substrate and dimethyl malonate as nucleophile.4^{d,13b,14} Thus, we repeated the allylic substitution of substrates **S1** and **S2** in PC (Table 1, entry 18; see also Table SI-1 in the Supporting Information for the use of PC for the allylic substitution of other substrates and nucleophiles). Gratifyingly, the enantioselectivities remained as high as those observed when dichloromethane was used.

OAc کی کھی ج		CO ₂ Me F	Pd/ L1-L7a-g (1 m	ol%) M	MeO ₂ C ₂ CO ₂ Me			
بر (rac)		CO ₂ Me	BSA / KOAc / CH	₂ Cl ₂	3. * r. *			
		OAc	; /~		OAc			
		Ph 🏹 S	´ `Ph 1		S2			
Entry	L	% Conv (h)	^b %ee ^c	% Conv	$(h)^b$ % ee^c			
1	L1a	100 (0.5)	17 (<i>R</i>)	100 (2) 15 (<i>S</i>)			
2	L1b	100 (0.5)	90 (<i>R</i>)	100 (2) 66 (<i>R</i>)			
3	L1c	100 (0.5)	75 (<i>S</i>)	100 (2) 61 (<i>S</i>)			
4	L1d	100 (0.5)	50 (R)	100 (2) 28 (<i>S</i>)			
5	L1e	100 (0.5)	25 (R)	100 (2) 11 (<i>S</i>)			
6	L1f	5 (0.5)	32 (<i>R</i>)	10 (2)	28 (S)			
7	L1g	100 (0.5)	4(R)	100 (2) 14 (<i>R</i>)			
8	L2b	100 (0.5)	90 (<i>R</i>)	100 (2) $62(R)$			
9	L3b	100 (0.5)	84 (<i>R</i>)	100 (2) $60(R)$			
10	L3e	100 (0.5)	63 (<i>R</i>)	100 (2) 77 (<i>S</i>)			
11	L4b	100 (0.5) ^d	97 (<i>R</i>)	100 (2) 85 (<i>R</i>)			
12	L5b	100 (0.5)	96 (<i>R</i>)	100 (2) 86 (<i>R</i>)			
13	L5c	100 (0.5)	80 (<i>S</i>)	100 (2) 84 (<i>S</i>)			
14	L5d	100 (0.5)	28 (R)	100 (2) 13 (<i>S</i>)			
15	L5e	100 (0.5)	40 (<i>R</i>)	100 (2) 11 (<i>S</i>)			
16	L6b	100 (0.5)	96 (<i>R</i>)	100 (2) 88 (<i>R</i>)			
17	L7b	100 (0.5)	97 (<i>R</i>)	100 (2) 87 (<i>R</i>)			
18 ^e	L5b	100 (1)	96 (<i>R</i>)	100 (4) 85 (<i>R</i>)			

 Table 1. Pd-catalyzed allylic alkylation of S1–S2 with dimethyl malonate as nucleophile

 using ligands L1–L7a–g.^a

^a 0.5 mol% [PdCl(η^3 -C₃H₅)]₂, ligand (0.011 mmol), substrate (1 mmol), CH₂Cl₂ (2 mL), BSA (3 equiv), nucleophile (3 equiv), KOAc (pinch) at rt. ^b Conversion measured by ¹H NMR. ^c Enantiomeric excesses measured by HPLC for dimethyl 2-(1,3-diphenylallyl)malonate (10) and by GC for dimethyl 2-(1,3-cyclohexanylallyl)malonate (11). ^d TOF= 2000 mol (mol h)⁻¹ calculated after 5 min from catalysis performed at 0.25 mol% of Pd. ^e Reactions carried out using PC as solvent at 40 °C.

2.3. Optimization of ligand parameters by DFT computational studies leading to fine-tuned phosphite-thioether ligands L8

With the ultimate goal of fine-tuning the ligands to increase enantioselectivity, we carried out DFT calculations of the transition states and key Pd-olefin intermediates involved in the enantiodetermining step. Previous mechanistic studies on Pd-catalyzed allylic alkylation have established the irreversible nucleophilic attack as the enantiodetermining step, although the corresponding transition state (TS) can be either early or late, depending on the nucleophile, ligands, and reaction conditions.¹⁵ In the latter case, the enantioselectivity of the final product is controlled by the formation of the most stable Pd-olefin complex, rather than by the difference in energy of the transition states leading to products.

Therefore, we started by calculating the relative stability of the transition states and the Pdolefin intermediates using the model substrate S1 and dimethyl malonate as nucleophile with ligands L5b-c. The goal was to evaluate the effect of the chiral axis of the biaryl phosphite moiety, as these ligands differ only in the configuration of this biaryl (see Table 1, entry 12 vs 13). Only the two syn-syn allyl complexes were calculated, neglecting the contribution of other allylic species of higher energy (*anti-anti* and *syn-anti*).3^d In this study, we have taken into account the configuration of the thioether and the attack of the nucleophile trans to P and S atoms. In contrast to P-N ligands, the trans effect exerted by the thioether and the phosphite are of a similar magnitude; indeed, previous studies have shown that small changes in the ligand can shift the *trans* preference in *P*,S-ligands.5^{b,16} The results of the most stable transition states ($TS_{(R)}$ and $TS_{(S)}$) and Pd-olefin intermediates (Pd-olefin_(R) and Pd-olefin_(S)) leading to the formation of both product enantiomers are shown in Table 2 (the full set of calculated TSs and Pd-olefin intermediates can be found in the Supporting Information). The energy differences of the calculated TSs match with the results obtained in the catalytic process, the value for L5b ($\Delta G^{\#}= 28 \text{ kJmol}^{-1}$; ee_{calc} > 99% (*R*)) being therefore higher than that of L5c ($\Delta G^{\#}$ = 10.8 kJmol⁻¹; ee_{calc} = 97% (S)). This is in agreement with the higher enantioselectivities achieved using L5b than L5c (96% (*R*) ee for L5b vs 80% (*S*) ee for L5c; Table 1, entries 12 and 13). Moreover, DFT correctly predicts the formation of the opposite product enantiomers when L5b and L5c are applied. It should be noted that, in contrast to what was observed with ligand L5b, both enantiomers of the substitution product obtained with ligand L5c arise from TSs with exo coordination of the substrate, with the nucleophilic attack *trans* to P (for the major enantiomer) and *trans* to S (for the minor enantiomer). Finally, the calculated energies of the Pd-olefin intermediates do not correlate well with the experimental results (Table 2). For both ligands, the most stable olefin complex corresponds to the *R*-enantiomer in more than 99% ee. The enantioselectivity is therefore not controlled by the rotation of the allylic system during the nucleophilic attack leading to the most stable Pd(0)-olefin complex.

Table 2. Calculated energies for the most stable transitions states (TS) and Pd- π -olefin complexes leading to the *R*- and *S*-enantiomers of the alkylated product of **S1** using dimethyl malonate as nucleophile.

Transition states (TS)									
Ligand	$\mathrm{TS}_{(R)}$	$TS_{(S)}$	% ee _{calc}	% ee _{exp}					
L5b	Ph Ph B C C C C C C C C C C C C C	R-S Ph Ph Ph Ph Ph Ph Ph Ph Ph Ph Ph Ph Ph	>99 (<i>R</i>)	96 (<i>R</i>)					
L5c	(R) R-S Ph Ph Pd-Po Ph E E 24.3 kJ/mol exo	R-S Ph Ph Ph Ph Ph Ph Ph Ph Ph Ph Ph Ph Ph	96 (<i>S</i>)	80 (<i>S</i>)					
Pd-olefin complexes									
Ligand	Pd-olefin _(R)	Pd-olefin _(S)	% ee _{calc}	% ee _{exp}					
L5b	Ph O kJ/mol	R-S 0 Ph Pd-P-0 E E 38.1 kJ/mol	>99 (<i>R</i>)	96 (<i>R</i>)					



Of all transition states (TSs) evaluated for both ligands, **L5b** and **L5c**, Figure 2 shows the two most stable. For the Pd/**L5b** catalytic system, it is seen that the *endo* TS is destabilized due to steric repulsion between one of the phenyl substituents of the substrate and the biaryl phosphite moiety. This increases the energy gap between the *endo* and *exo* TSs and could explain the preference for one of the pathways and, consequently, the higher enantiomeric excess achieved with Pd/**L5b** compared to Pd/**L5c**. In the Pd/**L5c** catalyst, this repulsion is less pronounced and therefore the two TSs have a more similar energy. The different steric constrains between both catalytic systems are reflected by the dihedral angles $\omega_1-\omega_3$ (Figure 2). Thus, the difference between the dihedral angles $\omega_1(C-O_1-P-Pd)$ and $\omega_3(C-O_1-P-O_2)$ of the TSs responsible for the formation of the *R*- and *S*-enantiomers (TS_(R) and TS_(S)) are higher for Pd/**L5b** than for Pd/**L5c** catalytic systems. It is also interesting to note the different spatial arrangement of the hydrogens of the methylene group of the ligand backbone. In the TSs of Pd/**L5c** the hydrogens are closer to the phosphite moiety than in the TSs of Pd/**L5b**, making the steric environment where the substrate is located less crowded. This again supports that the energy differences between the TSs are closer in Pd/**L5c** than in Pd/**L5b**.

Finally, we divided the energies of the transition states in: deformation energies of two moieties ([Pd-L5] and [substrate-nucleophile]) and in interaction energy between the two moieties. The results show very similar interaction energies for both ligands, but larger deformation energy for the [Pd/L5b] moiety than for [Pd/L5c]. This goes in line with previous findings showing that the transition states with L5c can accommodate better the substrate and nucleophile than those TSs containing L5b (see section SI-18 for details).



Figure 2. Most stable calculated transition states from **S1** using ligands (a) **L5b** and (b) **L5c** (hydrogen atoms have been omitted for clarity).

Based on the previous findings, we investigated whether the lower enantiomeric excesses recorded with the cyclic substrate **S2** (ee's up to 88%) could be improved by increasing the steric hindrance of the ligand. A simple way to do this is to introduce a thioether group that is bulkier than the 2,6-dimethylphenyl moiety, while maintaining the aryl groups (that have been shown to perform better than their alkylic counterparts). For this purpose, we ran analogous TS calculations for **S2** with ligand **L5b** (bearing the 2,6-dimethylphenyl thioether group) and with other ligands containing instead the bulkier 2,6-diisopropylphenyl or anthracenyl moieties. To accelerate the DFT calculation we used ammonia as model nucleophile.¹⁷ The results show that the enantioselectivity is affected by the steric effects of the thioether group to 35% with a 2,6-diisopropylphenyl thioether, and to 74% (*R*) with an anthracenyl thioether moiety. The results of these calculations prompted us to prepare two new ligands containing

an anthracenyl thioether group (**L8b** and **L8c**; Figure 1)ⁱⁱ and test them in the Pd-catalyzed alkylation of **S2**. To our delight, the introduction of this bulky aromatic moiety did affect positively the enantioselectivity, increasing from 86% ee to 94% ee (Table 3), as predicted by the theoretical calculations. This result is comparable to the best one reported in the literature for this challenging substrate.3 If we compare the calculated and experimental values (Table 3), we can conclude that, despite the fact that the calculated free energy differences are systematically lower than the experimental values, the general trend is reproduced well. The robustness of the theoretical model is demonstrated with the prediction of the new improved ligands **L8b**,**c** containing an anthracenyl moiety. Interestingly, ligand **L8b** also provided the highest enantioselectivity in the alkylation of linear substrate **S1** (ee's up to 99% (*R*), compared to previous best value 97% with ligand **L7b**).

 Table 3. Comparison between theoretical
 and experimental results in the Pd-catalyzed SR allylic substitution of S2. tBi $\Delta G^{\#}_{calc}$ R $\% \, e e_{exp}{}^a$ $\Delta G^{\#}_{exp}{}^{a}$ Ligand % ee_{calc} L5b 1.2 kJ/mol 9 (R) 86 (R) 6.3 kJ/mol 2.1 kJ/mol35 (R) L8b 6.8 kJ/mol 74 (*R*) 94 $(R)^{b}$ 8.6 kJ/mol

^a Reaction conditions: 0.5 mol% [PdCl(η^3 -C₃H₅)]₂, ligand (0.011 mmol), substrate (1 mmol), CH₂Cl₂ (2 mL), BSA (3 equiv), nucleophile (3 equiv), KOAc (pinch) at rt. ^b Ligand **L8c** provided the alkylated product **12** in 93% ee (*S*).

ⁱⁱ These ligands were prepared from (1*S*,2*S*)-1-(anthracen-9-ylthio)-2,3-dihydro-1H-inden-2-ol (**12**) as described in Scheme 1.

2.4. Allylic substitution of linear substrate S1 with several nucleophiles. Scope and limitations

We initially considered the allylic substitution of substrate **S1** with an extensive range of C-, N- and O-nucleophiles. Table 4 shows the results using ligand **L8b**, which had provided the best results in the allylic alkylation of **S1** with dimethyl malonate as model nucleophile. A variety of malonates, including the allyl-, butenyl-, pentenyl- and propargyl-substituted ones, reacted with **S1** to provide products **13–19** in high yields and enantioselectivities (ee's up to 99, entries 1–7). These substituted malonates are known to be more challenging nucleophiles for Pd-catalyzed allylic substitution, but they give rise to more interesting products from a synthetic point of view (see section 2.6 below). The addition of acetylacetone also proceeded with high enantiocontrol (entry 8, ee's up to 98%). High yields and enantioselectivities were also found in the addition of malononitrile and isopropyl cyanoacetate (products **21** and **22**; ee's up to 99%, entries 9 and 10) albeit the diastereoselectivity of the latter was low, as expected for such an acidic stereocentre.¹⁸

Pyrroles, which are electron-rich N-containing heterocycles interesting from the synthetic and biological point of view,¹⁹ also performed well as nucleophiles in this reaction. Despite their importance, only one catalytic system has been successful in the Pd-catalyzed allylic alkylation of **S1** type substrates with pyrroles, and this only at low temperature (-20 °C).²⁰ The difficulty of the transformation is more evident if we consider that, even two of the most successful ligands developed for this process (Trost diphosphine and phosphine-oxazoline PHOX), did not work with pyrroles.²⁰ Thus, we were pleased to see that using the Pd/**L8b** system we could reach ee's up to 99% and high yields working at room temperature (entries 11 and 12).

Chiral allylic amines are also ubiquitous in biologically active compounds, 3^n so we next studied the use of amine derivatives as nucleophiles. Benzylamine provided the substitution product **25** in high yield and enantioselectivity (99% ee; entry 13). To test the scope of allylic amination, the reaction of **S1** was evaluated using other N-nucleophilic compounds (entries 14–18). The combination Pd/L8b also proved highly efficient in the addition of *p*-methoxyand *p*-trifluoromethylbenzylamines (compounds **26** and **27**) and the furfurylamine **28** (entries 14–16), enantiocontrol being always excellent. The addition of morpholine, a cyclic secondary amine, also gave the expected product with high enantioselectivity (product **29**; entry 17), while allylamine proceeded with comparably high enantioselectivity (97% ee; entry 18). This is especially interesting given the fact that the amination product **30** is a key intermediate in the synthesis of complex molecules. For example, the Boc protected derivative of **30** can be further applied in metathesis reactions for the construction of a dihydropyrrole derivative (see section 2.6 below).

The exquisite enantiocontrol observed for C- and N-nucleophiles can also be extended to aliphatic alcohols (compounds 31-35, ee's up to 99%; entries 19-23). The effective allylic substitution with this type of O-nucleophiles opens up new synthetic avenues towards chiral ethers, which are important for the synthesis of biologically active molecules.²¹ Despite the potential of the resulting products, a general catalytic solution for the Pd-catalyzed allylic etherification has remained elusive and most of the few successful examples reported to date deal with phenols,²² while aliphatic alcohols have been less studied.4^{e,6a,23} Moreover, the enantioselectivities reported so far largely depend on the type of aliphatic alcohol and small modifications of their electronic properties 4^e, 6^{a,23} can have a large impact on this parameter. Using our streamlined ligand L8b, we found that benzylic alcohols gave excellent results regardless of the steric and electronic properties of the aryl group (entries 19-22). Allyl alcohol also furnished the desired product in high yield, and ee (entry 23). Even more outstanding are the almost perfect enantioselectivities (ee's up to 99%) and high yields achieved in the etherification of S1 with triphenylsilanol (entry 24), a rather unusual nucleophile that gives rise to a protected chiral alcohol.^{23e} Remarkably, enantioselectivities recorded with O-nucleophiles (entries 19-24) were, at the very least, as high as those obtained with dimethyl malonate.

			٥Ac		Pd/ L8b				
		Ph	Ph +	H–Nu	Ph + Ph				
Entry	Substrate	Product	% Yield ^b	% ee ^c	Fntry	Substrate	Product	% Vield ^b	% ee ^c
1	S1		94	99 (<i>R</i>)	13	Slostate S1		88	99 (<i>S</i>)
2	S 1	Bn0 OBn 14	92	98 (R)	14	S 1	MeO NH	81	99 (S)
3	S1	CO ₂ Me CO ₂ Me	92	97 (<i>S</i>)	15	S1	F3C NH 27	78	99 (S)
4	S1	CO ₂ Me CO ₂ Me	93	98 (<i>S</i>)	16	S1		83	97 (<i>S</i>)
5	S1		89	98 (<i>S</i>)	17	S1	0 V 29	87	98 (<i>S</i>)
6	S1		91	95(<i>S</i>)	18	S1		79	97 (<i>S</i>)
7	S1	CO ₂ Me CO ₂ Me	90	99 (<i>S</i>)	19 ^e	S1		92	99 (S)
8	S1		88	98 (R)	20 ^e	S1		90	99 (S)
9	S 1		84	99 (R)	21 ^e	S1	F ₃ C 0 33	91	98 (S)
10	S1		82 (60:40 dr)	98/97 ^d	22 ^e	S1		93	98 (<i>S</i>)
11 ^d	S 1		87	96 (<i>S</i>)	23 ^e	S1	35	83	96 (<i>S</i>)
12 ^d	S 1	N 24	85	>99 (<i>S</i>)	24 ^e	S1	Ph ₃ Si Q Q 36	78	99 (<i>R</i>)

Table 4. Pd-catalyzed allylic substitution of linear substrate **S1** with different types of C, N, and O nucleophiles using Pd/**L8b** catalytic system.^a

^a 0.5 mol% [PdCl(η^3 -C₃H₅)]₂, 1.1 mol% ligand, CH₂Cl₂ (2 mL), BSA (3 equiv), nucleophile (3 equiv), KOAc (pinch) at rt for 30 min. ^b Isolated yield. ^c Enantiomeric excesses measured by HPLC or GC. ^d 2 mol% [PdCl(η^3 -C₃H₅)]₂, 4.4 mol% ligand CH₂Cl₂ (2 mL), K₂CO₃ (2 equiv) at rt for 18 h. ^e 2 mol% [PdCl(η^3 -C₃H₅)]₂, 4.4 mol% ligand CH₂Cl₂ (2 mL), Cs₂CO₃ (3 equiv) at rt for 18 h.

2.5. Allylic substitution of several linear and cyclic substrates S2–S9 using several Cnucleophiles. Scope and limitations

After the broad scope of nucleophiles displayed by the catalytic system with S1, we turned our attention to the use of another five linear substrates (S3–S7) with electronic and steric requirements different from S1 (Table 5, entries 1–6). Advantageously, we found that the catalytic performance was neither affected by the introduction of electron-withdrawing and electron-donating groups (entries 1–3), nor by the introduction of *ortho-* and *metha*substituents at the phenyl groups of the substrate (entries 4–5). A remarkable enantioselectivity (entry 6) was still achieved in the Pd-catalyzed allylic alkylation of S7, a challenging substrate that typically gives rise to the corresponding substitution products in much lower enantioselectivities than S1 in otherwise identical conditions.

Finally, we wanted to see if the high enantioselectivities achieved in the allylic substitution of linear substrates were retained for their notoriously difficult cyclic analogues. To this end, a number of cyclic substrates with different ring sizes were tested using ligand L8b (Table 5; for the results using the Pd/L8c catalytic system see Table SI.2 in the Supporting Information). For substrate S2, a range of C-nucleophiles proved to give yields and enantioselectivities as high, if not higher, as those recorded with dimethyl malonate (ee's up to 97%, entries 7-11). The only exception was acetylacetone that led to somewhat lower enantioselectivity (entry 6). High enantioselectivities in both enantiomers of the substitution products were thus obtained using methyl-, allyl- and propargyl-substituted malonates (compounds 45–47; Table 5; entries 9–11 and Table SI.2). Furthermore, the biaryl phosphite group in Pd/L8b and Pd/L8c can adapt its chiral pocket to efficiently mediate the substitution of other cyclic substrates (entries 13-16). Excellent yields and enantioselectivities, comparable to the best reported in the literature, were obtained in the allylic alkylation of a 7membered cyclic substrate with different C-nucleophiles (products 51 and 52; entries 15 and 16). Even more interesting is that the good performance could be also extended to the allylic alkylation of a more challenging 5-membered cyclic substrate (compounds 49 and 50; entries 13 and 14).

	, n	LG f or	OAc	; +	F H— <mark>Nu</mark> ——	Pd/ L8b	► ⋼∕≫∕	Nu § or (Nu *	
	r. (ra	nc)	(rac)				37-4	* r. (2	43-52	
Entry	Substrate	Pro	oduct	%	% ee ^c	Entry	Substrate	Product	%	% ee ^c
				Yield ^b)				Yield ^b	
1	S 3	MeO		98	98 (<i>R</i>)	9	S 2	CO ₂ Me CO ₂ Me 45	87	91 (<i>R</i>)
2	S 3	3		87	97 (<i>R</i>)	10	S2	CO ₂ Me CO ₂ Me 46	84	94 (<i>R</i>)
3	S4	MeO		89	96 (<i>R</i>)	11	S 2	MeO ₂ C CO ₂ Me	86	97 (R)
4	S5	MeO MeO		91	97 (<i>R</i>)	12	S2		82	80 (-)
5	S 6	MeO		87	99 (<i>R</i>)	13	S8	MeO 49 OMe	80	84 (-)
6	S 7		O O Me	91	>95 (<i>R</i>)	14	S 8	MeO ₂ C CO ₂ Me	83	85 (-)
7	S2	EtO	43 OEt	87	95 (<i>R</i>)	15	S9	MeO O 51 OMe	90	96 (<i>R</i>)
8	S2	BnO	44 OBn	84	95 (<i>R</i>)	16	S 9	MeO ₂ C CO ₂ Me	92	96 (<i>R</i>)

 Table 5. Pd-catalyzed allylic substitution of substrates S2–S9 with several C nucleophiles

 using Pd/L8b catalytic system.^a

^a 0.5 mol% [PdCl(η^3 -C₃H₅)]₂, 1.1 mol% ligand, CH₂Cl₂ (2 mL), BSA (3 equiv), nucleophile (3 equiv), KOAc (pinch) at rt for 2 h. ^b Isolated yield. ^c Enantiomeric excesses measured by HPLC, GC or ¹H-NMR using [Eu(hfc)₃].

2.6 Synthetic applications of the allylic substitution compounds. Preparation of chiral functionalized (poly)carbocyclic and heterocyclic compounds 53–61

To illustrate the synthetic versatility of the compounds obtained from the enantioselective Pd-catalyzed allylic substitution, we have prepared a range of chiral functionalized carbocycles (53–56), heterocycles (57–58) and polycarbocycles (59–61). These compounds have been synthesized by straightforward reaction sequences involving allylic substitution of

the substrate followed by either ring-closing metathesis (Scheme 2) or Pauson-Khand enyne cyclization (Scheme 3).

According to this strategy, the alkylated compounds 16-18 (see Table 4 above) undergo clean ring-closing metathesis with no loss of enantiopurity, furnishing a number of 5, 6 and 7-membered carbocyles, in high yields and enantioselectivities (ee's ranging from 95–98%; Scheme 2). In an analogous manner, the O-heterocycle (*S*)-57 is achieved by sequential allylic etherification of **S1** with allylic alcohol and ring-closing metathesis reaction (Scheme 2); the corresponding N-heterocycle **58** performs similarly, albeit it requires protection of the amine with Boc prior to the ring-closing metathesis reaction, due to the azophilicity of ruthenium.



Scheme 2. Preparation of chiral functionalized carbo- and heterocyclic compounds 53–58.

The second derivatization we tackled was the Pauson-Khand reaction of the propargylated derivatives 47, 50 and 52 (see Table 5 above), which differ only in the size of the ring. Formation of the complex with $Co_2(CO)_8$, followed by thermal decomposition, gave rise to the [2+2+1] cycloadducts 59–61, which feature an architecturally complex tricyclic system with a *trans-cis* fusion (Scheme 3). Remarkably, the chiral information on the allylic substitution products was reliably conveyed to the final products, which were isolated as single diastereomers and with ee's replicating those of the starting materials. The relative

configuration of ketone **61** was assigned on the basis of a single crystal X-ray diffraction image, **59** and **60** being assigned by analogy.



Scheme 3. Preparation of chiral functionalized polycarbocyclic compounds 59–61. X-ray structure of compound 61 is also included.

2.7. Origin of enantioselectivity: study of the Pd- π -allyl intermediates

Our DFT calculations have established the nucleophilic attack as the enantiodetermining step (vide supra). With the aim of better understanding the catalytic process, we decided to prepare and characterize the Pd-allyl intermediates and determine their relative reactivity towards the nucleophile. Consequently, we studied the Pd- π -allyl compounds **62–65** [Pd(η^3 -allyl)(L)]BF₄ (L = L5b–c) by NMR and DFT studies. These Pd-intermediates containing cyclohexenyl and 1,3-diphenyl allyl groups were synthesized as previously reported (Scheme 4).²⁴ All complexes were characterized by ¹H, ¹³C and ³¹P NMR spectroscopyⁱⁱⁱ and mass spectrometry. Unfortunately, we were unable to obtain crystal of sufficient quality to perform

ⁱⁱⁱ The spectral assignments were confirmed using ¹H-¹H, ³¹P-¹H, ¹³C-¹H and ¹H-¹H NOESY experiments as well as DFT calculations.

X-ray diffraction measurements. The ESI-HR-MS showed the heaviest ions at m/z corresponding to the cation.

$$[PdCl(\eta^{3}-allyl)]_{2} + 2L \xrightarrow{AgBF_{4}} 2 [Pd(\eta^{3}-allyl)(L)]BF_{4} + 2 AgCl$$

$$62 allyl = cyclo-C_{6}H_{9}; L = L5b$$

$$63 allyl = cyclo-C_{6}H_{9}; L = L5c$$

$$64 allyl = 1,3-Ph_{2}-C_{3}H_{3}; L = L5b$$

$$65 allyl = 1,3-Ph_{2}-C_{3}H_{3}; L = L5c$$

Scheme 4. Preparation of $[Pd(\eta^3-allyl)(L)]BF_4$ complexes 62–65.

To understand why the opposite enantiomer is obtained when changing the configuration of the biaryl phosphite group, we compared the Pd-1,3-cyclohexenyl-allyl intermediate 62, which contains ligand L5b with its related counterpart Pd/L5c intermediate (63). The VT-NMR study (30 °C to -80 °C) showed the presence of essentially single isomers (ratio ca. 20:1; Scheme 5) for both intermediates (62 and 63). The major isomers were unambiguously assigned by NMR to be the exo isomer for 62 and the endo isomer for 63. In both cases, the thioether group had an S-configuration. For complex 62, the NOE indicated interactions between one of the tert-butyl groups of the phosphite moiety with the terminal allyl proton trans to the thioether group, whereas for compound 63 this interaction appeared with the methinic hydrogen of the CH-O group (Figure 2). The exo arrangement of the allyl group in complex 62 is further confirmed by a NOE interaction of the terminal allyl proton *trans* to the phosphite moiety with one of the methyl groups of the thioether moiety, while the other methyl of the thioether group presents a NOE interaction with the methinic hydrogen of the CH-S group (Figure 2). Similarly, the *endo* disposition of the allyl group in the complex **63** is further confirmed by the presence of NOE interactions with one of the methyls of the 2,6dimethylphenyl thioether group with the methinic hydrogen of the CH-S group, the central allyl proton and the terminal allyl proton *trans* to the phosphite moiety. The assignments are in agreement with the DFT calculations of the Pd- η^3 -cyclohexenylallyl complexes (see Supporting Information for the results of the full set of calculated Pd- η^3 -allyl intermediates). Thus, for Pd/L5b the major Pd- η^3 -exo isomer is 9.5 kJ/mol more stable than the most stable endo isomer, while for Pd/L5c the Pd- η^3 -endo isomer energy is 7.3 kJ/mol lower than the most stable exo isomer. The ¹³C NMR chemical shifts indicate that for both major isomers the most electrophilic allylic terminal carbon is *trans* to the phosphite group. Assuming that the nucleophilic attack takes place at the more electrophilic allyl carbon terminus, the fact that the observed stereochemical outcome of the reaction (86% ee (R) for Pd/L5b and 84% ee (S) for Pd/L5c) is similar to the diastereoisomeric excess (de 90%) of the Pd-intermediates indicates that both major and minor species react at a similar rate. In summary, the study of Pd-allyl intermediates shows that changes in the configuration of the phosphite moiety lead to changes in the ratio of the species that provide both enantiomers of the alkylated product. The enantioselectivity is therefore mainly controlled by the population of the Pd-allyl intermediates.



Scheme 5. Diastereoisomeric Pd- η^3 -allyl intermediates for S2 with ligands L5b and L5c. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbons and the relative DFT-calculated energies are also shown.



Figure 2. Relevant NOE contacts from the NOESY experiment of Pd- η^3 -allyl intermediates **62** and **63**.

Finally, to assess the impact of the phosphite chiral axis configuration on the enantioselectivity obtained for S1, we compared the corresponding Pd allylic intermediates with ligands L5b and L5c (64 and 65, respectively). In this case, L5b provided high enantioselectivity whereas L5c proved less selective, which is in contrast to the observation made for the alkylation of the cyclic substrate S2. The VT-NMR study (30 °C to -80 °C) of intermediates 64 and 65 showed a mixture of two isomers in equilibrium with ratios 1.3:1 and 2.3:1, respectively (Scheme 6). All isomers were assigned to be syn/syn, according to the NOE interaction between the two terminal protons of the allyl group. Unfortunately, the NOE contacts are not conclusive enough to unambiguously assign the 3D structure of these isomers. The final assignment of these Pd-allyl intermediates was performed by DFT studies (see Supporting Information) and further assessed by studying the reactivity of the Pdintermediates with sodium dimethyl malonate at low temperature by in situ NMR studies (Figure 3). The DFT calculated population of the different Pd-allyl species (i.e. ratio of 1.4:1 for complex 64) is in good agreement with the population obtained experimentally. Calculations indicate that for both systems, the most stable Pd-allyl intermediate is the exo isomer, the endo isomer being higher in energy (0.8 kJ/mol for Pd/L5b and 5.5 kJ/mol for Pd/L5c). On the other hand, the reactivity study of the Pd-intermediate 64 with sodium dimethyl malonate at low temperature reveals that the minor endo isomer reacts faster than the major isomer (Figure 3; $k_{endo}/k_{exo} \approx 8$). This reactivity pattern is in agreement with the previously presented TS calculations (see Section 2.2), which indicate that the most favourable (lowest in energy) transition state arises from the nucleophilic attack to the Pdallyl endo intermediate, being the pathway for the exo TS of much higher energy ($\Delta\Delta G^{\#}=28$ kJ/mol; Table 2). All these evidences further support the DFT calculations that suggest that for intermediates 64 the major isomer has an exo disposition, while the minor isomer has an endo spatial arrangement. In contrast, the reactivity study of the Pd-allyl complex 65 indicates that the major *exo* isomer is the isomer that reacts faster with a nucleophile (Figure 3; $k_{exo}/k_{endo} \approx 3$). Again, this finding is in agreement with the TS DFT calculations (vide supra, $\Delta\Delta G^{\#}$ = 10.8 kJ/mol, Table 2) and corroborates the DFT isomer assignment of the Pd-allyl intermediates observed in solution, having the major isomer an exo arrangement.

It should be pointed out that, albeit for the Pd/L5c catalytic system the relative population of the faster reacting isomer is much higher than that of Pd/L5b, the latter provides higher

enantioselectivity (96% ee for Pd/L5b vs 80% ee for Pd/L5c). Hence, in the case of S1 the enantioselectivity seems to be controlled by the different reactivity of the allyl intermediates towards the nucleophile (rather than their population, as was the case for S2). These results are in line with the previous TS DFT calculations (see section 2.3, Table 2) and therefore further corroborate that the energy gap between the most stable TSs leading to each of the product enantiomers is higher for the Pd/L5b catalytic system than for Pd/L5c.



Scheme 6. Diastereoisomer Pd- η^3 -allyl intermediates for S1 with ligands L5b and L5c. The relative amounts of each isomer are shown in parentheses. The chemical shifts (in ppm) of the allylic terminal carbons and the relative DFT-calculated energies are also shown.



Figure 3. ³¹P-{¹H}NMR spectra before and after the addition of sodium dimethyl malonate in CD₂Cl₂ at -80 °C of: (a) [Pd(η^3 -1,3-diphenylallyl)(**L5b**)]BF4 (64) (b) [Pd(η^3 -1,3-diphenylallyl)(**L5c**)]BF4 (65).

3. Conclusions

In summary, following a modular approach from enantiopure indene oxide, a phosphite/phosphinite-thioether ligand library has been prepared and benchmarked in the Pdcatalyzed asymmetric allylic substitution. After careful analysis of these results, and with the support of theoretical calculations, we have rationally designed a novel ligand that presents an improved enantioselectivity profile. The most remarkable feature of this optimized ligand is the broad scope demonstrated: linear and cyclic substrates, as well as a range of C-, N-, and O-nucleophiles (40 examples in total), all give rise to the desired products in excellent yields and enantioselectivities, even with the green propylene carbonate as solvent. Other advantages of the optimized ligand are that it is easily synthesized in only three steps from inexpensive indene and that it is solid and stable to air. Thorough mechanistic studies based on NMR spectroscopy have led to the identification of the species responsible for the catalytic performance, thus rationalizing the origin of the enantioselectivity. For enantioselectivities to be high, the ligand parameters therefore need to be correctly combined to either increase the difference in population of the possible Pd-allyl intermediates (for cyclic substrates) or to increase the relative rates of the nucleophilic attack for each of the possible Pd-allyl complexes (linear substrates). To assess the potential impact of this catalytic system in synthesis, the products have been employed in ring-closing metathesis or Pauson-Khand reactions, giving rise to a set of chiral (poly)carbocycles and heterocycles with retention of the enantioselectivity.

4. Experimental section

4.1. General considerations

All reactions were carried out using standard Schlenk techniques under an atmosphere of argon. Solvents were purified and dried by standard procedures. Phosphorochloridites were easily prepared in one step from the corresponding biaryls.²⁵ Enantiopure (–)-indene oxide 2^{10} and phosphinite-thioether ligands $L1d^{12a}$ and $L5d^{12a}$ were prepared as previously described. Racemic substrates $S1-S9^{26}$ and Pd-allyl complexes $[Pd(\eta^3-1,3-Ph_2-C_3H_3)(\mu-Cl)]_2^{27}$ and $[Pd(\eta^3-cyclohexenyl)(\mu-Cl)]_2^{28}$ were prepared as previously reported. ¹H, ¹³C{¹H}, and ³¹P{¹H} NMR spectra were recorded using a 400 MHz spectrometer. Chemical shifts are relative to that of SiMe₄ (¹H and ¹³C) as internal standard or H₃PO₄ (³¹P) as external standard.

¹H, ¹³C and ³¹P assignments were made on the basis of ¹H-¹H gCOSY, ¹H-¹³C gHSQC and ¹H-³¹P gHMBC experiments.

4.2. General procedure for the regio- and stereospecific ring opening of 2. Preparation of thioether-alcohols 3-9 and 12

A solution of (–)-indene oxide **2** (2 mmol, 264 mg) in dioxane (4.5 mL/mmol of indene oxide) is treated with the corresponding thiol (3 mmol). Then, a solution of NaOH (3 mmol, 120 mg) in water (0.45 mL/mmol of indene oxide) is added dropwise. The reaction mixture is capped and stirred at 55 °C until the epoxide is consumed according to TLC analysis (*ca.* 45-60 min). After this, the mixture is cooled to room temperature, diluted with water and extracted with CH_2Cl_2 (3 x 20 mL). The combined organic layers are dried over Na_2SO_4 and concentrated to give a residue that is purified by flash chromatography on silica gel (eluent specified in each case) to give the desired thioether-alcohol.

(1S,2S)-1-(Isopropylthio)-2,3-dihydro-1H-inden-2-ol (3). Yield: 308 mg (74%), white from cyclohexane/EtOAc solid. SiO₂-chromatography (gradient = 100:0 to cyclohexane/EtOAc = 80:20). ¹H NMR (400 MHz, CDCl₃): δ = 1.34 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.7 Hz), 1.38 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.7 Hz), 2.07 (bs, 1H, OH), 2.86 (dd, 1H, CH₂, ²J_{H-H} =16.1, ${}^{3}J_{H-H}$ =4.4 Hz), 3.14 (hept, 1H, CH, ${}^{i}Pr$, ${}^{3}J_{H-H}$ =6.7 Hz), 3.38 (dd, 1H, CH₂, ${}^{2}J_{H-H}$ =16.1, ${}^{3}J_{\text{H-H}}$ =6.2 Hz), 4.13 (d, 1H, CH-S, ${}^{3}J_{\text{H-H}}$ =4.1 Hz), 4.46-4.49 (m, 1H, CH-O), 7.22 (bs, 3H, CH=), 7.36 (m, 1H, CH=).¹³C NMR (100.6 MHz, CDCl₃): δ= 23.8 (CH₃), 24.2 (CH₃), 35.5 (CH), 39.9 (CH₂), 55.9 (CH-S), 79.9 (CH-O), 125.1 (CH=), 125.4 (CH=), 127.1 (CH=), 127.9 (CH=), 140.0 (C), 141.3 (C).

(1*S*,2*S*)-1-(Propylthio)-2,3-dihydro-1*H*-inden-2-ol (4). Yield: 325 mg (78%), white solid. SiO₂-chromatography (gradient from cyclohexane/EtOAc = 100:0 to cyclohexane/EtOAc = 80:20). ¹H NMR (400 MHz, CDCl₃): δ = 1.01 (t, 3H, CH₃, ⁿPr, ³*J*_{H-H} =7.3 Hz), 1.66 (sext, 2H, CH₂, ⁿPr, ³*J*_{H-H} =7.3 Hz), 2.10 (bs, 1H, OH), 2.53 (dt, 1H, CH₂, ²*J*_{H-H} =12.3, ³*J*_{H-H} =7.3 Hz), 2.60 (dt, 1H, CH₂, ²*J*_{H-H} =12.3, ³*J*_{H-H} =7.3 Hz), 2.87 (dd, 1H, CH₂, ²*J*_{H-H} =16.1, ³*J*_{H-H} =4.7 Hz), 3.37 (dd, 1H, CH₂, ²*J*_{H-H} =16.1, ³*J*_{H-H} =6.3 Hz), 4.09 (d, 1H, CH-S, ³*J*_{H-H} =4.3 Hz), 4.49 (quint, 1H, CH-O, ³*J*_{H-H} =5.0 Hz), 7.32-7.40 (m, 1H, CH=), 7.20-7.25 (m, 3H, CH=). ¹³C NMR (100.6 MHz, CDCl₃): δ = 13.6 (CH₃), 23.2 (CH₂), 33.0 (CH₂), 39.8 (CH₂), 57.0 (CH-S), 79.3 (CH-O), 125.1 (CH=), 125.3 (CH=), 127.1 (CH=), 127.9 (CH=), 140.1 (C), 140.6 (C). (1*S*,2*S*)-1-(*tert*-Butylthio)-2,3-dihydro-1*H*-inden-2-ol (5). Yield: 320 mg (72%), pale orange solid. SiO₂-chromatography (gradient from cyclohexane/EtOAc = 100:0 to cyclohexane/EtOAc = 80:20). ¹H NMR (400 MHz, CDCl₃): δ = 1.45 (s, 3H, CH₃, ^tBu), 2.29 (bs, 1H, OH), 2.86 (dd, 1H, CH₂, ²*J*_{H-H} =15.8, ³*J*_{H-H} =5.6 Hz), 3.32 (dd, 1H, CH₂, ²*J*_{H-H} =15.8, ³*J*_{H-H} =6.4 Hz), 4.03 (d, 1H, CH-S, ³*J*_{H-H} =5.3 Hz), 4.39-4.44 (m, 1H, CH-O), 7.19-7.25 (m, 3H, CH=), 7.38 (m, 1H, CH=), 7.36 (m, 1H, CH=).¹³C NMR (100.6 MHz, CDCl₃): δ = 31.7 (CH₃, ^tBu), 39.9 (CH₂), 43.7 (C, ^tBu), 54.6 (CH-S), 80.8 (CH-O), 124.8 (CH=), 125.6 (CH=), 127.2 (CH=), 127.7 (CH=), 139.7 (C), 142.0 (C).

(15,25)-1-(Phenylthio)-2,3-dihydro-1*H*-inden-2-ol (6). Yield: 373 mg (77%), white solid. SiO₂-chromatography (gradient from cyclohexane/EtOAc = 100:0 to cyclohexane/EtOAc = 80:20). ¹H NMR (400 MHz, CDCl₃): δ = 2.09 (bs, 1H, OH), 2.82 (dd, 1H, CH₂, ${}^{2}J_{H-H} = 16.3$, ${}^{3}J_{H-H} = 3.5$ Hz), 3.32 (dd, 1H, CH₂, ${}^{2}J_{H-H} = 16.3$, ${}^{3}J_{H-H} = 6.2$ Hz), 4.50 (dt, 1H, CH-O, ${}^{3}J_{H-H} = 6.2$ Hz, ${}^{3}J_{H-H} = 3.4$ Hz), 4.55 (d, 1H, CH-S, ${}^{3}J_{H-H} = 3.3$ Hz), 7.19-7.24 (m, 4H, CH=), 7.26-7.31 (m, 2H, CH=), 7.34-7.37 (m, 1H, CH=), 7.40-7.43 (m, 2H, CH=).¹³C NMR (100.6 MHz, CDCl₃): δ= 39.9 (CH₂), 59.1 (CH-S), 79.6 (CH-O), 125.2 (CH=), 125.7 (CH=), 126.9 (CH=), 127.2 (CH=), 128.3 (CH=), 129.0 (CH=), 130.9 (CH=), 135.2 (CH=), 139.9 (CH=), 135.2 (C), 139.9 (C), 140.6 (C).

(1*S*,2*S*)-1-((2,6-Dimethylphenyl)thio)-2,3-dihydro-1*H*-inden-2-ol (7). Yield: 427 mg (79%), white solid. SiO₂-chromatography (gradient from cyclohexane/EtOAc = 100:0 to cyclohexane/EtOAc = 80:20). ¹H NMR (400 MHz, CDCl₃): δ = 1.75 (bs, 1H, OH), 2.47 (s, 6H, CH₃), 2.82 (dd, 1H, CH₂, ²*J*_{H-H} =16.6, ³*J*_{H-H} =2.1 Hz), 3.52 (dd, 1H, CH₂, ²*J*_{H-H} =16.6, ³*J*_{H-H} =5.5 Hz), 4.32 (d, 1H, CH-S, ³*J*_{H-H} =2.0 Hz), 4.36 (tt, 1H, CH-O, ³*J*_{H-H} =5.5 Hz, ³*J*_{H-H} =2.1 Hz), 6.94 (d, 1H, CH=, ³*J*_{H-H} =7.5 Hz), 7.06-7.16 (m, 4H, CH=), 7.18-7.26 (m, 2H, CH=). ¹³C NMR (100.6 MHz, CDCl₃): δ = 21.9 (CH₃), 40.3 (CH₂), 58.8 (CH-S), 78.5 (CH-O), 125.3 (CH=), 125.4 (CH=), 126.7 (CH=), 128.1 (CH=), 128.2 (CH=), 128.7 (CH=), 132.0 (CH=), 140.4 (C), 140.7 (C), 143.6 (C).

(1*S*,2*S*)-1-((4-(Trifluoromethyl)phenyl)thio)-2,3-dihydro-1*H*-inden-2-ol (8). Yield: 478 mg (77%), yellow oil. SiO₂-chromatography (gradient from cyclohexane/EtOAc = 100:0 to cyclohexane/EtOAc = 80:20). ¹H NMR (400 MHz, CDCl₃): δ = 2.06 (d, 1H, OH, ³*J*_{H-H} =4.9 Hz), 2.91 (dd, 1H, CH₂, ²*J*_{H-H} =16.5, ³*J*_{H-H} =3.5 Hz), 3.42 (dd, 1H, CH₂, ²*J*_{H-H} =16.5, ³*J*_{H-H} =6.0 Hz), 4.55 (m, 1H, CH-O), 4.69 (d, 1H, CH-S, ³*J*_{H-H} =3.2 Hz), 7.21-7.30 (m, 3H, CH=), 7.36-7.41 (m, 1H, CH=), 7.46-7.57 (m, 4H, CH=). ¹³C NMR (100.6 MHz, CDCl₃): δ = 40.2 (CH₂),

58.0 (CH-S), 78.6 (CH-O), 124.1 (CH=), 125.4 (CH=), 125.7 (CH=), 125.8 (q, CH=, ${}^{3}J_{\text{H-F}}$ =3.8 Hz), 127.4 (CH=), 128.2 (q, C, ${}^{2}J_{\text{H-F}}$ =32.8 Hz), 128.7 (CH=), 128.8 (CH=), 139.0 (C), 140.6 (C), 141.3 (C).

(1*S*,2*S*)-1-((4-Methoxyphenyl)thio)-2,3-dihydro-1*H*-inden-2-ol (9). Yield: 541 mg (79%), yellow oil. SiO₂-chromatography (gradient from cyclohexane/EtOAc = 100:0 to cyclohexane/EtOAc = 75:25). ¹H NMR (400 MHz, CDCl₃): δ = 1.97 (d, 1H, OH, ³*J*_{H-H} =5.2 Hz), 2.81 (dd, 1H, CH₂, ²*J*_{H-H} =16.3, ³*J*_{H-H} =3.5 Hz), 3.25 (dd, 1H, CH₂, ²*J*_{H-H} =16.3, ³*J*_{H-H} =6.1 Hz), 3.79 (s, 3H, CH₃O), 4.38 (d, 1H, CH-S, ³*J*_{H-H} =3.3 Hz), 4.50 (tt, 1H, CH-O, ³*J*_{H-H} =6.1 Hz, ³*J*_{H-H} =3.5 Hz), 6.82 (d, 2H, CH=, ³*J*_{H-H} =8.7 Hz), 7.19-7.27 (m, 3H, CH=), 7.35-7.42 (m, 3H, CH=). ¹³C NMR (100.6 MHz, CDCl₃): δ = 39.9 (CH₂), 55.3 (CH-S), 60.6 (CH₃O), 78.6 (CH-O), 114.6 (CH=), 124.4 (CH=), 125.2 (CH=), 125.6 (CH=), 127.0 (CH=), 128.1 (CH=), 135.1 (C), 140.2 (C), 140.6 (C), 159.6 (C).

(15,2*S*)-1-(Anthracen-9-ylthio)-2,3-dihydro-1H-inden-2-ol (12). Yield: 308 mg (45%), yellow solid. SiO₂-chromatography (gradient from cyclohexane/EtOAc = 100:0 to cyclohexane/EtOAc = 80:20).. ¹H NMR (400 MHz, CDCl₃): δ = 1.33 (d, 1H, OH, ³*J*_{H-H} =5.1 Hz), 2.80 (dd, 1H, CH₂, ²*J*_{H-H} =16.4, ³*J*_{H-H} =2.6 Hz), 3.58 (dd, 1H, CH₂, ²*J*_{H-H} =16.4, ³*J*_{H-H} =5.6 Hz), 4.40 (m, 1H, CH-O), 4.55 (d, 1H, CH-S, ³*J*_{H-H} =2.2 Hz), 7.16 (t, 1H, CH=, ³*J*_{H-H} =7.5 Hz), 7.24 (t, 1H, CH=, ³*J*_{H-H} =7.5 Hz), 7.25-7.30 (m, 2H, CH=), 7.52 (ddd, 2H, CH=, ³*J*_{H-H} =8.0 Hz, ³*J*_{H-H} =6.5 Hz, ⁴*J*_{H-H} =1.0 Hz), 8.05 (d, 2H, CH=, ³*J*_{H-H} =8.4 Hz), 8.54 (s, 1H, CH=), 8.98 (dq, 2H, CH=, ³*J*_{H-H} =8.9, ⁴*J*_{H-H} =1.0 Hz). ¹³C NMR (100.6 MHz, CDCl₃): δ = 40.1 (CH₂), 61.2 (CH-S), 78.7 (CH-O), 125.3 (CH=), 125.4 (CH=), 125.7 (CH=), 126.6 (CH=), 126.9 (CH=), 127.0 (CH=), 127.7 (CH=), 128.4 (CH=), 129.1 (CH=), 129.6 (C), 131.8 (C), 134.9 (C), 140.2 (C), 140.8 (C).

4.3. General procedure for the preparation of phosphite-thioether ligands L1-L8a-c

The corresponding phosphorochloridite (1.1 mmol) produced in situ was dissolved in toluene (5 mL), and pyridine (0.3 mL, 3.9 mmol) was added. The corresponding thioether-hydroxyl compound (1 mmol) was azeotropically dried with toluene (3 x 2 mL) and then dissolved in toluene (5 mL) to which pyridine (0.3 mL, 3.9 mmol) was added. The alcohol solution was transferred slowly to a solution of phosphorochloridite. The reaction mixture was stirred at 80

°C for 90 min, after which the pyridine salts were removed by filtration. Evaporation of the solvent gave a white foam, which was purified by flash chromatography in silica (Hexane/Toluene/NEt₃ = 7/3/1) to produce the corresponding ligand as a white solid.

L1a. Yield: 320.8 mg (50%). ³¹P NMR (161.9 MHz, C₆D₆): δ =141.5 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.09 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.4 Hz), 1.07 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.8 Hz), 1.26 (s, 9H, CH₃, ⁱBu), 1.28 (s, 9H, CH₃, ⁱBu), 1.54 (s, 9H, CH₃, ⁱBu), 1.56 (s, 9H, CH₃, ⁱBu), 2.90-2.96 (m, 2H, CH₂, CH ⁱPr), 3.23 (dd, 1H, CH₂, ²J_{H-H} =16.4 Hz, ³J_{H-H} =5.6 Hz), 4.57 (b, 1H, CH-S), 5.18 (m, 1H, CH-OP), 6.94-7.12 (m, 3H, CH=), 7.31 (s, 1H, CH=), 7.33 (d, 1H, CH=, ⁴J_{H-H} =2.8 Hz), 7.34 (d, 1H, CH=, ⁴J_{H-H} =2.4 Hz), 7.57 (d, 1H, CH=, ⁴J_{H-H} =2.4 Hz), 7.59 (d, 1H, CH=, ⁴J_{H-H} =2.4 Hz), ¹³C NMR (100.6 MHz, C₆D₆): δ =23.4 (CH₃, ⁱPr), 23.8 (CH₃, ⁱPr), 31.0 (d, CH₃, ⁱBu, J_{C-P} =7.6 Hz), 31.1 (CH₃, ⁱBu), 34.2 (C, ⁱBu), 34.9 (C, ⁱBu), 35.3 (CH, ⁱPr), 39.1 (CH₂), 54.7 (CH-S), 82.8 (CH-OP), 124.1-146.6 (aromatic carbons). MS HR-ESI [found 669.3498, C₄₀H₅₅O₃PS (M-Na)⁺ requires 669.3502].

L1b. Yield: 220.8 mg (37%). ³¹P NMR (161.9 MHz, C₆D₆): δ =130.2 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.03 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.8 Hz), 1.23 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.4 Hz), 1.49 (s, 9H, CH₃, ⁱBu), 1.53 (s, 9H, CH₃, ⁱBu), 1.64 (s, 3H, CH₃), 1.76 (s, 3H, CH₃), 2.01 (s, 3H, CH₃), 2.07 (s, 3H, CH₃), 2.71-2.77 (m, 1H, CH, ⁱPr), 2.88 (d, 1H, CH₂, ²J_{H-H} =16.4 Hz), 3.27 (dd, 1H, CH₂, ²J_{H-H} =16.4 Hz, ³J_{H-H} =4.8 Hz), 4.81 (b, 1H, CH-S), 4.92 (m, 1H, CH-OP), 6.89-7.12 (m, 3H, CH=), 7.19 (m, 2H, CH=), 7.35 (d, 1H, CH=, ³J_{H-H} =8.8 Hz). ¹³C NMR (100.6 MHz, C₆D₆): δ =16.3 (CH₃), 16.6 (CH₃), 20.0 (CH₃), 20.1 (CH₃), 23.3 (CH₃, ⁱPr), 24.2 (CH₃, ⁱPr), 31.3 (CH₃, ⁱBu), 31.4 (d, CH₃, ⁱBu, J_{C-P} =5.3 Hz), 34.6 (C, ⁱBu), 34.7 (C, ⁱBu), 34.8 (CH, ⁱPr), 39.3 (CH₂), 55.0 (CH-S), 82.9 (CH-OP), 124.9-146.4 (aromatic carbons). MS HR-ESI [found 613.2903, C₃₆H₄₇O₃PS (M-Na)⁺ requires 613.2876].

L1c. Yield: 202.0 mg (34%). ³¹P NMR (161.9 MHz, C₆D₆): δ =139.4 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.09 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.8 Hz), 1.12 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.8 Hz), 1.47 (s, 9H, CH₃, ^tBu), 1.61 (s, 9H, CH₃, ^tBu), 1.69 (s, 3H, CH₃), 1.75 (s, 3H, CH₃), 2.05 (s, 6H, CH₃), 2.85-2.91 (m, 1H, CH, ⁱPr), 3.37 (d, 1H, CH₂, ²J_{H-H} =16.4 Hz), 3.38 (dd, 1H, CH₂, ²J_{H-H} =16.8 Hz, ³J_{H-H} =6.0 Hz), 4.24 (d, 1H, CH-S, ³J_{H-H} =4.8 Hz), 5.07-5.11 (m, 1H, CH-OP), 6.95-7.03 (m, 3H, CH=), 7.18 (s, 1H, CH=), 7.23 (2H, CH=). ¹³C NMR (100.6 MHz, C₆D₆): δ =16.3 (CH₃, ⁱPr), 16.5 (CH₃, ⁱPr), 20.1 (CH₃), 23.5 (CH₃), 23.9 (CH₃), 31.3 (CH₃, ^tBu), 31.4 (d, CH₃, ^tBu, J_{C-P} =5.3 Hz), 34.6 (C, ^tBu), 34.8 (CH, ⁱPr), 39.7 (CH₂), 54.7 (CH-S), 82.7 (d,

CH-OP, ${}^{2}J_{C-P} = 6.1$ Hz), 124.9-145.5 (aromatic carbons). MS HR-ESI [found 613.2869, C₃₆H₄₇O₃PS (M-Na)⁺ requires 613.2876].

L2b. Yield: 352.2 mg (59%). ³¹P NMR (161.9 MHz, C₆D₆): δ =132.4 (s). ¹H NMR (400 MHz, C₆D₆): δ =0.82 (pt, 3H, CH₃, Pr, ³J_{H-H}=7.2 Hz), 1.41-1.53 (m, 2H, CH₂, Pr), 1.53 (s, 9H, CH₃, ¹Bu), 1.58 (s, 9H, CH₃, ¹Bu), 1.69 (s, 3H, CH₃), 1.80 (s, 3H, CH₃), 2.05 (s, 3H, CH₃), 2.09 (s, 3H, CH₃), 2.23-2.30 (m, 1H, CH₂, Pr), 2.44-2.50 (m, 2H, CH₂, Pr), 2.93 (d, 1H, CH₂, ²J_{H-H}=16.8 Hz), 3.25 (dd, 1H, CH₂, ²J_{H-H}=16.4 Hz, ³J_{H-H}=5.2 Hz), 4.72 (b, 1H, CH-S), 4.90-4.94 (m, 1H, CH-OP), 6.92 (d, 1H, CH=, ³J_{H-H}=6.4 Hz), 7.00-7.03 (m, 2H, CH=), 7.22 (d, 2H, CH=, ³J_{H-H}=8.0 Hz), 7.37 (d, 2H, CH=, ³J_{H-H}=6.8 Hz). ¹³C NMR (100.6 MHz, C₆D₆): δ =13.2 (CH₃, Pr), 16.2 (CH₃), 16.5 (CH₃), 20.0 (CH₃), 20.1 (CH₃), 23.0 (CH₂, Pr), 31.3 (CH₃, ¹Bu, J_{C-P}=5.3 Hz), 31.4 (CH-S, ³J_{C-P}=3.0 Hz), 82.4 (CH-OP), 124.8-146.1 (aromatic carbons). MS HR-ESI [found 613.2903, C₃₆H₄7O₃PS (M-Na)⁺ requires 613.2876].

L3b. Yield: 283.6 mg (47%). ³¹P NMR (161.9 MHz, C₆D₆): δ =134.2 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.34 (s, 9H, CH₃, ¹Bu), 1.48 (s, 9H, CH₃, ¹Bu), 1.58 (s, 9H, CH₃, ¹Bu), 1.69 (s, 3H, CH₃), 1.74 (s, 3H, CH₃), 2.05 (s, 3H, CH₃), 2.06 (s, 3H, CH₃), 2.71 (d, 1H, CH₂, ²*J*_{H-H} =16.0 Hz), 3.12 (dd, 1H, CH₂, ²*J*_{H-H} =16.4 Hz, ³*J*_{H-H} =5.2 Hz), 4.56 (b, 1H, CH-S), 5.22-5.25 (m, 1H, CH-OP), 6.83 (d, 1H, CH=, ³*J*_{H-H} =7.2 Hz), 6.96-7.21 (m, 1H, CH=), 7.41 (d, 1H, CH=, ³*J*_{H-H} =7.6 Hz). ¹³C NMR (100.6 MHz, C₆D₆): δ =16.3 (CH₃), 16.5 (CH₃), 20.0 (CH₃), 20.1 (CH₃), 31.4 (CH₃, ¹Bu), 34.6 (C, ¹Bu), 34.7 (C, ¹Bu), 38.8 (CH₂), 43.5 (C, ¹Bu), 54.0 (d, CH-S, ³*J*_{C-P} =3.8 Hz), 83.7 (CH-OP), 124.7-145.7 (aromatic carbons). MS HR-ESI [found 627.3026, C₃₇H₄₉O₃PS (M-Na)⁺ requires 627.3032].

L4b. Yield: 284.8 mg (41%). ³¹P NMR (161.9 MHz, C₆D₆): δ =135.3 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.44 (s, 9H, CH₃, ¹Bu), 1.51 (s, 9H, CH₃, ¹Bu), 1.69 (s, 3H, CH₃), 1.77 (s, 3H, CH₃), 2.06 (s, 3H, CH₃), 2.07 (s, 3H, CH₃), 2.73 (d, 1H, CH₂, ²*J*_{H-H} =16.8 Hz), 2.93 (dd, 1H, CH₂, ²*J*_{H-H} =17.2 Hz, ³*J*_{H-H} =5.6 Hz), 4.95 (b, 1H, CH-S), 5.03-5.06 (m, 1H, CH-OP), 6.80-6.82 (m, 1H, CH=), 6.90-7.01 (m, 5H, CH=), 7.21 (d, 2H, CH=, ³*J*_{H-H} =3.6 Hz), 7.27-7,29 (m, 3H, CH=). ¹³C NMR (100.6 MHz, C₆D₆): δ =16.2 (CH₃), 16.5 (CH₃), 20.0 (CH₃), 20.1 (CH₃), 31.3 (CH₃, ¹Bu), 31.4 (CH₃, ¹Bu), 34.5 (C, ¹Bu), 34.6 (C, ¹Bu), 39.2 (d, CH₂, ³*J*_{C-P} =3.0 Hz), 58.8 (d, CH-S, ³*J*_{C-P} =3.8 Hz), 81.6 (d, CH-OP, ²*J*_{C-P} =4.6 Hz), 124.8-145.7 (aromatic carbons). MS HR-ESI [found 647.2737, C₃₉H₄₅O₃PS (M-Na)⁺ requires 647.2719].

L5b. Yield: 324.6 mg (42%). ³¹P NMR (161.9 MHz, C₆D₆): δ =135.7 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.42 (s, 9H, CH₃, ^tBu), 1.52 (s, 9H, CH₃, ^tBu), 1.68 (s, 3H, CH₃), 1.79 (s, 3H, CH₃), 2.04 (s, 3H, CH₃), 2.08 (s, 3H, CH₃), 2.31 (s, 6H, CH₃), 2.90 (d, 1H, CH₂, ²*J*_{H-H} =16.8 Hz), 3.38 (dd, 1H, CH₂, ²*J*_{H-H} =16.8 Hz, ³*J*_{H-H} =4.8 Hz), 4.80-4.83 (m, 1H, CH-OP), 4.91 (s, 1H, CH-S), 6.67 (d, 1H, CH=, ³*J*_{H-H} =7.2 Hz), 6.83 (pt, 1H, CH=, ³*J*_{H-H} =7.2 Hz), 6.88-7.03 (m, 5H, CH=), 719 (d, 1H, CH=, ⁴*J*_{H-H} =2.4 Hz). ¹³C NMR (100.6 MHz, C₆D₆): δ =16.2 (CH₃), 16.5 (CH₃), 20.0 (CH₃), 20.1 (CH₃), 21.7 (CH₃), 31.2 (CH₃, ^tBu), 31.3 (d, CH₃, ^tBu, *J*_{C-P} =5.4 Hz), 34.5 (C, ^tBu), 34.6 (C, ^tBu), 39.3 (d, CH₂, ³*J*_{C-P} =3.8 Hz), 58.0 (d, CH-S, ³*J*_{C-P} =3.8 Hz), 81.3 (d, CH-OP, ²*J*_{C-P} =4.6 Hz), 124.8-145.6 (aromatic carbons). MS HR-ESI [found 675.3026, C₄₁H₄₉O₃PS (M-Na)⁺ requires 675.3032].

L5c. Yield: 276.4 mg (36%). ³¹P NMR (161.9 MHz, C₆D₆): δ =137.5 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.44 (s, 9H, CH₃, ¹Bu), 1.56 (s, 9H, CH₃, ¹Bu), 1.69 (s, 3H, CH₃), 1.74 (s, 3H, CH₃), 2.05 (s, 3H, CH₃), 2.06 (s, 3H, CH₃), 2.25 (s, 6H, CH₃), 3.19 (d, 1H, CH₂, ²*J*_{H-H} =16.8 Hz), 3.50 (dd, 1H, CH₂, ²*J*_{H-H} =17.2 Hz, ³*J*_{H-H} =4.8 Hz), 4.60 (b, 1H, CH-S), 4.93 (m, 1H, CH-OP), 6.46 (d, 1H, CH=, ³*J*_{H-H} =7.2 Hz), 6.79 (m, 1H, CH=), 6.86-7.22 (m, 6H, CH=), 7.22 (s, 1H, CH=). ¹³C NMR (100.6 MHz, C₆D₆): δ =16.2 (CH₃), 16.4 (CH₃), 20.0 (CH₃), 20.7 (CH₃), 31.3 (CH₃, ¹Bu), 34.5 (C, ¹Bu), 34.6 (C, ¹Bu), 40.2 (d, CH₂, ³*J*_{C-P} =3.0 Hz), 57.5 (d, CH-S, ³*J*_{C-P} =3.8 Hz), 81.0 (d, CH-OP, ³*J*_{C-P} =7.6 Hz), 124.8-145.5 (aromatic carbons). MS HR-ESI [found 675.3041, C₄₁H₄₉O₃PS (M-Na)⁺ requires 675.3032].

L6b. Yield: 336.1 mg (52%). ³¹P NMR (161.9 MHz, C₆D₆): δ =135.6 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.41 (s, 9H, CH₃, ¹Bu), 1.47 (s, 9H, CH₃, ¹Bu), 1.69 (s, 3H, CH₃), 1.74 (s, 3H, CH₃), 2.05 (s, 3H, CH₃), 2.06 (s, 3H, CH₃), 2.67 (d, 1H, CH₂, ²*J*_{H-H} =16.8 Hz), 2.99 (dd, 1H, CH₂, ²*J*_{H-H} =16.8 Hz, ³*J*_{H-H} =6.0 Hz), 4.92 (b, 1H, CH-S), 4.96-5.01 (m, 1H, CH-OP), 6.82-6.84 (m, 1H, CH=), 6.99-7.19 (m, 8H, CH=), 7.24-7.26 (m, 1H, CH=). ¹³C NMR (100.6 MHz, C₆D₆): δ =16.9 (CH₃), 17.1 (CH₃), 20.6 (CH₃), 20.7 (CH₃), 31.9 (d, CH₃, ¹Bu, ³*J*_{C-P} =5.3 Hz), 32.0 (CH₃,¹Bu), 35.2 (C,¹Bu), 35.3 (C,¹Bu), 39.6 (CH₂), 58.3 (d, CH-S, ³*J*_{C-P} =3.8 Hz), 81.7 (d, CH-OP, ³*J*_{C-P} =4.6 Hz), 125.7-146.2 (aromatic carbons). MS HR-ESI [found 715.2610, C₄₀H₄₄F₃O₃PS (M-Na)⁺ requires 715.2593].

L7b. Yield: 321 mg (49%). ³¹P NMR (161.9 MHz, C₆D₆): δ =135.4 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.47 (s, 9H, CH₃, ¹Bu), 1.55 (s, 9H, CH₃, ¹Bu), 1.71 (s, 3H, CH₃), 1.82 (s, 3H, CH₃), 2.07 (s, 3H, CH₃), 2.10 (s, 3H, CH₃), 2.78 (d, 1H, CH₂, ²*J*_{H-H} =16.8 Hz), 2.87 (dd, 1H, CH₂, ²*J*_{H-H} =16.8 Hz, ³*J*_{H-H} =5.6 Hz), 3.16 (s, CH₃, *p*-OMe) 4.88 (b, 1H, CH-S), 5.04-5.07 (m,

1H, CH-OP), 6.54 (d, 2H, CH=, ${}^{3}J_{H-H}$ =8.8 Hz), 6.82 (d, 1H, CH=, ${}^{3}J_{H-H}$ =6.8 Hz), 6.96-7.23 (m, 6H, CH=), 7.31 (d, 1H, CH=, ${}^{3}J_{H-H}$ =7.2 Hz). 13 C NMR (100.6 MHz, C₆D₆): δ =16.7 (CH₃), 16.9 (CH₃), 20.4 (CH₃), 20.5 (CH₃), 31.7 (CH₃, 1 Bu), 31.8 (CH₃, 1 Bu), 35.0 (C, 1 Bu), 35.1 (C, 1 Bu), 39.9 (CH₂), 54.8 (CH₃, *p*-MeO), 60.5 (d, CH-S, ${}^{3}J_{C-P}$ =3.8 Hz), 82.4 (d, CH-OP, ${}^{3}J_{C-P}$ =4.6 Hz), 114.7-160.4 (aromatic carbons). MS HR-ESI [found 677.2851, C₄₀H₄₇O₄PS (M-Na)⁺ requires 677.2825].

L8b. Yield: 94.3 mg (27%). ³¹P NMR (161.9 MHz, C₆D₆): δ =136.1 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.28 (s, 9H, CH₃, ¹Bu), 1.39 (s, 9H, CH₃, ¹Bu), 1.65 (s, 3H, CH₃), 1.79 (s, 3H, CH₃), 2.02 (s, 3H, CH₃), 2.03 (s, 3H, CH₃), 2.91 (d, 1H, CH₂, ²J_{H-H} =16.8 Hz), 3.45 (dd, 1H, CH₂, ²J_{H-H} =16.8 Hz, ³J_{H-H} =5.2 Hz), 4.92-4.95 (m, 1H, CH-OP), 5.22 (b, 1H, CH-S), 6.55-6.63 (m, 2H, CH=), 6.90 (s, 2H, CH=), 7.11-7.28 (m, 6H, CH=), 7.73 (d, 2 H, CH=, ³J_{H-H} =8,0 Hz), 8.16 (s, 1, CH=), 8.96 (d, 2 H, CH=, ³J_{H-H} =8,4 Hz). ¹³C NMR (100.6 MHz, C₆D₆): δ =16.2 (CH₃), 16.6 (CH₃), 20.0 (CH₃), 20.1 (CH₃), 31.1 (CH₃, ¹Bu), 31.3 (d, CH₃, ¹Bu, J_{C-P} =5.4 Hz), 34.4 (C, ¹Bu), 39.7 (CH₂), 60.1 (CH-S), 81.4 (CH-OP), 124.8-145.7 (aromatic carbons). MS HR-ESI [found 747.3048, C₄₇H₄₉O₃PS (M-Na)⁺ requires 747.3028].

L8c. Yield: 102 mg (29%). ³¹P NMR (161.9 MHz, C₆D₆): δ =134.7 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.33 (s, 9H, CH₃, ¹Bu), 1.45 (s, 9H, CH₃, ¹Bu), 1.66 (s, 3H, CH₃), 1.84 (s, 3H, CH₃), 2.01 (s, 3H, CH₃), 2.07 (s, 3H, CH₃), 2.94 (d, 1H, CH₂, ²*J*_{H-H} =16.2 Hz), 3.54 (dd, 1H, CH₂, ²*J*_{H-H} =16.2 Hz, ³*J*_{H-H} =6.2 Hz), 5.02 (m, 1H, CH-OP), 5.27 (b, 1H, CH-S), 6.64 (m, 2H, CH=), 6.95 (s, 2H, CH=), 7.11-7.27 (m, 6H, CH=), 7.72 (d, 2 H, CH=, ³*J*_{H-H} =7.6 Hz), 8.04 (s, 1, CH=), 8.92 (d, 2 H, CH=, ³*J*_{H-H} =8.0 Hz). ¹³C NMR (100.6 MHz, C₆D₆): δ =16.4 (CH₃), 16.5 (CH₃), 20.2 (CH₃), 20.7 (CH₃), 31.4 (CH₃, ¹Bu), 31.7 (CH₃, ¹Bu), 34.6 (C, ¹Bu), 34.8 (C, ¹Bu), 40.1 (CH₂), 58.4 (CH-S), 81.4 (CH-OP), 125.2-146.9 (aromatic carbons). MS HR-ESI [found 747.3032, C₄₇H₄₉O₃PS (M-Na)⁺ requires 747.3028].

4.4. General procedure for the preparation of phosphinite-thioether ligands L1-L8d-g

The corresponding thioether-hydroxyl compound (0.5 mmol) and DMAP (6.7 mg, 0.055 mmol) were dissolved in toluene (1 ml), and triethylamine was added (0.09 ml, 0.65 mmol) at rt, followed by the addition of the corresponding chlorophosphine (0.55 mmol) via syringe. The reaction was stirred for 20 min at room temperature. The solvent was removed *in vacuo*,

and the product was purified by flash chromatography on alumina (toluene/NEt₃ = 100/1) to produce the corresponding ligand as an oil.

L1e. Yield: 257.8 mg (61%). ³¹P NMR (161.9 MHz, C₆D₆): δ =98.2 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.08 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.8 Hz), 1.21 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.4 Hz), 2.37 (s, 3H, CH₃, *o*-Tol), 2.41 (s, 3H, CH₃, *o*-Tol), 2.91-3.01 (m, 2H, CH ⁱPr, CH₂), 3.30 (dd, 1H, CH₂, ²J_{H-H}=16.4 Hz, ³J_{H-H}=6.0 Hz), 4.49 (b, 1H, CH-S), 4.82 (m, 1H, CH-OP), 6.91-7.15 (m, 9H, CH=), 7.36 (d, 1H, CH=, ³J_{H-H}=6.8 Hz), 7.52 (m, 2H, CH=). ¹³C NMR (100.6 MHz, C₆D₆): δ =20.3 (d,CH₃, *o*-Tol, ³J_{C-P}=4.0 Hz), 20.5 (d,CH₃, *o*-Tol, ³J_{C-P}=4.4 Hz), 23.2 (CH₃, ⁱPr), 23.8 (CH₃, ⁱPr), 35.3 (CH, ⁱPr), 39.1 (d, CH₂, ³J_{C-P}=6.1 Hz), 54.7 (d, CH-S, ³J_{C-P}=6.1 Hz), 87.7 (d, CH-OP, ²J_{C-P}=20,7 Hz), 124.8-141.4 (aromatic carbons).

L1f. Yield: 128.9 mg (61%). ³¹P NMR (161.9 MHz, C₆D₆): δ =140.4 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.08-1.20 (m, 6H, CH₂, Cy), 1.22 (d, 3H, CH₃, ⁱPr, ³J_{H-H}=6.4 Hz), 1.25-1.35 (m, 5H, CH₂, Cy), 1.38 (d, 3H, CH₃, ⁱPr, ³J_{H-H}=6.4 Hz), 1.48-1.61 (m, 5H, CH₂, Cy), 1.69 (b, 5H, CH, CH₂, Cy), 1.86 (m, 2H, CH₂, Cy), 2.98 (d, 1H, CH₂, ²J_{H-H}=16.0 Hz), 3.10-3.17 (m, 1H, CH ⁱPr), 3.41 (dd, 1H, CH₂, ²J_{H-H}=16.4 Hz, ³J_{H-H}=5.6 Hz), 4.51-4.54 (m, 2H, CH-S, CH-OP), 6.99-7.15 (m, 3H, CH=), 7.40 (d, 1H, CH=, ³J_{H-H}=8.0 Hz), 7.52 (m, 2H, CH=). ¹³C NMR (100.6 MHz, C₆D₆): δ =23.3 (CH₃, ⁱPr), 24.0 (CH₃, ⁱPr), 26.5-27.1 (CH₂, Cy), 28.1 (CH₂, Cy), 28.3 (CH₂, Cy), 28.5 (CH₂, Cy), 35.2 (CH, ⁱPr), 37.6 (d, CH, ¹J_{C-P}=8.5 Hz), 37.8 (d, CH, ¹J_{C-P}=9.9 Hz), 39.3 (d, CH₂, ³J_{C-P}=6.1 Hz), 54.8 (d, CH-S, ³J_{C-P}=6.1 Hz), 87.7 (d, CH-OP, ²J_{C-P}=18.4 Hz), 124.8-141.5 (aromatic carbons).

L1g. Yield: 129.9 mg (54%). ³¹P NMR (161.9 MHz, C₆D₆): δ =114.5 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.11 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.8 Hz), 1.22 (d, 3H, CH₃, ⁱPr, ³J_{H-H} =6.4 Hz), 2.04 (s, 3H, *p*-CH₃, Mes), 2.06 (s, 3H, *p*-CH₃, Mes), 2.39 (s, 12H, *o*-CH₃, Mes), 2.85-2.99 (m, 2H, CH ⁱPr, CH₂), 3.33 (dd, 1H, CH₂, ²J_{H-H} =16.4 Hz, ³J_{H-H} =5.6 Hz), 4.46 (b, 1H, CH-S), 4.66 (m, 1H, CH-OP), 6.63 (s, 1H, CH=), 6.64 (s, 1H, CH=), 6.65 (s, 1H, CH=), 6.66 (s, 1H, CH=), 6.94-7.05 (m, 2H, CH=), 7.12 (m, 1H, CH=), 7.31 (m, 1H, CH=). ¹³C NMR (100.6 MHz, C₆D₆): δ =20.6 (*p*-CH₃, Mes), 22.1 (d, *o*-CH₃, Mes, ³J_{C-P} =3.0 Hz), 22.2 (d, *o*-CH₃, Mes, ³J_{C-P} =3.1 Hz), 23.2 (CH₃, ⁱPr), 23.9 (CH₃, ⁱPr), 35.2 (CH, ⁱPr), 38.9 (d, CH₂, ³J_{C-P} =6.8 Hz), 54.6 (d, CH-S, ³J_{C-P} =7.6 Hz), 87.6 (d, CH-OP, ²J_{C-P} =22,1 Hz), 124.7-141.6 (aromatic carbons).

L3e. Yield: 147,6 mg (31%). ³¹P NMR (161.9 MHz, C₆D₆): δ =97.7 (s). ¹H NMR (400 MHz, C₆D₆): δ =1.24 (s, 9H, CH₃, ^tBu), 2.32 (s, 3H, CH₃, *o*-Tol), 2.41 (s, 3H, CH₃, *o*-Tol),

2.92 (dd, 1H, CH₂, ${}^{2}J_{\text{H-H}}$ =16.4 Hz, ${}^{3}J_{\text{H-H}}$ =2.8 Hz), 3.22 (dd, 1H, CH₂, ${}^{2}J_{\text{H-H}}$ =16.0 Hz, ${}^{3}J_{\text{H-H}}$ =5.2 Hz), 4.40 (b, 1H, CH-S), 4.82-4.85 (m, 1H, CH-OP), 6.89-7.12 (m, 9H, CH=), 7.39 (d, 1H, CH=, ${}^{3}J_{\text{H-H}}$ =7.2 Hz), 7.50-7.54 (m, 2H, CH=). 13 C NMR (100.6 MHz, C₆D₆): δ =20.2 (d, CH₃, ${}^{3}J_{\text{C-P}}$ =15.3 Hz), 20.4 (d, CH₃, ${}^{3}J_{\text{C-P}}$ =16.0 Hz), 31.3 (CH₃, 4 Bu), 39.0 (d, CH₂, ${}^{3}J_{\text{C-P}}$ =6.0 Hz), 43.4 (C, 4 Bu), 53.6 (d, CH-S, ${}^{3}J_{\text{C-P}}$ =6.8 Hz), 88.5 (d, CH-OP, ${}^{3}J_{\text{C-P}}$ =20.6 Hz), 124.6-142.3 (aromatic carbons). MS HR-ESI [found 457.1731, C₂₇H₃₁OPS (M-Na)⁺ requires 457.1725].

L5e. Yield: 260.6 mg (54%). ³¹P NMR (161.9 MHz, C₆D₆): δ =98.5 (s). ¹H NMR (400 MHz, C₆D₆): δ =2.30 (s, 3H, CH₃, *o*-Tol), 2.35 (s, 3H, CH₃, *o*-Tol), 2.44 (s, 6H, CH₃), 3.14 (d, 1H, CH₂, ²J_{H-H} =16.8 Hz), 3.54 (dd, 1H, CH₂, ²J_{H-H} =16.8 Hz, ³J_{H-H} =5.2 Hz), 4.78-4.82 (m, 1H, CH-OP), 4.83 (b, 1H, CH-S), 6.91-7.14 (m, 12H, CH=), 7.23 (s, 1H, CH=), 7.32-7.35 (m, 1H, CH=), 7.40-7.44 (m, 1H, CH=). ¹³C NMR (100.6 MHz, C₆D₆): δ =20.1 (d, CH₃, ³J_{C-P} =6.1 Hz), 20.3 (d, CH₃, ³J_{C-P} =6.8 Hz), 21.7 (CH₃), 39.4 (d, CH₂, ³J_{C-P} =6.9 Hz), 58.0 (d, CH-S, ³J_{C-P} =6.9 Hz), 85.7 (d, CH-OP, ³J_{C-P} =5.4 Hz), 124.9-143.5 (aromatic carbons).

4.5. Typical procedure for the allylic alkylation of disubstituted linear (S1 and S3-S7) and cyclic (S2 and S8-S9) substrates

A degassed solution of $[PdCl(\eta^3-C_3H_5)]_2$ (0.9 mg, 0.0025 mmol) and the corresponding ligand (0.0055 mmol) in dichloromethane (0.5 mL) was stirred for 30 min. Subsequently, a solution of the corresponding substrate (0.5 mmol) in dichloromethane (1.5 mL), nucleophile (1.5 mmol), *N*,*O*-bis(trimethylsilyl)-acetamide (370 µL, 1.5 mmol) and a pinch of KOAc were added. The reaction mixture was stirred at room temperature. After the desired reaction time the reaction mixture was extracted with Et₂O (5 mL) and saturated NH₄Cl (aq) (25 mL) was added. The mixture was extracted with Et₂O (3 x 10 mL) and the extract dried over MgSO₄. Conversions were measured by ¹H NMR and enantiomeric excesses were determined either by HPLC (compounds **11**, **13-22**, **37-41** and **43-46**) or by GC (compounds **12**, **47-48** and **50-52**) or by ¹H NMR using [Eu(hfc)₃] (compounds **42** and **49**). For characterization and ee determination details see Supporting Information.

4.6. Typical procedure for the allylic alkylation of disubstituted linear substrate S1 using pyrroles

A degassed solution of $[PdCl(\eta^3-C_3H_5)]_2$ (1.8 mg, 0.005 mmol) and the corresponding phosphite/phosphinite-thioether (0.011 mmol) in dichloromethane (0.5 mL) was stirred for 30 min. Subsequently, a solution of the corresponding substrate (0.5 mmol) in dichloromethane (1.5 mL), the corresponding pyrrole (0.4 mmol) and K₂CO₃ (110 mg, 0.8 mmol) were added. The reaction mixture was stirred at room temperature. After 18 h, the reaction mixture was diluted with Et₂O (5 mL) and saturated NH₄Cl (aq) (25 mL) was added. The mixture was extracted with Et₂O (3 x 10 mL) and the extract dried over MgSO₄. Conversions were measured by ¹H NMR and enantiomeric excesses were determined by HPLC. For characterization and ee determination details see Supporting Information.

4.7. Typical procedure for the allylic amination of disubstituted linear substrate S1

A degassed solution of $[PdCl(\eta^3-C_3H_5)]_2$ (0.9 mg, 0.0025 mmol) and the corresponding ligand (0.0055 mmol) in dichloromethane (0.5 mL) was stirred for 30 min. Subsequently, a solution of *rac*-1,3-diphenyl-3-acetoxyprop-1-ene (**S1**) (0.5 mmol) in dichloromethane (1.5 mL), the corresponding amine (1.5 mmol), *N*,*O*-bis(trimethylsilyl)-acetamide (370 µL, 1.5 mmol) and a pinch of KOAc were added. The reaction mixture was stirred at room temperature. After 2 hours, the reaction mixture was diluted with Et₂O (5 mL) and saturated NH₄Cl (aq) (25 mL) was added. The mixture was extracted with Et₂O (3 x 10 mL) and the extract dried over MgSO₄. Conversions were measured by ¹H NMR and enantiomeric excesses were determined by HPLC. For characterization and ee determination details see Supporting Information.

4.8. Typical procedure for the allylic etherification and silylation of disubstituted linear substrate S1

A degassed solution of $[PdCl(\eta^3-C_3H_5)]_2$ (0.9 mg, 0.0025 mmol) and the corresponding ligand (0.0055 mmol) in dichloromethane (0.5 mL) was stirred for 30 min. Subsequently, a solution of *rac*-1,3-diphenyl-3-acetoxyprop-1-ene (**S1**) (31.5 mg, 0.125 mmol) in dichloromethane (1.5 mL) was added. After 10 minutes, Cs₂CO₃ (122 mg, 0.375 mmol) and the corresponding alcohol or silanol (0.375 mmol) were added. The reaction mixture was stirred at room temperature. After 18 h, the reaction mixture was diluted with Et₂O (5 mL) and saturated NH₄Cl (aq) (25 mL) was added. The mixture was extracted with Et₂O (3 x 10 mL) and the extract dried over MgSO₄. Conversions were measured by ¹H NMR and enantiomeric excesses were determined by HPLC. For characterization and ee determination details see Supporting Information.

4.9. Typical procedure for the preparation of chiral carbo- and heterocyclic compounds 53-58

A solution of Grubbs II catalyst (5 mg, 0.006 mmol) and the corresponding alkylated product (0.12 mmol) in CH_2Cl_2 (3 mL) was stirred for 16 h. The solution was directly purified by flash chromatography (Hex/EtOAc 95:5) to obtained the desired compounds. For characterization and ee determination details see Supporting Information.

4.10. Typical procedure for the preparation of chiral tricyclic compounds 59-61

A solution of the starting enyne (0.187 mmol) in 1 mL of *tert*-butylbenzene was added to a solution of $Co_2(CO)_8$ (83 mg, 0.243 mmmol) in 0.5 mL of *tert*-butylbenzene under air. The flask was rinsed with 0.5 mL more of the same solvent. The resulting mixture was stirred at room temperature for 1 h, until full consumption of the starting material was observed by TLC. After that, the system was heated at 170 °C for a further hour. Then, it was cooled to room temperature, filtered on Celite with CH₂Cl₂ and concentrated in vacuo. The crude mixture was purified by flash column chromatography on silica gel eluting with cyclohexane/EtOAc (gradient form 90:10 to 70:30) to furnish the desired tricyclic compound as a white solid. For characterization and ee determination details see Supporting Information.

4.11. General procedure for the preparation of [Pd(η^3 -allyl)(L)]BF₄ complexes 62-65

The corresponding ligand (0.05 mmol) and the complex $[Pd(\mu-Cl)(\eta^3-1,3-allyl)]_2$ (0.025 mmol) were dissolved in CD₂Cl₂ (1.5 mL) at room temperature under argon. AgBF₄ (9.8 mg, 0.05 mmol) was added after 30 minutes and the mixture was stirred for 30 minutes. The mixture was then filtered over celite under argon and the resulting solutions were analyzed by NMR. After the NMR analysis, the complexes were precipitated as pale yellow solids by adding hexane.

[Pd(η³-1,3-cyclohexenylallyl)(L5b)]**BF**₄ (62). ³¹P NMR (CD₂Cl₂, 298 K), δ: 106.7 (s, 1P). ¹H NMR(CD₂Cl₂, 298 K), δ: 0.78 (m, 1H, CH₂, allyl), 1.03 (m, 1H, CH₂, allyl), 1.39 (s, 9H, CH₃, 'Bu), 1.42-1.56 (m, 2H, CH₂, allyl), 1.52 (s, 9H, CH₃, 'Bu), 1.62-1.71 (m, 2H, CH₂, allyl), 1.81 (s, 3H, CH₃), 1.97 (s, 3H, CH₃), 2.23 (s, 3H, CH₃), 2.41 (s, 3H, CH₃), 2.68 (s, 3H, CH₃, SR group), 2.91 (s, 3H, CH₃, SR group), 3.23 (dd, 1H, CH₂, ${}^{2}J_{H-H}$ = 12.4 Hz, ${}^{3}J_{H-H}$ = 7.2 Hz), 3.51 (dd, 1H, CH₂, ${}^{2}J_{H-H}$ = 12.4 Hz, ${}^{3}J_{H-H}$ = 6.4 Hz), 4.00 (m, 1H, CH allyl *trans* to S), 4.81 (m, 1H, CH-O), 5.15 (d, 1H, CH-S, ${}^{3}J_{H-H}$ = 6.8 Hz), 5.24 (m, 1H, CH allyl central), 5.46 (m, 1H, CH allyl *trans* to P), 6.71 (d, 1H, CH=, ${}^{3}J_{H-H}$ = 6 Hz), 7.18 (m, 1H, CH=), 7.3-7.5 (m, 7H, CH=). 13 C NMR (CD₂Cl₂, 298 K), δ : 18.1 (CH₃), 18.2 (CH₃), 20.9 (CH₂ allyl), 21.9 (CH₃), 22.0 (CH₃), 24.5 (CH₃, SR group), 25.7 (CH₃, SR group), 28.6 (CH₂ allyl), 29.6 (CH₂ allyl), 32.9 (CH₃, 'Bu), 33.5 (CH₃, 'Bu), 36.6 (C, 'Bu), 36.9 (C, 'Bu), 39.4 (d, CH₂, J_{C-P} = 6 Hz), 55.2 (CH-S), 83.6 (d, CH-O, J_{C-P} = 4.8 Hz), 84.3 (d, CH allyl *trans* to S, J_{C-P} = 6.5 Hz), 100.6 (d, CH allyl *trans* to P, J_{C-P} = 30.5 Hz), 115.0 (d, CH allyl central, J_{C-P} = 8.7 Hz), 125.5-147.3 (aromatic carbons). MS HR-ESI [found 839.2869, C₄₇H₅₈O₃PPdS (M-BF₄)⁺ requires 839.2874].

[Pd(η³-1,3-cyclohexenylallyl)(L5c)]BF₄ (63). ³¹P NMR (CD₂Cl₂, 298 K), δ: 106.2 (s, 1P). ¹H NMR(CD₂Cl₂, 298 K), δ: 1.18 (m, 1H, CH₂, allyl), 1.47 (s, 9H, CH₃, 'Bu), 1.55 (s, 9H, CH₃, 'Bu), 1.51-1.72 (m, 3H, CH₂, allyl), 1.84 (s, 3H, CH₃), 1.86 (m, 1H, CH₂, allyl), 2.14 (s, 3H, CH₃), 2.14 (m, 1H, CH₂), 2.38 (s, 3H, CH₃), 2.44 (s, 3H, CH₃), 2.58 (s, 3H, CH₃, SR group), 2.70 (s, 3H, CH₃, SR group), 3.29 (dd, 1H, CH₂, ²*J*_{H-H}= 12.4 Hz, ³*J*_{H-H}= 6.8 Hz), 3.56 (dd, 1H, CH₂, ²*J*_{H-H}= 12.4 Hz, ³*J*_{H-H}= 6 Hz), 4.08 (m, 1H, CH allyl *trans* to S), 5.20 (m, 1H, CH allyl *trans* to P), 5.28 (m, 2H, CH-S and CH allyl central), 5.34 (m, 1H, CH-O), 6.14 (d, 1H, CH₌, ³*J*_{H-H}= 6.4 Hz), 7.04 (m, 1H, CH=), 7.3-7.5 (m, 7H, CH=). ¹³C NMR (CD₂Cl₂, 298 K), δ: 18.1 (CH₃), 18.2 (CH₃), 21.0 (CH₂ allyl), 21.9 (CH₃), 22.0 (CH₃), 24.7 (CH₃, SR group), 24.8 (CH₃, SR group), 29.5 (CH₂ allyl), 30.96 (CH₂ allyl), 33.1 (CH₃, 'Bu), 33.5 (CH₃, 'Bu), 36.6 (C, 'Bu), 36.8 (C, 'Bu), 40.1 (d, CH₂, *J*_{C-P}= 5.4 Hz), 56.8 (d, CH-S, *J*_{C-P}= 3.2 Hz), 85.1 (m, CH-O and CH allyl *trans* to S), 103.8 (d, CH allyl *trans* to P, *J*_{C-P}= 28.6 Hz), 115.0 (d, CH allyl central, *J*_{C-P}= 7.9 Hz), 125.4-147.0 (aromatic carbons). MS HR-ESI [found 839.2870, C₄₇H₅₈O₃PPdS (M-BF₄)⁺ requires 839.2874].

[Pd(η^3 -1,3-diphenylallyl)(L5b)]BF4 (64). Major isomer (57%): ³¹P NMR (CD₂Cl₂, 298 K), δ : 102.8 (s, 1P). ¹H NMR(CD₂Cl₂, 298 K), δ : 1.25 (s, 9H, CH₃, ¹Bu), 1.59 (s, 9H, CH₃, ¹Bu), 1.66 (s, 3H, CH₃), 1.69 (s, 3H, CH₃), 1.97 (s, 3H, CH₃, SR group), 2.18 (s, 3H, CH₃), 2.24 (s, 3H, CH₃), 2.95 (s, 3H, CH₃, SR group), 2.97 (m, 1H, CH₂), 3.41 (dd, 1H, CH₂, ²*J*_{H-H}= 12.4 Hz, ³*J*_{H-H}= 7.2 Hz), 4.70 (m, 1H, CH-S), 4.75 (m, 1H, CH allyl *trans* to S), 4.85 (m, 1H,

CH-O), 5.46 (m, 1H, CH allyl *trans* to P), 6.18 (dd, 1H, CH allyl central, ${}^{3}J_{H-H}$ = 10.8 Hz, ${}^{3}J_{H-H}$ _H= 9.2 Hz), 6.36 (d, 1H, CH=, ${}^{3}J_{H-H}$ = 6.0 Hz), 6.71-7.52 (m, 14H, CH=). ${}^{13}C$ NMR (CD₂Cl₂, 298 K), δ: 16.9 (CH₃), 17.3 (CH₃), 20.7 (CH₃), 20.9 (CH₃), 22.5 (CH₃, SR group), 24.0 (CH₃, SR group), 31.5 (CH₃, ^tBu), 32.1 (CH₃, ^tBu), 36.4 (C, ^tBu), 36.6 (C, ^tBu), 37.6 (b, CH₂), 53.6 (CH-S), 80.2 (d, CH allyl trans to S, J_{C-P}= 7.3 Hz), 80.4 (d, CH-O, J_{C-P}= 6.8 Hz), 93.0 (d, CH allyl trans to P, J_{C-P}= 21.2 Hz), 110.8 (d, CH allyl trans to P, J_{C-P}= 8.4 Hz), 122.7-144.5 (aromatic carbons). Minor isomer (43%): ³¹P NMR (CD₂Cl₂, 298 K), δ: 102.6 (s, 1P). ¹H NMR(CD₂Cl₂, 298 K), δ: 1.50 (s, 9H, CH₃, ^tBu), 1.58 (s, 9H, CH₃, ^tBu), 1.63 (s, 3H, CH₃), 1.80 (s, 3H, CH₃), 2.20 (s, 3H, CH₃, SR group), 2.42 (s, 3H, CH₃), 2.48 (s, 3H, CH₃), 3.14 (s, 3H, CH₃, SR group), 2.97 (m, 1H, CH₂), 3.30 (dd, 1H, CH₂, ${}^{2}J_{H-H}$ = 12.4 Hz, ${}^{3}J_{H-H}$ = 7.2 Hz), 4.15 (m, 1H, CH allyl trans to S), 4.89 (m, 1H, CH-O), 5.02 (m, 1H, CH-S), 5.39 (m, 1H, CH allyl *trans* to P), 6.36 (d, 1H, CH=, ${}^{3}J_{H-H}$ = 6.0 Hz), 6.52 (t, 1H, CH allyl central, ${}^{3}J_{H-H}$ = 9.6 Hz), 6.71-7.52 (m, 14H, CH=). ¹³C NMR (CD₂Cl₂, 298 K), δ: 16.9 (CH₃), 17.3 (CH₃), 20.8 (CH₃), 20.8 (CH₃), 22.5 (CH₃, SR group), 23.3 (CH₃, SR group), 30.6 (CH₃, ^tBu), 31.0 (CH₃, ^tBu), 36.5 (C, ^tBu), 36.8C, ^tBu), 37.6 (b, CH₂), 53.5 (CH-S), 79.8 (d, CH allyl *trans* to S, J_{C-P}= 6.3 Hz), 92.3(d, CH-O, J_{C-P}= 6.2 Hz), 103.2 (d, CH allyl trans to P, J_{C-P}= 23 Hz), 112.1 (d, CH allyl trans to P, J_{C-P}= 10 Hz), 122.7-144.5 (aromatic carbons). MS HR-ESI [found 951.3184, C₅₆H₆₂O₃PPdS (M-BF₄)⁺ requires 951.3187].

[Pd(η³-1,3-diphenylallyl)(L5c)]BF4 (65). Major isomer (70%): ³¹P NMR (CD₂Cl₂, 298 K), δ: 104.3 (s, 1P). ¹H NMR(CD₂Cl₂, 298 K), δ: 1.41 (s, 9H, CH₃, ¹Bu), 1.63 (s, 3H, CH₃), 1.65 (s, 3H, CH₃), 1.77 (s, 12H, CH₃, ¹Bu and CH₃), 2.27 (s, 3H, CH₃), 2.47 (s, 3H, CH₃, SR group), 3.06 (s, 3H, CH₃, SR group), 3.01 (m, 1H, CH₂), 3.36 (dd, 1H, CH₂, ${}^{2}J_{H-H}$ = 12.8 Hz, ${}^{3}J_{H-H}$ = 6.8 Hz), 5.01 (m, 1H, CH-O), 5.06 (m, 1H, CH-S), 5.12 (m, 1H, CH allyl *trans* to S), 5.26 (d, 1H, CH allyl *trans* to P, ${}^{3}J_{H-H}$ = 10 Hz), 5.95 (d, 1H, CH=, ${}^{3}J_{H-H}$ = 6.0 Hz), 6.73 (m, 1H, CH allyl central), 6.87-7.51 (m, 14H, CH=). ¹³C NMR (CD₂Cl₂, 298 K), δ: 18.0 (CH₃), 18.4 (CH₃), 21.9 (CH₃), 22.1 (CH₃), 24.4 (CH₃, SR group), 25.8 (CH₃, SR group), 33.8 (CH₃, ¹Bu), 34.2 (CH₃, ¹Bu), 36.7 (C, ¹Bu), 36.9 (C, ¹Bu), 40.0 (d, CH₂, *J*_{C-P}= 6.7 Hz), 56.9 (d, CH-S, *J*_{C-P}= 2.7 Hz), 84.7 (d, CH-O, *J*_{C-P}= 5.4 Hz), 88.6 (d, CH allyl *trans* to S, *J*_{C-P}= 8.3 Hz), 124.7-146.9 (aromatic carbons). Minor isomer (30%): ³¹P NMR (CD₂Cl₂, 298 K), δ: 107.1 (s, 1P). ¹H NMR(CD₂Cl₂, 298 K), δ: 1.64 (s, 9H, CH₃, ¹Bu), 1.72 (s, 9H, CH₃, ¹Bu), 1.74 (s, 3H, CH₃), 1.82 (s, 3H, CH₃), 2.31 (s, 3H, CH₃), 2.33 (s, 3H, CH₃), 2.38 (s, 3H, CH₃, SR group), 2.60 (s,

3H, CH₃, SR group), 3.01 (m, 1H, CH₂), 3.47 (dd, 1H, CH₂, ${}^{2}J_{H-H}$ = 12.8 Hz, ${}^{3}J_{H-H}$ = 6.8 Hz), 4.85 (d, 1H, CH-S, ${}^{3}J_{H-H}$ = 5.6 Hz), 5.21 (m, 1H, CH allyl *trans* to S), 5.51 (m, 1H, CH-O), 5.70 (d, 1H, CH allyl *trans* to P, ${}^{3}J_{H-H}$ = 10 Hz), 6.00 (d, 1H, CH=, ${}^{3}J_{H-H}$ = 6.0 Hz), 6.58 (m, 1H, CH allyl central), 6.87-7.51 (m, 14H, CH=). 13 C NMR (CD₂Cl₂, 298 K), δ : 18.2 (CH₃), 18.3 (CH₃), 21.9 (CH₃), 22.0 (CH₃), 24.2 (CH₃, SR group), 26.1 (CH₃, SR group), 32.8 (CH₃, 'Bu), 34.1 (CH₃, 'Bu), 36.6 (C, 'Bu), 36.7 (C, 'Bu), 39.6 (d, CH₂, J_{C-P} = 7.5 Hz), 58.1 (d, CH-S, J_{C-P} = 2.5 Hz), 86.7 (d, CH-O, J_{C-P} = 7.5 Hz), 95.0 (d, CH allyl *trans* to S, J_{C-P} = 5.4 Hz), 95.9 (d, CH allyl *trans* to P, J_{C-P} = 23.7 Hz), 113.0 (d, CH allyl *trans* to P, J_{C-P} = 9.1 Hz), 124.7-146.9 (aromatic carbons). MS HR-ESI [found 951.3182, C₅₆H₆₂O₃PPdS (M-BF₄)⁺ requires 951.3187].

4.12. Study of the reactivity of the [Pd(η^3 -allyl)(L))]BF₄ with sodium dimethyl malonate by in situ NMR²⁹

A solution of *in situ* prepared $[Pd(\eta^3-allyl)(L)]BF_4$ (L= phosphite-thioether, 0.05 mmol) in CD_2Cl_2 (1 mL) was cooled in the NMR at -80 °C. At this temperature, a solution of cooled sodium dimethyl malonate (0.1 mmol) was added. The reaction was then followed by ³¹P NMR. The relative reaction rates were calculated using a capillary containing a solution of triphenylphosphine in CD_2Cl_2 as external standard.

4.13. Computational details

Geometries of all transition states and intermediates were optimized using the Gaussian 09 program,³⁰ employing the B3LYP³¹ density functional and the LANL2DZ³² basis set for palladium and the 6-31G* basis set for all other elements.³³ Solvation correction was applied in the course of the optimizations using the PCM model with the default parameters for dichloromethane.³⁴ The complexes were treated with charge +1 and in the single state. No symmetry constraints were applied. Normal mode analysis of all transition states revealed a single imaginary mode corresponding to the expected nucleophilic attack of the nucleophile to one of the two allylic termini carbons. The energies were further refined by performing single point calculations using the above-mentioned parameters, with the exception that the 6-311+G**³⁵ basis set was used for all elements except palladium, and by applying dispersion correction using DFT-D3³⁶ model. All energies reported are Gibbs free energies at 298.15 K and calculated as G_{reported} = G_{6-31G*} + (E_{6-311+G***} - E_{6-31G*}) + E_{DFT-D3}.

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