

## Precision medicine and lymphoma

James A Heward\*, Emil A Kumar, Koorosh Korfi, Jessica Okosun and Jude Fitzgibbon

Author Affiliations: Centre for Haemato-Oncology, Barts Cancer Institute, Queen Mary University of London, Charterhouse Square, London, EC1M 6BQ, United Kingdom.

\* **Corresponding Author:** James Heward, Centre for Haemato-Oncology, Barts Cancer Institute, Queen Mary University of London, Charterhouse Square, London, EC1M 6BQ, United Kingdom; Tel: +44 (0)20 7882 8780; Email: [j.a.heward@qmul.ac.uk](mailto:j.a.heward@qmul.ac.uk).

### Financial support and sponsorship

Our research is supported by Cancer Research UK (15968 awarded to JF, 22742 awarded to JO) and Bloodwise program grant [15002] through the Precision Medicine for Aggressive Lymphoma (PMAL) consortium. EAK is in receipt of fellowship funding from The Medical College of Saint Bartholomew's Hospital Trust.

## **Abstract**

### ***Purpose of review***

The treatment of the germinal center (GC) lymphomas, diffuse large B-cell (DLBCL) and follicular lymphoma (FL), has changed little beyond the introduction of immuno-chemotherapies. However, there exists a substantial group of patients within both diseases, for which improvements in care will involve appropriate tailoring of treatment.

### ***Recent findings***

DLBCL consists of two major subtypes with striking differences in their clinical outcomes paralleling their underlying genetic heterogeneity. Recent studies have seen advances in the stratification of GC lymphomas, through comprehensive profiling of 1001 DLBCLs alongside refinements in the identification of high-risk FL patients using m7-FLIPI and 23G models. A new wave of novel therapeutic agents is now undergoing clinical trials for GC lymphomas, with BCR and EZH2 inhibitors demonstrating preferential benefit in subgroups of patients. The emergence of cell-free DNA has raised the possibility of dynamic disease monitoring to potentially mitigate the complexity of spatial and temporal heterogeneity, whilst predicting tumor evolution in real-time.

### ***Summary***

Altogether knowledge of the genomic landscape of GC lymphomas is offering welcome opportunities in patient risk stratification and therapeutics. The challenge ahead is to establish how best to combine upfront or dynamic prognostication with precision therapies, whilst retaining practicality in clinical trials and the real-world setting.

***Key words***

Follicular lymphoma, diffuse large B-cell lymphoma, precision medicine, targeted therapies.

## **Introduction**

The treatment of the two most common forms of germinal center (GC) non-Hodgkin lymphomas (NHL), diffuse large B-cell (DLBCL) and Follicular lymphoma (FL), was revolutionized by the addition of rituximab to chemotherapeutic regimens (R-CHOP). Over half of DLBCL patients are now curable by R-CHOP, whilst despite FL being considered an incurable disease, the majority of FL patients live for upwards of 15 years. Beyond these successes, prognosis remains poor for a third of relapsed-refractory DLBCL patients, and for a quarter of FL patients who are prone to early relapse and/or transformation to a more aggressive lymphoma (tFL). There is renewed optimism that the molecular characterization of these diseases will allow for the prediction of high-risk patients and the tailoring of treatments based on a patient's molecular profile. In this review, we will focus on the prognostic strategies to identify high-risk patients and the progress now being made in developing targeted therapies for these GC lymphomas.

### **The molecular landscape of GC lymphomas.**

The introduction of next generation sequencing (NGS) sparked a dramatic increase in our understanding of the molecular events that drive GC lymphomas. There are now well over 200 recurring gene mutations described, although only a minority occur in greater than 5-10% of patients. The significance of gene translocations targeting *MYC*, *BCL2*, and *BCL6* are all well established, as is the recognition of widespread disruption of the epigenome, predominantly driven by somatic mutations within *KMT2D*, *CREBBP*, *EZH2* and linker histones [1-13]. DLBCL displays a greater degree of genetic heterogeneity than FL, and can be readily divided into at least two major

subtypes based on the “cell of origin” (COO): GC B cell (GCB)-like and activated B cell (ABC)-like DLBCL [14,15]. The ABC-DLBCL subtype has the poorer prognosis by far (40% 3-year overall survival (OS)) and is typified by constitutive NF-κB activation and signaling driven by somatic mutations in the B cell receptor (BCR) and NF-κB pathway genes (*CARD11*, *CD79B*, *MYD88*, and *TNFAIP3*) [16-20], whereas GCB-DLBCL has a better prognosis (80% 3-year OS) and more closely resembles FL. While there are specific gene mutations that are enriched in each group, the overall landscape is complex and contrary to earlier expectations, BCR and NF-κB pathways mutations also arise in GCB-DLBCL and FL, albeit at a lower frequency [2,11].

### **Identifying high-risk DLBCL patients**

Overall, progress has been sluggish in discriminating high- from low-risk patients within the distinct COO entities with the exception of the “high-grade B cell lymphomas, with *MYC* and *BCL2* and/or *BCL6* rearrangements”, also known as double- and triple-hit lymphomas, and the “double-expressors” that highly express both *BCL2* and *MYC* in the absence of rearrangements [21,22]. Both groups demonstrate an aggressive phenotype associated with poor outcomes and therefore are a high priority for clinical intervention. Nevertheless, we are likely to see an acceleration in improved prognostic models and refined patient subsets in the foreseeable future, as our understanding of the landscape of coding aberrations in DLBCL nears completion. A monumental study by Reddy and colleagues has set the pace with an extensive analysis of 1001 newly diagnosed DLBCL patients, integrating mutational profiling, gene expression analysis and clinical information, which has offered novel insights into combinatorial factors influencing outcome for near

enough the first time. While many of these observations will require validation, (for example *KLH14* mutations being predictive of high-risk ABC-DLBCL, and *EZH2* mutations predicting low-risk GCB-DLBCL), the size and depth of the data allowed the authors to devise an encouraging prognostic multivariate genomic risk model [11]. Such large collaborative studies will undoubtedly be the benchmark for future studies and provides a framework for developing prospective risk-adapted strategies for DLBCL.

### **A new era in FL stratification?**

Molecular stratification of FL has lagged behind DLBCL, with the COO classification first described nearly 20 years ago [14]. Traditional attempts at risk stratification in the era of immunochemotherapy have been based on clinical parameters such as the Follicular Lymphoma International Prognostic Index (FLIPI) scores, the FLIPI and FLIPI-2, but are for the most part fairly ineffective tools for directing management decisions [23,24]. While molecular prognostication has long been opined as having more potential, it is only of late that we are seeing inroads into the development of tools capable of navigating the clinical heterogeneity within FL and able to predict high-risk patients. The earliest studies by the Lymphoma/Leukemia Molecular Profiling Project stratified FL based on the patterns of two gene signatures associated with T cell and myeloid cell infiltration [25]. However, we may well have turned the corner with the recent development of the m7-FLIPI, a clinico-genetic risk model that incorporates the clinical features of the FLIPI and the mutational status of 7 genes (*EP300*, *FOXO1*, *CREBBP*, *CARD11*, *MEF2B*, *ARID1A*, *EZH2*) [26], and an alternative prognostic strategy based on the expression of 23 genes (23G model)

from both the tumor and microenvironment [27]. It is likely that these models will continue to evolve as they are impacted by treatment changes, such as the move towards chemotherapy-free interventions, and we need to lay the groundwork for these changes by developing the infrastructure to enable head-to-head comparisons and reach a consensus on the level of discrimination needed to guide treatment, especially for an indolent disease such as FL where a high prognostic accuracy is necessary.

### **Upfront prognostication verses dynamic disease monitoring**

All of the aforementioned clinical and molecular prognostic strategies are based on upfront testing and prognostication. However, sampling of a single site at diagnosis fails to account for the temporal [1,28-31] and spatial heterogeneity [32] observed in GC lymphomas. The assessment of cell-free DNA (cfDNA) from plasma samples offers the possibility of more effectively sampling the heterogeneity in GC lymphomas, and dynamically monitoring changes in prognostic markers and actionable targets in a minimally invasive manner. Whilst it has not been widely applied so far, early studies have suggested a number of possible applications including the ability to classify the COO in DLBCL patients and to predict transformation of FL [33-35]. In an era where there is a headlong shift away from excision biopsies, it is likely that non-invasive cfDNA-based diagnostics and prognostics will have a significant future role within GC lymphomas.

### **Ibrutinib preferentially benefits ABC-DLBCL patients**

Running in tandem with efforts at exploiting genetic data for defining risk, access to genetic profiles are also pointing the way towards novel therapies. Ibrutinib, which inhibits the BTK enzyme responsible for propagating pro-survival signals from the BCR, which is constitutively active in ABC-DLBCLs (Figure 1) [36], has proved to be revolutionary in the treatment of chronic lymphocytic leukemia [37], and accumulating data suggest that it may be efficacious in poor-risk, BCR/NF- $\kappa$ B-dependent ABC-DLBCL patients. In a Phase I/II trial of 80 patients, single agent ibrutinib was well tolerated and had an overall response rate (ORR) of 37% in ABC-DLBCL, compared to only 5% in GCB-DLBCL patients [36], and it is encouraging that all non-GCB patients achieved complete remission in a Phase I study examining ibrutinib in combination with rituximab-based chemotherapy [38]. Following the success of these trials, ibrutinib is being tested in several phase III trials for ABC-DLBCL, that includes combination with R-CHOP for newly diagnosed ABC-DLBCL patients (NCT01855750), and in relapsed/refractory ABC-DLBCL patients undergoing stem cell transplant (NCT02443077).

In FL, single agent ibrutinib was shown to only have modest activity [39], despite evidence that BCR signaling has a role in FL pathogenesis [40-44]. Combination studies with rituximab have improved on the single-agent outcomes [45] but it seems sensible that upfront selection should be a feature of future trials, with evidence that *MYD88* mutations are associated with a positive outcome in DLBCL [36], while *CARD11* mutations appear to influence ibrutinib resistance in FL [39].

**EZH2 inhibitors may selectively benefit *EZH2*-mutant patients**



The recent recognition of gain-of-function mutations in *EZH2* demonstrates the speed by which discoveries relating to the molecular pathogenesis of a disease are translatable into potential clinical benefit. EZH2 is a histone methyltransferase that acts as the catalytic subunit of the polycomb repressor complex 2 and catalyzes repressive mono-, di- and tri-methylation of the histone 3 lysine 27 (H3K27) residue. In GC lymphomas, its important role in regulating the normal GC reaction [46] is subverted by heterozygous mutations most commonly altering tyrosine 646 (Y646) within the catalytic SET domain [47]. These gain-of-function mutations alter the catalytic activity of the mutant EZH2 enzyme so that it preferentially catalyzes the conversion of H3K27me1 into the strongly repressive H3K27me2/3 marks, whilst the wild-type protein continues to deposit H3K27me1 [48].

Selective EZH2 inhibitors have been developed by Epizyme (EPZ6438, tazemetostat [49]), GlaxoSmithKline (GSK126 [50]) and Constellation Pharmaceuticals (CPI-1205 [51]), with pre-clinical data indicating that these compounds are generally more active in mutant cell lines and able to re-activate genes repressed or silenced by mutant EZH2 [49-51]. Phase I/II clinical trials have now been launched for all of these compounds to examine their efficacy for NHL with recent interim data suggesting that tazemetostat is efficacious for *EZH2*-mutated FL patients (92% ORR in mutant vs 26% in wild-type) and to a lesser extent in DLBCL (29% ORR in mutant vs 15% in wild-type) [52]. Altogether, these results are encouraging and highlight the attractions of precision medicine although given that *EZH2* mutations have been provisionally linked with better outcomes in GC lymphomas [26,53], EZH2 inhibitors will need to

demonstrate significant benefit over the current standard of care to justify their use as a targeted therapy in this sub-group of patients.

### **Revisiting mTOR inhibitors in the light of recent developments**

It is also possible that armed with new insights into the genetic basis of GC lymphomas, we can revisit historical clinical studies and explore opportunities for precision treatments. Inhibition of the mTOR (mammalian Target Of Rapamycin) pathway, previously evaluated in FL, serves as a notable example with two phase II trials of second generation mTOR inhibitors, temsirolimus and everolimus, demonstrating promising results in multiply relapsed cohorts (ORR 53.8% [54] and ORR 38% [55], respectively). These trials however were performed before the discovery of unique mutations of the *RRAGC* and the V-ATPase *ATP6V1B2* and *ATP6AP1* genes in around 30% of FL patients, enforcing mTORC1 activation [4,,56,57]. With the spectrum of different molecular lesions that we are now aware of in GC lymphomas, there may be opportunities to re-examine previously trialed agents, to determine whether events such as mTOR pathway mutations are predictive of response. Equally this also reaffirms the need and value of rigorously collecting biopsy material as part of clinical trials for later correlative studies.

### **New strategies to reverse the early loss of CREBBP in precursor cells**

Given the frequency and recognition of *CREBBP* mutations as one of the earliest events in GC lymphomas [8-11, 58-60], and their widespread role in promoting GC-lymphoma development [4,58-62], the successful targeting of these lesions would offer an exciting new therapy with the potential to eradicate disease-propagating

cells. Indeed, phylogenetic analysis has revealed that overt FL, and subsequent relapses of the disease, are likely to develop from long-lived pre-malignant cells known as common progenitor cells (CPCs), which are believed to be t(14;18)-positive and typically contain mutations within the histone regulatory genes *CREBBP* and *KMT2D* [1,3-5,63-66]. There is a persuasive argument to suppose that inhibition of the histone deacetylase (HDAC) enzymes that normally oppose CREBBP by removing acetylation marks could mitigate the deep-rooted loss of histone acetyltransferases CREBBP and EP300. Overall, whilst pan-HDAC inhibitors have demonstrated efficacy in GC lymphomas [67-69], the occurrence of significant toxicities, alongside the absence of a biomarker and an unclear mechanism of action, have limited progress beyond phase II trials. Their fortunes may well change in the future, with recent studies exploring the relationship between *CREBBP* mutations and HDAC isoforms indicating that HDAC3 opposes the activity of CREBBP at enhancers and hyper-represses these enhancers following the loss of *CREBBP*, resulting in an increased dependency on HDAC3 for survival [58]. Targeting the HDAC3 isoform thus offers a potential therapeutic strategy with the promise of re-activating CREBBP-regulated genes whilst minimizing toxicity associated with pan-HDAC inhibitors.

## **Conclusion**

Our understanding of the biology of the GC lymphomas has increased dramatically with the introduction of NGS, and our ability to parallel clinical and molecular heterogeneity. There are signs that we are on the edge of a new precision era for GC lymphomas. Several new upfront prognostic strategies have been published for FL, and the seminal study by Reddy et al examining 1001 DLBCL patients has highlighted

the need for similar large scale, multi-institutional collaborative studies across all lymphomas, including FL. Furthermore, cfDNA assessment have raised the possibility of undertaking dynamic disease monitoring accounting for spatial and temporal heterogeneity and forecasting the trajectory of the tumor's evolution in real-time.

The identification of therapeutics targeted towards specific molecular lesions has also seen significant advances in GC lymphomas, including the observations of selective benefit for ibrutinib in ABC-DLBCL and tazemetostat in EZH2 mutant FL. The challenge now is to establish how best to combine upfront prognostication with dynamic monitoring, whilst retaining practicality, to ensure that patients receive the best possible treatment.

### **Key points**

- Specific sub-groups of high-risk DLBCL patients within the COO entities are now recognized.
- Notable advances made in identifying high-risk FL with the development of novel molecular prognostication tools.
- Ibrutinib and tazemetostat are showing preferential benefit in specific populations of GC lymphoma patients.

### **Acknowledgements**

None.

### **Financial support and sponsorship**

Our research is supported by Cancer Research UK (15968 awarded to JF, 22742 awarded to JO) and Bloodwise program grant [15002] through the Precision Medicine for Aggressive Lymphoma (PMAL) consortium. EAK is in receipt of fellowship funding from The Medical College of Saint Bartholomew's Hospital Trust.

### **Conflicts of interest**

JF declares grants from Epizyme and personal fees from Roche, Gilead, Janssen, and Epizyme.

### **Figure legend**

**Figure 1. Actionable mutations in GC lymphomas.** Diagram indicating key recurring and potentially targetable mutations. Highlighted therapeutics include ibrutinib for ABC-DLBCL [36] and EZH2 inhibitors for *EZH2*-mutant FL patients [49-52].

## References

1. Okosun J, Bödör C, Wang J, Araf S, Yang C-Y, Pan C, et al. Integrated genomic analysis identifies recurrent mutations and evolution patterns driving the initiation and progression of follicular lymphoma. *Nat Genet.* 2013; 46:176–181.
  2. Krysiak K, Gomez F, White BS, Matlock M, Miller CA, Trani L, et al. Recurrent somatic mutations affecting B-cell receptor signaling pathway genes in follicular lymphoma. *Blood* 2017; 129:473–483.
- \* This study described the occurrence of low-frequency BCR mutations in follicular lymphoma
3. Pasqualucci L, Khiabani H, Fangazio M, Vasishtha M, Messina M, Holmes AB, et al. Genetics of follicular lymphoma transformation. *Cell Rep.* 2014; 6:130–40.
  4. Green MR, Kihira S, Liu CL, Nair RV, Salari R, Gentles AJ, et al. Mutations in early follicular lymphoma progenitors are associated with suppressed antigen presentation. *Proc Natl Acad Sci* 2015; 112:1116–1125.
  5. Korfi K, Ali S, Heward JA, Fitzgibbon J. Follicular lymphoma, a B cell malignancy addicted to epigenetic mutations. *Epigenetics* 2017; 12:370–377.
  6. Li H, Kaminski MS, Li Y, Yildiz M, Ouillette P, Jones S, et al. Mutations in linker histone genes HIST1H1 B, C, D, and E; OCT2 (POU2F2); IRF8; and ARID1A underlying the pathogenesis of follicular lymphoma. *Blood* 2014; 123:1487–1498.
  7. Araf S, Okosun J, Koniali L, Fitzgibbon J, Heward J. Epigenetic dysregulation in follicular lymphoma. *Epigenomics* 2015; 8:77–84.
  8. Genetic heterogeneity of diffuse large B-cell lymphoma. *Proc Natl Acad Sci* 2013; 110:1398–1403.
  9. Frequent mutation of histone-modifying genes in non-Hodgkin lymphoma. *Nature* 2011; 476:298–303.
  10. Testoni M, Zucca E, Young KH, Bertoni F. Genetic lesions in diffuse large B-cell lymphomas. *Ann Oncol* 2015; 26:1069–1080.
  11. Reddy A, Waldrop A, Leppä S, Pasanen A, Meriranta L, Karjalainen-Lindsberg M-L, et al. Genetic and Functional Drivers of Diffuse Large B Cell Lymphoma. *Cell* 2017;171: 481–494.
- \*\* This seminal publication performed an integrative analysis of 1001 DLBCL patients
12. Bödör C, Grossmann V, Popov N, Okosun J, O'Riain C, Tan K, et al. EZH2

mutations are frequent and represent an early event in follicular lymphoma. *Blood* 2013; 122:3165–3168.

13. Boice M, Salloum D, Mourcin F, Sanghvi V, Cell RA, 2016. Loss of the HVEM tumor suppressor in lymphoma and restoration by modified CAR-T cells. *Cell* 2016; 167:405-418.
14. Alizadeh AA, Eisen MB, Davis RE, Ma C, Lossos IS, Rosenwald A, et al. Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. *Nature* 2000; 403:503–511.
15. Hans CP, Weisenburger DD, Greiner TC, Gascoyne RD, Delabie J, Ott G, et al. Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a tissue microarray. *Blood* 2004; 103: 275–282.
16. Lenz G, Wright GW, Emre NCT, Kohlhammer H, Dave SS, Davis RE, et al. Molecular subtypes of diffuse large B-cell lymphoma arise by distinct genetic pathways. *Proc Natl Acad Sci* 2008; 105:13520–13525.
17. Lenz G, Davis RE, Ngo VN, Lam L, George TC, Wright GW, et al. Oncogenic CARD11 Mutations in Human Diffuse Large B Cell Lymphoma. *Science* 2008; 319:1676–1679.
18. Davis RE, Ngo VN, Lenz G, Tolar P, Young RM, Romesser PB, et al. Chronic active B-cell-receptor signalling in diffuse large B-cell lymphoma. *Nature* 2010; 463:88-92.
19. Ngo VN, Young RM, Schmitz R, Jhavar S, Xiao W, Lim K-H, et al. Oncogenically active MYD88 mutations in human lymphoma. *Nature* 2011; 470:115-120.
20. Compagno M, Lim WK, Grunn A, Nandula SV, Brahmachary M, Shen Q, et al. Mutations of multiple genes cause deregulation of NF-kappaB in diffuse large B-cell lymphoma. *Nature* 2009; 459:717–721.
21. Rosenthal A, reviews AYB, 2017. High grade B-cell lymphoma with rearrangements of MYC and BCL2 and/or BCL6: Double hit and triple hit lymphomas and double expressing lymphoma. *Blood Rev* 2016; 31: 37-42.
22. Swerdlow SH, Campo E, Pileri SA, Harris NL, Stein H, Siebert R, et al. The 2016 revision of the World Health Organization classification of lymphoid neoplasms. *Blood* 2016; 127:2375–2390.
23. Solal-Céligny P, Roy P, Colombat P, White J, Armitage JO, Arranz-Saez R, et al. Follicular Lymphoma International Prognostic Index. *Blood* 2004; 104:1258–1265.

24. Federico M, Bellei M, Marcheselli L, Luminari S, Lopez-Guillermo A, Vitolo U, et al. Follicular Lymphoma International Prognostic Index 2: A New Prognostic Index for Follicular Lymphoma Developed by the International Follicular Lymphoma Prognostic Factor Project. *J Clin Oncol* 2009; 27:4555–4562.
25. Dave SS, Wright G, Tan B, Rosenwald A, Gascoyne RD, Chan WC, et al. Prediction of survival in follicular lymphoma based on molecular features of tumor-infiltrating immune cells. *N Engl J Med* 2004; 351:2159–2169.
26. Pastore A, Jurinovic V, Kridel R, Hoster E, Staiger AM, Szczepanowski M, et al. Integration of gene mutations in risk prognostication for patients receiving first-line immunochemotherapy for follicular lymphoma: a retrospective analysis of a prospective clinical trial and validation in a population-based registry. *The Lancet Oncology* 2015; 16:1111–1122.
27. Huet S, Tesson B, Jais J-P, Feldman AL, Magnano L, Thomas E, et al. A gene-expression profiling score for prediction of outcome in patients with follicular lymphoma: a retrospective training and validation analysis in three international cohorts. *The Lancet Oncology* 2018; 19:549-561.

\*\* This study identified a 23 gene signature for stratifying high and low-risk FL patients

28. Kridel R, Chan FC, Mottok A, Boyle M, Farinha P, Tan K, et al. Histological Transformation and Progression in Follicular Lymphoma: A Clonal Evolution Study. *PLoS Med* 2016; 13:e1002197.
29. A Predictive Model for Aggressive Non-Hodgkin's Lymphoma. *N Engl J Med* 1993; 329:987–994.
30. Juskevicius D, Lorber T, Gsponer J, Perrina V, Ruiz C, Stenner-Liewen F, et al. Distinct genetic evolution patterns of relapsing diffuse large B-cell lymphoma revealed by genome-wide copy number aberration and targeted sequencing analysis. *Leukemia* 2016; 30:2385.
31. Jiang Y, Redmond D, Nie K, Eng KW, Clozel T, Martin P, et al. Deep sequencing reveals clonal evolution patterns and mutation events associated with relapse in B-cell lymphomas. *Genome biology* 2014; 15:432.
32. Araf S, Wang J, Korfi K, et al. Genomic profiling reveals spatial intra-tumor heterogeneity in follicular lymphoma. *Leukemia* 2018. doi:10.1038/s41375-018-0043y
33. Scherer F, Kurtz DM, Newman AM, Stehr H, Craig AFM, Esfahani MS, et al. Distinct biological subtypes and patterns of genome evolution in lymphoma revealed by circulating tumor DNA. *Sci Transl Med* 2016;



8:364ra155–5.

34. Roschewski M, Staudt LM, Wilson WH. Dynamic monitoring of circulating tumor DNA in non-Hodgkin lymphoma. *Blood* 2016; 127:3127–3132.
35. Rossi D, Diop F, Spaccarotella E, Monti S, Zanni M, Rasi S, et al. Diffuse large B-cell lymphoma genotyping on the liquid biopsy. *Blood* 2017; 129:1947–1957.
36. Wilson WH, Young RM, Schmitz R, Yang Y, Pittaluga S, Wright G, et al. Targeting B cell receptor signaling with ibrutinib in diffuse large B cell lymphoma. *Nat Med* 2015; 21:922–926.
37. Khan M, Gibbons JL, Ferrajoli A. Spotlight on ibrutinib and its potential in frontline treatment of chronic lymphocytic leukemia. *Onco Targets Ther.* 2017; 10:1909–1914.
38. Sauter CS, Matasar MJ, Schoder H, Devlin SM, Drullinsky P, Gerecitano J, et al. A Phase I Study of Ibrutinib in Combination with R-ICE in Patients with Relapsed or Primary Refractory DLBCL. *Blood* 2018.
39. Bartlett NL, Costello BA, LaPlant BR, Ansell SM, Kuruvilla JG, Reeder CB, et al. Single-Agent Ibrutinib in Relapsed or Refractory Follicular Lymphoma: A Phase 2 Consortium Trial. *Blood* 2017; 131:182-190.
40. Zhu D, McCarthy H, Ottensmeier CH, Johnson P, Hamblin TJ, Stevenson FK. Acquisition of potential N-glycosylation sites in the immunoglobulin variable region by somatic mutation is a distinctive feature of follicular lymphoma. *Blood* 2002; 99:2562–2568.
41. Amin R, Mourcin F, Uhel F, Pangault C, Ruminy P, Dupré L, et al. DC-SIGN–expressing macrophages trigger activation of mannosylated IgM B-cell receptor in follicular lymphoma. *Blood* 2015; 126:1911–1920.
42. Linley A, Krysov S, Ponzoni M, Johnson PW, Packham G, Stevenson FK. Lectin binding to surface Ig variable regions provides a universal persistent activating signal for follicular lymphoma cells. *Blood* 2015; 126:1902–1910.
43. Recurrent somatic mutations affecting B-cell receptor signaling pathway genes in follicular lymphoma. *Blood* 2016; 129:473–483.
44. Irish JM, Myklebust JH, Alizadeh AA, Houot R, Sharman JP, Czerwinski DK, et al. B-cell signaling networks reveal a negative prognostic human lymphoma cell subset that emerges during tumor progression. *Proc Natl Acad Sci* 2010; 107:12747–12754.
45. Fowler N, Nastoupil L, de Vos S, Knapp M, Flinn IW, Chen RW, et al.

Ibrutinib Combined with Rituximab in Treatment-Naive Patients with Follicular Lymphoma: Arm 1 + Arm 2 Results from a Multicenter, Open-Label Phase 2 Study. *Blood* 2016; 128:1804–1814.

46. De Silva NS, Klein U. Dynamics of B cells in germinal centres. *Nature reviews Immunology* 2015; 15:137-148.
47. Morin RD, Johnson NA, Severson TM, Mungall AJ, An J, Goya R, et al. Somatic mutations altering EZH2 (Tyr641) in follicular and diffuse large B-cell lymphomas of germinal-center origin. *Nat Genet* 2010; 42:181–185.
48. Sneeringer CJ, Scott MP, Kuntz KW, Knutson SK, Pollock RM, Richon VM, et al. Coordinated activities of wild-type plus mutant EZH2 drive tumor-associated hypertrimethylation of lysine 27 on histone H3 (H3K27) in human B-cell lymphomas. *Proc Natl Acad Sci* 2010; 107:20980–20985.
49. Knutson SK, Kawano S, Minoshima Y, Warholc NM, Huang K-C, Xiao Y, et al. Selective inhibition of EZH2 by EPZ-6438 leads to potent antitumor activity in EZH2-mutant non-Hodgkin lymphoma. *Mol Cancer Ther* 2014; 13:842–854.
50. McCabe MT, Ott HM, Ganji G, Korenchuk S, Thompson C, Van Aller GS, et al. EZH2 inhibition as a therapeutic strategy for lymphoma with EZH2-activating mutations. *Nature* 2012; 492:108–112.
51. Garapaty-Rao S, Nasveschuk C, Gagnon A, Chan EY, Sandy P, Busby J, et al. Identification of EZH2 and EZH1 Small Molecule Inhibitors with Selective Impact on Diffuse Large B Cell Lymphoma Cell Growth. *Chem Bio* 2013; 20:1329–1339.
52. Morschhauser F, Salles G, McKay P, Tilly H, schmitt A, Gerecitano J, et al. Interim report from a phase 2 multicenter study of tazemostat, an EZH2 inhibitor: clinical activity and favourable safety in patients with relapsed or refractory B-cell non-hodgkin lymphoma. *International Conference on Malignant Lymphoma*. 2017.
53. Huet S, Xerri L, Tesson B, Mareschal S, Taix S, Mescam-Mancini L, et al. EZH2 alterations in follicular lymphoma: biological and clinical correlations. *Blood Cancer J* 2017 7: e555–565.
54. Smith SM, van Besien K, Karrison T, Dancey J, McLaughlin P, Younes A, et al. Temsirolimus has activity in non-mantle cell non-Hodgkin's lymphoma subtypes: The University of Chicago phase II consortium. *J Clin Oncol* 2010; 28:4740–4746.
55. Witzig TE, Reeder CB, LaPlant BR, Gupta M, Johnston PB, Micallef IN, et al. A phase II trial of the oral mTOR inhibitor everolimus in relapsed aggressive lymphoma. *Leukemia* 2011; 25:341–347.

56. Okosun J, Wolfson RL, Wang J, Araf S, Wilkins L, Castellano BM, et al. Recurrent mTORC1-activating RRAGC mutations in follicular lymphoma. *Nat Genet* 2016; 48:183–188.
57. Ying ZX, Jin M, Peterson LF, Bernard D, Saiya-Cork K, Yildiz M, et al. Recurrent Mutations in the MTOR Regulator RRAGC in Follicular Lymphoma. *Clin Cancer Res.* 2016; 22:5383–5393.
58. Jiang Y, Ortega-Molina A, Geng H, Ying H-Y, Hatzi K, Parsa S, et al. CREBBP Inactivation Promotes the Development of HDAC3-Dependent Lymphomas. *Cancer Discov* 2017; 7:38–53.
59. García-Ramírez I, Tadros S, González-Herrero I, Martín-Lorenzo A, Rodríguez-Hernández G, Moore D, et al. Crebbp loss cooperates with Bcl2 overexpression to promote lymphoma in mice. *Blood* 2017; 129:2645–2656.
60. Hashwah H, Schmid CA, Kasser S, Bertram K, Stelling A, Manz MG, et al. Inactivation of CREBBP expands the germinal center B cell compartment, down-regulates MHCII expression and promotes DLBCL growth. *Proc Natl Acad Sci* 2017;114:9701–9706.
61. Zhang J, Vlasevska S, Wells VA, et al. The Crebbp Acetyltransferase is a Haploinsufficient Tumor Suppressor in B Cell Lymphoma. *Cancer Discov* 2017; 7:322-337.
62. Horton SJ, Giotopoulos G, Yun H, Vohra S, Sheppard O, Bashford-Rogers R, et al. Early loss of Crebbp confers malignant stem cell properties on lymphoid progenitors. *Nat Cell Biol* 2017; 19:1093–1104.
63. Green MR, Alizadeh AA. Chromatin modifying gene mutations in follicular lymphoma. *Blood* 2017; 121:1604-1611.
64. Green MR, Gentles AJ, Nair RV, Irish JM, Kihira S, Liu CL, et al. Hierarchy in somatic mutations arising during genomic evolution and progression of follicular lymphoma. *Blood* 2013; 121:1604–1611.
65. Carlotti E, Wrench D, Matthews J, Iqbal S, Davies A, Norton A, et al. Transformation of follicular lymphoma to diffuse large B-cell lymphoma may occur by divergent evolution from a common progenitor cell or by direct evolution from the follicular lymphoma clone. *Blood* 2009; 113:3553–3557.
66. Weigert O, Kopp N, Lane AA, Yoda A, Dahlberg SE, Neuberg D, et al. Molecular ontogeny of donor-derived follicular lymphomas occurring after hematopoietic cell transplantation. *Cancer Discov* 2012; 2:47–55.
67. Ogura M, Ando K, Suzuki T, Ishizawa K, Oh SY, Itoh K, et al. A multicentre phase II study of vorinostat in patients with relapsed or refractory

indolent B-cell non-Hodgkin lymphoma and mantle cell lymphoma. *Br J Haematol* 2014; 165:768–776.

68. Kirschbaum M, Frankel P, Popplewell L, Zain J, Delioukina M, Pullarkat V, et al. Phase II Study of Vorinostat for Treatment of Relapsed or Refractory Indolent Non-Hodgkin's Lymphoma and Mantle Cell Lymphoma. *J Clin Oncol* 2011; 9:1198-1203.
69. Assouline SE, Nielsen TH, Yu S, Alcaide M, Chong L, MacDonald D, et al. Phase 2 study of panobinostat +/- rituximab in relapsed diffuse large B cell lymphoma and biomarkers predictive of response. *Blood* 2016; 128:185-194.